

NEW ZEALAND
DEPARTMENT OF SCIENTIFIC AND INDUSTRIAL RESEARCH

BULLETIN 196

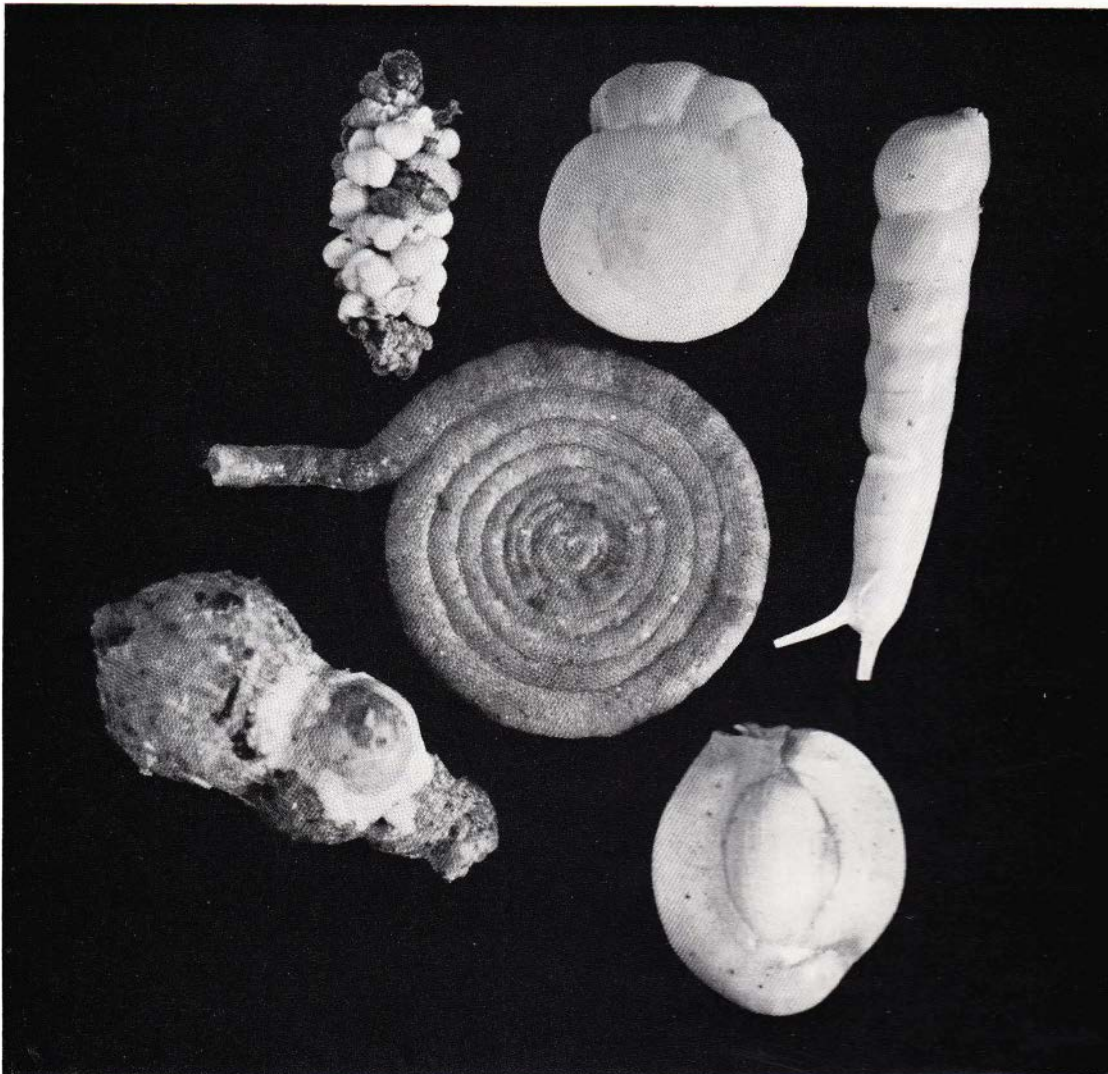
**A Key to the Recent Genera
of the
Foraminiferida**

by
K. B. Lewis

New Zealand Oceanographic Institute
Memoir No. 45

1970

A KEY TO THE RECENT GENERA
OF THE
FORAMINIFERIDA



Magnification $\times 16$ approx.

Photograph: J. Whalan

A selection of foraminifera from NZOI Sta. C 488, which was situated 30 km to the east of Cape Palliser, the most southerly point of the North Island, New Zealand. The sample, consisting of mud, was collected from a depth of 459 m.

The arenaceous species illustrate a wide variation in test form and the material selected for test construction, e.g.: UNILOCCULAR GLOBOSE (top left) *Brachysiphon corbuliformis* Chapman (composed of agglutinated glauconite grains and pelagic foraminifera only); UNILOCCULAR ENROLLED TUBULAR (centre) *Ammodiscus tenuis* Brady (composed of silt grains); UNISERIAL (bottom left) *Hormosina globulifera* Brady (composed of fine sand, mainly quartz); ATTACHED LOW TROCHOSPIRAL (bottom left, attached to *Hormosina*) *Tritaxis fusca* (Williamson) (composed of silt-sized grains).

The calcareous species are: QUINQUELOCULINE (bottom right) *Quinqueloculina* sp. (with an opaque white (parcellaneous) test); UNCOILING UNISERIAL (top right) *Vaginulopsis spinigera* (Brady) (with a translucent (hyaline) test and radiate aperture); LOW TROCHOSPIRAL (top centre) *Heterolepa subhardingerii* (Parr) (with a translucent (hyaline) final chamber and distinctly perforate wall).

NEW ZEALAND
DEPARTMENT OF SCIENTIFIC AND INDUSTRIAL RESEARCH

BULLETIN 196

**A Key to the Recent Genera
of the
Foraminiferida**

by
K. B. Lewis

**New Zealand Oceanographic Institute
Memoir No. 45**

January 1970

Price: \$2.50

This publication should be referred to as:
Bull. N.Z. Dep. scient. ind. Res. 196

© Crown Copyright 1970

A. R. SHEARER, Government Printer, Wellington, New Zealand--1969



FOREWORD

Over the last few years the attention paid to studies of the Recent Foraminifera has notably increased. These investigations have been considerably aided by the publication by A. R. Loeblich and H. Tappan of their revised classification of the Foraminifera in the "Treatise on Invertebrate Paleontology" series. The present memoir extends the general usefulness of this work by providing a key to the Recent genera.

J. W. BRODIE, Director,
N.Z. Oceanographic Institute,
Wellington.

CONTENTS

	<i>Page</i>
Abstract	7
Introduction	7
Using the Key	7
Key to the Foraminiferida	8
Key to the Suborders of Foraminiferida	8
Key to New Zealand Genera with Tectinous Tests, Chiefly Suborder Allogromiina	8
Key to Recent Genera of Suborder Textulariina	8
Ammodiscacea	8
Lituolacea	14
Key to Recent Genera of Suborder Miliolina	22
Miliolacea	22
Key to Recent Genera of Suborder Rotaliina with Classification of Superfamilies	30
Nodosariacea	35
Buliminacea	40
Discorbacea	45
Spirillinacea	49
Rotaliacea	49
Globigerinacea	53
Orbitoidacea	55
Cassidulinacea	59
Carterinacea	65
Robertinacea	65
Illustrated Glossary	67
Useful Techniques	79
References	83
Index	84

A Key to the Recent Genera of the Foraminiferida

K. B. LEWIS

New Zealand Oceanographic Institute, D.S.I.R. Wellington

Abstract

A KEY with illustrated glossary is provided to all the Recent agglutinated and calcareous foraminiferal genera recorded by Loeblich and Tappan in the *Treatise on Invertebrate Paleontology*, with special reference to those occurring in the New Zealand region. The few tectinuous forms recorded from this area are also included. A summary is given of techniques used by workers on Recent foraminifers.

INTRODUCTION

THE recently proposed classification of the Foraminiferida by Loeblich and Tappan (1964a) incorporates much up-to-date knowledge of these animals, but does not, unlike a number of its predecessors, contain a key. The construction of a workable key to all the genera from Cambrian to Recent would have been an enormous undertaking; however, a key to the Recent genera has been prepared in conjunction with, and to facilitate, current research on the Recent foraminiferal fauna of New Zealand.

The key form is simple, with the parts of the couplet together (cf. Metcalf 1954), except in a note on the natural classification of Suborder Rotaliina which is illustrated in a small separate key with the parts of the couplet separated and successive couplets indented.

The classification proposed by Loeblich and Tappan is adhered to throughout and the order of families and subfamilies is the same as in the *Treatise on Invertebrate Paleontology*. New genera proposed by these authors since the *Treatise* went to press have been included, and notes made of their revisions to the classification.

The key in many instances cuts across the natural classification, but each genus is recorded only under the

family name or subfamily in which it appears in the *Treatise*. All taxa have a reference to their description in the *Treatise*.

Using the Key

To use the key: beginning with couplet 1 (p. 8) choose one of the alternatives and proceed to the couplet indicated to the right of that alternative, e.g. if the specimen is calcareous, proceed to couplet 2, and continue in this way until a determination is reached. The key progresses to family and subfamily, but may continue through several different families before reaching a generic identification. The generic name is followed by a reference to a description and figure by Loeblich and Tappan either in the *Treatise on Invertebrate Paleontology* or in their modifications of the work, and genera recorded from the New Zealand Region (Eade, 1967) are indicated with †, e.g. *Astrorhiza*† C184 [103] is described in the *Treatise* on p. C184 and illustrated in Fig. 103, and species recorded from the New Zealand region are listed in *A Checklist of Recent New Zealand Foraminifera*.

KEY TO THE FORAMINIFERIDA

KEY TO THE SUBORDERS

1. Tectinous test (chiefly Subord. ALLOGROMIINA—New Zealand forms only)	..	3
Agglutinated test (Subord. TEXTULARIINA)	5
Calcareous test	2
2. Test imperforate, porcellanous (Subord. MILIOLINA)	184
Test perforate, hyaline (Subord. ROTALIINA)	276

KEY TO THE NEW ZEALAND GENERA OF THE SUBORDER ALLOGROMIINA (Loeblich and Tappan 1964 p. C164)

Sarcodina with tectinous tests cannot be identified without detailed work on the living animal and are therefore outside the scope of this key. However the few forms recorded from New Zealand can readily be distinguished by test morphology.

Forms with biflagellate gametes are included in the Family Lagnidae C164
Forms with amoeboid gametes are referred to the Family Allogromiidae C173

3. Attached (commonly with few adherent sand grains) <i>Iridia</i> †	..	C167 [86]
Free living	4
4. Elongate tubular	<i>Shepherdella</i> †
Ovate to spherical	<i>Allogromia</i> † ..
		C182 [102] C173 [92]

Note: *Gromia*† (p. C40 Fig. 21) has filopodia and uniflagellate gametes. It is not strictly a foraminifer.

KEY TO THE RECENT GENERA OF THE SUBORDER TEXTULARIINA (Loeblich and Tappan 1964, p. C184)

5. Unilocular, single rounded chamber and/or non-septate tube or tubes	..	Superfam. AMMODISCACEA ..	6
Multilocular, test with many distinct chambers	Superfam. LITUOLACEA ..	84

AMMODISCACEA (C184)

6. Essentially a globose or discoidal chamber, may have tubular arms, may be in groups	Fam. SACCAMMINIDAE ..	37
Essentially tubular chamber; straight, enrolled or branching	7	
7. Enrolled tubular second chamber	..	Fam. AMMODISCIDAE ..	73
Straight or branching tubular chambers, never enrolled	8	
8. With smooth outer covering of tectine, branching in one plane	Fam. SCHIZAMMINIDAE ..	36
No outer covering of tectine, straight or branching in one or many planes	Fam. ASTORRHIZIDAE ..	9

ASTRORHIZIDAE (C184)

9.	Test free	10		
	Test attached or mass of entwined tubes	Subfam. DENDROPHRYINAE	...	29
10.	Simple tube, with open ends, rarely branching	Subfam. RHIZAMMININAE	...	18
	With proloculus or central chamber, tube open at one end only	11		
11.	Central chamber with radiating arms ..	Subfam. ASTRORHIZINAE	..	13
	Proloculus continued to single, non-septate, tube	12		
12.	Wall labyrinthic	Subfam. BOTELLININAE	..	28
	Wall not labyrinthic	Subfam. HIPPOCREPININAE	..	23

ASTRORHIZINAE (C184)

13.	Central chamber with tectinous sides, arms and "framework" agglutinated	<i>Vanhoeffenella</i>	..	C186 [104]
	Central chamber completely agglutinated	14		
14.	Central chamber discoidal with short arms ..	<i>Astrorhiza</i> †	..	C184 [103]
	Central chamber not discoidal	15		
15.	Central chamber large spherical, few short arms ..	16		
	Central chamber small, often indistinct	17		
16.	Tubular arms with open ends	<i>Astrammina</i>	..	C184 [103]
	Domed or conical protruberances with small opening at extremity	56		
17.	Arms numerous, irregular, very fine grained ..	<i>Radicula</i>	..	C185 [104]
	Arms few, usually straight, medium-coarse grained ..	<i>Rhabdammina</i> †	..	C185 [103]

Note: *Marsipella*† C186 [105] is elongate fusiform.

RHIZAMMININAE (C186)

18.	Branching	19		
	Not branching	20		
19.	Tubes radiating from small central chamber, seen when test broken or sectioned	17		
	No rounded cavity at point of branching	<i>Rhizammina</i> †	C186 [105]
	Note: <i>Schizammina</i> C194 [107] has labyrinthic wall. <i>Dendronina</i> † C192 [108] is constructed of aligned sponge spicules.			
20.	Elongate fusiform, slightly wider in middle ..	21		
	Simple tube	22		
21.	Test medium to coarse-grained, may include unaligned sponge spicules	<i>Marsipella</i> †	..	C186 [105]
	Test fine-grained or composed completely of aligned sponge spicules	53		
22.	Tube small, thin-walled, usually curved; fine arenaceous groundmass with irregular covering of larger fragments and commonly other foraminifer tests	<i>Rhizammina</i> †	C186 [105]
	Tube large, thick, almost straight; sand and sponge spicules in calcareous cement; one end of tube may be blocked with waste material	<i>Bathysiphon</i> †	C186 [105]

Note: *Protobotellina* C190 [106] has base blocked with sand grains.

HIPPOCREPININAE (C187)

23. Tube branching Saccorhiza† C190 [106]
 Tube not branching 24
24. Test conical; possibly very elongate cone .. 25
 Test cylindrical 26
25. Test simple; elongate cone; aperture open end of tube Jaculella† C190 [106]
 Test with irregular transverse constrictions; small terminal aperture Hippocrepina C188 [106]
 Note: *Pelosina* C200 [112, 113] has no transverse constrictions.
26. With bulbous proloculus; tubular portion generally smaller diameter than proloculus 27
 No bulbous base; simple tube with one end closed by arenaceous material Protobotellina C190 [106]
 Note: *Bathysiphon*† C186 [105] has end closed by black waste material. *Brachysiphon*† C196 [112] is much shorter. *Nubeculariella* C192 [108] is tectinous with a few large sand grains.
27. Tube with internal partial septa 91
 Tube without partial septa Hyperammina† C190 [106]
 Note: *Lagenammina* C200 [113] has flask-shaped test with more bulbous chamber and narrower tube.

BOTELLININAE (C190)

28. Botellina† C190 [107]
 Note: *Schizammina* C194 [107] has smooth tectinous outer surface.

DENDROPHRYINAE (C192)

29. Tubes growing upwards from attached base, or mass not firmly attached to substrate 30
 Tubes growing along and firmly attached to substrate for at least part of their length 35
30. Tubes simple or branching, growing upwards from attachment 31
 Complicated mass of tubes 34
31. Short pseudochitinous tube with a few large grains attached Nubeculariella C192 [108]
 Test normally agglutinated, elongate, often branching 32
32. Erect portion of test conical; diameter of tube increasing from attachment upwards Halyphysema C192 [108]
 Erect portion tubular; if anything, diameter decreases slightly away from attachment 33
33. May branch; wall of fine sand and sponge spicules aligned in direction of growth Dendronina† C192 [108]
 Branching, spreading; wall of fine sand, not sponge spicules Dendrophrya C192 [108]
 Note: *Saccodendron* C205 [118] has hemispherical to irregularly ovate chamber attached to substrate.
34. Mass of anastomosing tubes, may be organised into radiating tubes joined by networks of connecting tubes Syringamina† C192 [108]
 Central mass with radiating tubular portions, each expanding distally into a globular or conical mass Normanina C192 [108]
35. Proloculus with single unbranched elongate tube .. 81
 Two or more tubes from periphery of globular chamber or attached branching tubes, or both 60

SCHIZAMMINIDAE (C192)

36. Tubular with dichotomous branching *Schizammina* C194 [107]
 Flabelliform; margin with projecting tubules *Jullienella* C194 [107]

SACCAMMINIDAE (C194)

37. Interior not divided into chamberlets;
 wall not labyrinthic 38
 Interior divided into chamberlets; every
 gradation between partial septa and a
 thick labyrinthic wall Subfam. DIFFUSILININAE 65
38. Free living 39
 Attached Subfam. HEMISPHAERAMMININAE 57
39. No aperture Subfam. PSAMMOSPHAERINAE 40
 With definite apertures, possibly at end
 of tubes Subfam. SACCAMMININAE 45

PSAMMOSPHAERINAE (C194)

40. Single globular or discoidal chamber 41
 Several loosely joined globular chambers *Sorosphaera* C196 [110, 111]
 Note: *Psammophax* is synonymised with *Sorosphaera*, but original figures of *P. consociata* and Ross
 Sea material show single rounded aperture in each group of chambers (J. P. Kennett, 1968).
Normanina C192 [108] has tubes radiating from central mass which expand distally into globular
 chambers.
41. Discoidal 42
 Spherical 43
42. Without radiating arms; central area tectinous *Amphifenestrella* C195 [111]
 With radiating arms; central area tectinous or
 completely arenaceous 13
43. Simple globular chamber; no projections *Psammospaera*† C195 [110]
 With numerous projections 44
44. Projections irregular, without small openings at ends *Storthospaera*† C196 [111]
 Projections more regular, with small apertures at
 ends 16

SACCAMMININAE (C196)

45. Single aperture 46
 Two or more tubular extensions, or “apertures”
 where the extensions have been broken off 52
46. Hemispherical; aperture on concave side *Causia* C200 [113]
 Globular, fusiform, or subcylindrical 47
47. Test composed mainly of longitudinally aligned
 sponge spicules 48
 Sponge spicules not principal constituent 49
48. Test oviform to fusiform or cylindrical; rounded
 aperture *Technitella*† C202 [115]
 Test spherical with slit-like aperture *Pilulina* C201 [112]
49. Globular; main chamber does not have parallel
 sides 50
 Subcylindrical; with parallel sides *Brachysiphon*† C196 [112]
 Note: *Protobotellina* C190 [106] has a comparatively elongate cylinder.
50. Test almost spherical; neck, if any, short 51
 Test flask-shaped; aperture on elongate neck *Lagenammina* C200 [113]
 Note: *Hyperammina*† C190 [106] has a comparatively wide tube and small proloculus. *Pelosina*†
 C200 [112, 113] has opening at both ends.



51. Aperture slit-like *Pilulina* C201 [112]
 Aperture circular *Saccamina*† C196 [112]
 Note: *Pelosphaera* C201 [112, 114] has tectinous entosolenian tube. *Psammophax* is synonymised with *Sorosphaera* C196 [110, 111]. However, original figures of *P. consociata* and Ross Sea material show single rounded aperture in each group of chambers (J. P. Kennett, 1968).
52. Test fusiform to almost globular; projections at opposite ends 53
 Test roughly spherical or discoidal; two or more projections around test 54
53. Test composed mainly of longitudinally aligned sponge spicules *Technitella*† C202 [115]
 Test composed of fine mineral grains *Pelosina*† C200 [112, 113]
 Test composed of coarse mineral grains possibly with some sponge spicules 21
 Note: *Vanhoeffenella* C186 [104] has chamber centres of tectin.
54. Roughly spherical; projections randomly distributed 55
 Discoidal; projections around periphery 13
55. Projections sturdy, arenaceous 56
 Projections fragile, flexible, argillaceous, tubular, bifurcating near extremities; if projection broken, tectinous entosolenian tube can be seen *Pelosphaera* C201 [112, 114]
56. True tubular arms 16
 Domed or conical protruberances with small opening at extremity *Thuramina*† C202 [115]

HEMISPHERAMMININAE (C202)

57. Solitary chamber 58
 Several chambers in loose whorl, ring, or linear series 64
58. With tubular extensions which remain attached to substrate for at least part of their length 59
 Test roughly hemispherical, without tubular extensions 62
59. Tubes branching 60
 Tubes not branching 61
60. Hemispherical to ovate chamber; extensions arising from peripheral area and may grow free of attachment *Saccodendron* C205 [118]
 No central chamber; branching tubes attached to substrate throughout *Sagenina* C205 [117]
61. Single elongate tube 81
 Two or more short protruberances *Tholosina*† C205 [117]
 Note: *Iridia* C167 [86] has wall with tectinous base and little cement.
62. With apertures (may be small at periphery) 63
 Without apertures *Hemisphaerammina*† C202 [115]
 Note: *Iridia* C167 [86] has wall with tectinous base and little cement.
63. Aperture at periphery *Tholosina*† C205 [117]
 Aperture at summit of chamber *Colonamina* C204 [117]
 Note: *Verrucina* C210 [119] has thick labyrinthic wall.
64. Chambers hemispherical in loose whorl or ring; aperture at summit of each chamber *Ammopemphix* C202 [117]
 Chambers pyriform in linear series; aperture terminal against attachment *Goatapitigba* C795 [652]

DIFFUSILININAE (C205)

65.	Free-living	**	**	**	**	**	66		
	Attached	**	**	**	**	**	70		
66.	Spherical	**	**	**	**	**	67		
	Discoidal to lenticular	**	**	**	**	**	68		
67.	Thick labyrinthic wall; small central cavity	**	**	**	**	**		<i>Oryctoderma</i>	C208 [120]
	Final chamber enveloping early planispiral development	**	**	**	**	**	117		
68.	Thin wall; cavity divided by radial septa; test fairly thick; oblong in section	**	**	**	**	**		<i>Daitrona</i> †	C205 [120]
	Labyrinthic wall; test much flattened in section	**	**	**	**	**	69		
69.	Very compressed central cavity; fine branching tubules extending to periphery	**	**	**	**	**		<i>Masonella</i>	C208 [119]
	Inner layer of loosely cemented larger grains; outer layer well cemented fine grains; concentric growth rings; often two eccentric perforations through test	**	**	**	**	**		<i>Discobotellina</i>	C205 [119]
70.	Irregular mass; interior ramifying tubules; apertures at ends of one to four irregularly spaced pustules	**	**	**	**	**		<i>Diffusilina</i>	C205 [119]
	Regular subspherical or hemispherical chambers, sometimes with single aperture	**	**	**	**	**	71		
71.	With single partial septum	**	**	**	**	**		<i>Crithionina</i>	C205 [120]
	With several or numerous short radial portions	**	**	**	**	**	72		
72.	With single aperture in depressed area at summit of test	**	**	**	**	**		<i>Verrucina</i>	C210 [119]
	No aperture	**	**	**	**	**		<i>Pseudowebbinella</i>	C208 [120]

AMMODISCIDAE (C210)

73.	Test free	**	**	**	**	**	Subfam. AMMODISCINAE	**	74
	Test attached	**	**	**	**	**	Subfam. TOLYPAMMININAE	**	81

AMMODISCINAE (C210)

74.	Axis of coiling the same throughout (planispiral and trochospiral forms)	**	**	**	**	**	75		
	Axis of coiling changing (streptospiral and irregularly coiled forms)	**	**	**	**	**	79		
75.	Tubular chamber laterally compressed, tending to uncoil	**	**	**	**	**		<i>Psammonyx</i>	C212 [124]
	Tubular chamber circular or semicircular in section; closely coiled	**	**	**	**	**	76		
76.	Wall tectinous; outer whorl bordered by an agglutinated layer	**	**	**	**	**		<i>Spirillinoides</i>	C212 [124]
	Wall arenaceous throughout	**	**	**	**	**	77		
77.	Trochospiral at some stage	**	**	**	**	**	78		
	Planispiral, no trochospiral stage	**	**	**	**	**		<i>Ammodiscus</i> †	C210 [122]
	Note: <i>Discobotellina</i> C205 [119] has annular growth rings, not an enrolled tube.								
78.	Low trochospiral in early stages, but later planispiral or trochospiral in reverse direction	**	**	**	**	**		<i>Ammodiscoides</i> †	C210 [122]
	Elongate, high trochospiral	**	**	**	**	**		<i>Turritellella</i> †	C212 [122]
79.	Streptospiral to planispiral to final uncoiled zigzag growth	**	**	**	**	**		<i>Ammovertellina</i>	C210 [123]
	No final zigzag stage	**	**	**	**	**	80		



80. Initial high-spired coil to planispiral stage with axis of coiling almost perpendicular to original axis .. *Usbekistania* C212 [125]
 Streptospiral or irregular coil .. *Glomospira*† C212 [122]
 Note: *Lituotuba* C214 [126] has later stages uncoiling.

TOLYPAMMININAE (C213)

81. Growth in zigzag fashion *Ammovertella* C214 [126]
 Growth not zigzag 82
 82. Early planispiral, streptospiral or irregular coil; later uncoiling *Lituotuba* C214 [126]
 No early coiled stage 83
 83. After globular proloculus, winds irregularly over surface of attachment *Tolypamina*† C213 [126]
 Flask-shaped chamber with straight tubular neck *Ammolagena*† C214 [126]

LITUOLACEA (C214)

84. Uniserial series Fam. HORMOSINIDAE 89
 Biserial series; may have final uniserial stage Fam. TEXTULARIIDAE 132
 Coiled or irregular, at least in initial stages 85
 85. Irregular development of globose chambers 86
 Milioline arrangement with two (rarely three) tubular chambers per whorl Fam. RZEHAKINIDAE 99
 Planispiral or streptospiral, at least in early stages 87
 Trochospiral, at least in early stages 88
 86. Free 40
 Attached 64
 87. Close coiled or uncoiling to uniserial series Fam. LITUOLIDAE 104
 Uncoiling to biserial series Fam. TEXTULARIIDAE 132
 88. Low trochospiral Fam. TROCHAMMINIDAE 146
 High trochospiral usually with later biserial or uniserial stage Fam. ATAXOPHRAGMIIDAE 161
 Note: Fam. NOURIIDAE (polymorphine spiral) and Fam. PAVONITINIDAE (microgranular calcite wall) key out with Fam. ATAXOPHRAGMIIDAE.

HORMOSINIDAE (C214)

89. Free-living 90
 Attached 130
 90. Chambers long and tubular or irregularly globose; each chamber typically with individual aperture Subfam. ASCHEMONELLINAE 91
 Chambers globose or short and flattened; only single terminal aperture Subfam. HORMOSININAE 92

ASCHEMONELLINAE (C214)

91. With rounded apertures on tubular necks, one or more on each chamber; commonly branching *Aschemonella* C214 [127]
 With only single terminal aperture; not branching; chambers long and tubular; sutures often indistinct *Kalamopsis* C215 [127]

HORMOSININAE (C215)

92.	Uniserial throughout . .	++	++	++	93		
	Note: Examine initial end very carefully. Specimens may have to be broken, sectioned, or immersed in aniseed oil to reveal initial coil.						
	Uniserial with initial coil or biserial stage			++	97		
93.	Branching . .	++	++	++	94	<i>Protoschista</i>	C215 [130]
	Not branching	++	++	++			
94.	Aperture curved slit, usually in slight depression on terminal face	++	++	++	95	<i>Sulcophax</i>	C217 [128]
	Aperture rounded, often on apertural neck			++			
95.	Aperture circular, no tooth	++	++	++	96		
	Aperture rounded, with single tooth			++	174		
	Aperture radiate because of several ribs or teeth on inside of apertural neck	++	++	++		<i>Nodosinum</i>	C215 [129]
	Note: <i>Nubeculina</i> C456 [346] has porcellaneous neck.						
	Aperture cribrate	++	++	++	141		
96.	Very fine-grained; surface smooth; chambers almost spherical . .	++	++	++		<i>Hormosina</i> †	C215 [128]
	Medium to coarsely arenaceous; surface rough; chambers not necessarily spherical			++		<i>Reophax</i> †	C216 [128]
97.	Initial biserial stage . .	++	++	++	132		
	Initial planispiral stage	++	++	++	104		
	Initial trochospiral stage	++	++	++	161		

NOURIIDAE (C220)

98.		++	++	++		<i>Nouria</i> †	C220 [132]
-----	--	----	----	----	--	-----------------	------------

RZEHAKINIDAE (C220)

99.	Three chambers to whorl	++	++	++		<i>Ammofintina</i> . .	C220 [133]
	Two chambers to whorl, arranged in various planes about vertical axis . .	++	++	++	100		
100.	Test planispiral or sigmoidal . .	++	++	++	101		
	Test triloculine	++	++	++		<i>Trilocularena</i> . .	C224 [134]
	Test quinqueloculine . .	++	++	++	102		
	Chamber added in more than five planes about vertical axis	++	++	++	257*		
101.	Aperture rounded on neck and without tooth; no early quinqueloculine stage . .	++	++	++		<i>Spirolocamina</i>	C222 [134]
	Aperture rounded, with small tooth, not on pronounced neck; early quinqueloculine stage or sigmoidal arrangement	++	++	++	244*		
	Aperture cribrate; early quinqueloculine stage	++	++	++	257*		
102.	Margin of aperture crenulate	++	++	++	245*		
	Margin of aperture smooth	++	++	++	103		
103.	Wall with calcareous base; tooth and lip of aperture porcellaneous; disintegrates rapidly in dil. HCl	++	++	++	245*		
	Wall without calcareous base; almost insoluble in dil. HCl . .	++	++	++		<i>Miliamina</i> † . .	C220 [134]

*These genera have a calcareous layer below the arenaceous layer and are therefore placed in the Miliolidae.

LITUOLIDAE (C225)

104.	Planispiral or streptospiral throughout; not uncoiling	++	++	++	105		
	Uncoiling after early planispiral stage	++	++	++	107		



105.	Wall simple	106		
	Wall labyrinthic	Subfam. CYCLAMMININAE	..	119
106.	Only one to three chambers visible externally; test globose	Subfam. SPHAERAMMININAE	..	116
	More than three chambers visible externally; test rarely globose	Subfam. HAPLOPHRAGMOIDINAE	..	109
107.	Test free	Subfam. LITUOLINAE	..	122
	Test attached	108		
108.	Wall labyrinthic	Subfam. COSCINOPHRAGMATINAE		131
	Wall not labyrinthic	Subfam. PLACOPSILININAE	..	130

HAPLOPHRAGMOIDINAE (C225)

109.	Axis of coiling the same throughout	110		
	Streptospirally coiled	115		
110.	Septa thick and arenaceous representing previous apertural faces; wall finely to fairly coarsely arenaceous not appearing labyrinthic					111		
	Septa thin and tectinous; wall coarsely arenaceous giving interior labyrinthic appearance	<i>Discammina</i> †	..	C226 [136, 137]
111.	Interiomarginal aperture, bordered by lip on upper margin only	112		
	Areal aperture, may be a slit very low down an apertural face with only faint lip between it and previous whorl, i.e. lips on upper and lower margins	114		
112.	Aperture equatorial; test symmetrical; typically short axis of coiling	113		
	Aperture to one side of periphery; test slightly asymmetrical; globose, with elongate axis of coiling	<i>Adercotryma</i> †	..	C225 [135]
	Note: <i>Thalmannammina</i> C226 [136] has axis of coiling changing throughout growth.							
113.	Involute to slightly evolute	<i>Haplophragmoides</i> †	..	C225 [135]
	Evolute	<i>Trochamminoides</i>	..	C227 [136]
114.	Aperture slit-like, or series of pores in line, commonly low down on apertural face	<i>Cribrostomoides</i> †	..	C225 [136]
	Aperture multiple, rarely single, rounded with prominent raised lips scattered over apertural face	<i>Trochamminita</i> †	..	C226 [135]
	Note: Later chambers of <i>Trochamminita</i> are often irregular so may appear to be trochospiral.							
115.	Aperture areal, usually rounded with lip	<i>Recurvoidea</i>	..	C226 [136]
	Note: <i>Trochamminita</i> † C226 [135] may have last few chambers irregular. <i>Ammofofintina</i> C220 [133] has three tubular chambers per whorl.							
	Aperture interiomarginal	<i>Thalmannammina</i>	..	C226 [136]
	Note: <i>Adercotryma</i> † C225 [135] has axis of coiling same throughout and a symmetrical aperture. <i>Ammosphaeroidina</i> † C259 [174] has only two or three globular chambers visible.							

SPHAERAMMININAE (C227)

116.	Free-living	117		
	Attached	<i>Ammosphaerulina</i>	..	C227 [139]
117.	Aperture interiomarginal or slit paralleling and close to basal margin	148		
	Aperture areal, rounded or slit-like, not close to basal margin	118		

118. Aperture with tooth; wall fine-grained .. ** *Sphaerammina* C227 [139, 140]
 Aperture simple slit, no tooth; wall coarse-grained *Canepaia* ** C228 [141]

CYCLAMMININAE (C228)

119. Septa arenaceous; coiling involute to slightly evolute 120
 Septa thin tectinous; coiling evolute to partly involute, wall very coarsely arenaceous ** 110
120. Interiomarginal aperture only, has single lip on upper margin ** ** ** ** *Reticulophragmium* ** C233 [142]
 With areal aperture, may be a slit low down on apertural face with only very faint lip between it and previous whorl, i.e. lips on upper and lower margins .. ** ** ** ** 121
121. Slit-like aperture low down on apertural face ** *Alveolophragmium*† ** C228 [143]
 Intermarginal aperture plus series of rounded pores with raised margins scattered over apertural face ** ** ** ** *Cyclammina*† ** C228 [142]

LITUOLINAE (C238)

122. Uniserial portion triangular or quadrate in section *Triplasia* ** ** C245 [155]
 Uniserial portion circular or compressed in section 123
123. Biserial stage between planispiral and uniserial stages ** ** ** ** 124
 No biserial stage in development .. ** ** 125
124. Aperture rounded; periphery typically rounded ** *Ammobaculoides* ** C241 [151]
 Aperture slit-like; periphery acutely angular ** 133
125. Initially close-coiled; almost straight uniserial portion growing off at tangent to original coil ** 126
 Gradually uncoiling series; chambers elongate, extending down towards coil at inner margin ** 129
126. Aperture cribrate ** ** ** ** *Lituola* ** ** C238 [151]
 Aperture single .. ** ** ** 127
127. Aperture terminal but displaced towards outer rim; sutures oblique; test compressed *Ammomarginulina* ** C241 [151]
 Aperture terminal on axis running through centre of uniserial portion; sutures roughly perpendicular to growth; usually circular or oval in section 128
128. Chambers separated by porcellaneous calcite septa or long porcellaneous stolons 221
 Chambers separated by normal arenaceous septa, i.e., terminal portion of earlier chamber *Ammobaculites*† ** C239 [151]
 Chambers separated by straight tectinous septa .. *Ammoscalaria*† ** C241 [152]
 Undivided tube but growth constrictions give the appearance of sutures ** ** ** 82
129. Aperture rounded terminal, at peripheral angle .. *Ammotium*† ** ** C241 [152]
 Aperture transverse areal slit near centre of terminal face; secondary aperture cribrate at lower end of final chamber near proloculus ** ** *Ammoastuta* ** C238 [151]

PLACOPSILININAE (C247)

130. Aperture single or double row of pores; test rarely branching 131
 Aperture slit-like with projecting teeth; test may be branching *Haddonia* C248 [159]
 Aperture rounded; test not branching *Placopsilina*† C247 [159]
 Note: *Goatapitigba* C795 [652] has few pyriform chambers. *Lituotuba* C214 [126] has growth constrictions but no true septa.

COSCINOPHRAGMITINAE (C248)

131. *Bdelloidina* C250 [126]

TEXTULARIIDAE (C250)

132. No initial coiled stage (mainly Subfam. TEXTULARIINAE plus Subfam. TAWI-TAWIINAE and Subfam. PSEUDOBOLIVININAE) 136
 Note: Examine initial end very carefully. Specimens may have to be broken, sectioned, or soaked in aniseed oil to reveal initial coil.
 Initial planispiral or streptospiral stage .. Subfam. SPIROPECTAMMININAE .. 133
 Initial trochospiral stage 161

SPIROPECTAMMININAE (C251)

133. Aperture low interiomarginal arch; no uniserial stage 134
 Aperture areal terminal in adult uniserial development 124
 134. Initial planispiral coil 135
 Initial streptospiral coil *Morulaepectra* C251 [163]
 135. Acutely angled periphery; broad low chambers, outer edges curving down towards proloculus; finely arenaceous; adult or gerontic stage may be uniserial with areal slit in plane of compression *Vulvulina* C251 [163]
 Periphery rounded to fairly angular; chambers not low and broad; never any final uniserial development *Spiropectamina*† C251 [163]
 Note: Microspheric specimens of *Textularia*† C253 [165] may have initial coil.

TEXTULARIINAE (C253)

136. Aperture low interiomarginal arch 137
 Aperture areal in adult; in early stages may be interiomarginal arch or slit-like up apertural face 138
 137. Test attached, at least in early stages *Textularioides* C255 [165]
 Test free throughout life *Textularia*† C253 [165]
 Note: *Textulariella*† C299 [202] has labyrinthic interior and exterior of microgranular calcite.
 138. Aperture multiple in adult 139
 Aperture single in adult 142
 139. Test compressed; aperture linear series of openings across terminal portion of final chamber 140
 Test rounded; aperture group of pores not in linear series 141
 140. Interior simple *Poritextularia* C254 [165]
 Interior labyrinthic 145

141. Uniserial portion absent or of only few chambers;
aperture one to three rounded openings .. 144
Uniserial portion well developed; aperture cribrate *Cribrrobigenina* .. C254 [166]
Note: *Liebusella*† C287 [195] has internal chamber partitions.
142. No true uniserial portion with horizontal sutures,
although may tend towards uniseriality .. 143
Well developed uniserial portion with horizontal
sutures .. *Bigenina*† .. C254 [165]
Note: *Vulvulina* C251 [163] has terminal slit-like aperture in plane of compression.

PSEUDOBOLIVININAE (C255)

143. Roughly quadrangular in section .. *Siphotextularia*† .. C258 [168]
Not quadrangular in section .. 144
144. Twisted biserial; aperture slit-like to rounded,
terminal on short neck .. *Pseudobolovina*† .. C255 [167]
Plane of growth not twisted; aperture one to three
rounded openings, without neck .. *Planctostoma* .. C256 [168]

TAWITAWIINAE (C258)

145. .. *Tawitawia* .. C258 [170]

TROCHAMMINIDAE (C259)

146. Interior simple .. Subfam. TROCHAMMININAE .. 147
Interior partly divided by infolding of
septial wall .. Subfam. REMANEICINAE .. 160

TROCHAMMININAE (C259)

147. Total number of visible chambers few (less than
five), usually globose; form of coiling often
difficult to see .. 148
Chambers numerous, usually somewhat flattened;
obvious true trochospiral coil .. 151
148. Aperture areal, may be slit-like, paralleling and near
to basal margin .. 149
Aperture interiomarginal .. 150
149. Coiling trochospiral; chambers domed to high-
oviform; aperture slit or oval, close to basal
margin .. *Cystamina*† .. C263 [177]
Coiling planispiral; chambers rounded, embracing;
aperture slit well removed from basal margin .. 116
150. Coiling trochospiral arenaceous isomorph of
Globigerina .. *Trochamina*† .. C259 [173]
Coiling streptospiral arenaceous isomorph of
Sphaeroidina .. *Ammosphaeroidina*† .. C259 [174]
Note: *Thalmanamma* C226 [136] has more than three chambers visible externally. *Rugidia* C587
[462] is calcareous, perforate with rugose appearance. *Adercotryma*† C225 [135] is planispirally coiled
with eccentric aperture.
151. Test free, not deformed .. 152
Test attached or deformed to shape of some
attachment .. 159
152. With final stage uncoiling biserial .. 169
Coiled throughout .. 153
153. Low interiomarginal aperture only, extraumbilical-
umbilical, usually difficult to see .. 154
With areal aperture or slit up apertural face .. 156

154. Aperture rounded at end of forward directed siphon-like lobe at umbilical margin of final chamber *Siphotrochammina* .. C266 [174]
Aperture low arch or slit-like 155
155. Ventral sutures sinuate; final chamber appears to be T-shaped *Tiphotrocha* .. C266 [174]
Ventral sutures curved not sinuate *Trochammina*† .. C259 [173]
- Note: *Thalmanammina* C226 [136] is streptospirally coiled. *Adercotryma*† C225 [135] is planispirally coiled with eccentric aperture.
156. Having slit-like aperture extending up apertural face; may have secondary aperture 157
Having rounded areal apertures without slit up apertural face 158
157. With numerous circular openings near peripheral angle *Arenoparrella* .. C262 [173]
Without secondary cribrate aperture *Trochamminula* .. C266 [173]
158. Two pairs of openings symmetrical on apertural face, lower pair large and ovate, upper pair small and rounded *Entzia* .. C264 [176]
One to several openings randomly placed on apertural face often with lips *Jadammina* .. C265 [176]
- Note: *Tritaxis*† C266 [177] is without primary low interiomarginal arch; flattened on umbilical side; usually attached in later stages. *Trochamminita* C226 [135] is planispiral with final irregular chambers. *Recurvoides*† C226 [136] is streptospirally coiled.
159. Rather coarse-grained; three or four chambers per whorl; free-living early stage with areal ovate aperture with lip *Trixtaxis*† .. C266 [177]
Fine-grained; more than four chambers per whorl; flexible test bending round attachment *Rotaliammina* .. C265 [174]

REMANEICINAE (C266)

160. Test of agglutinated material *Remaneica*† .. C266 [178]
Test of secreted fusiform crystals of calcite 723

ATAXOPHRAGMIIDAE (C268)

161. Without internal pillars or partitions 162
With internal vertical pillars or partitions Subfam. ATAXOPHRAGMIINAE .. 181
162. Without tooth protruding into aperture 163
With apertural tooth Subfam. VALVULININAE .. 174
163. Initial part triserial, usually triangular in section Subfam. VERNEUILININAE .. 164
Initial part not triserial nor triangular, usually more than three chambers in initial whorl Subfam. GLOBOTEXTULARIINAE .. 169

VERNEUILININAE (C268)

164. Triserial and triangular throughout 165
Triserial reducing to biserial and even uniserial 166
Triserial increasing to five or more chambers per whorl 177
165. Aperture interiomarginal, arched *Verneuilina*† .. C268 [179]
Aperture areal, terminal, rounded *Barbourinella* .. C268 [179]
166. Final stage biserial 167
Final stage uniserial 168

167. Final stage irregularly spreading, aperture terminal rounded with lip on short neck Rudigaudryina .. C272 [181]
 Straight biserial series; aperture interiomarginal, arched Gaudryina† .. C269 [179]
 Note: Many species of *Gaudryina* show tendency toward *Tritaxia* C272 [182] by developing final uniserial stage with terminal aperture in gerontic specimens. Character of major part of population must determine generic position.
168. Final stage irregularly spreading Rudigaudryina .. C272 [181]
 Straight uniserial stage Tritaxia .. C272 [182]
 Note: See note for *Gaudryina* above.

GLOBOTEXTULARIINAE (C273)

169. Growth into high spire; axis of coiling or growth same throughout 170
 Coiling in low trochospiral; later biserial portion growing with axis perpendicular to axis of coil Plectotrochammina .. C279 [185]
170. Final stage with four chambers in the whorl Globotextularia† .. C273 [183]
 Final stage triserial Eggerella .. C275 [186]
 Note: *Tritaxis*† C266 [177] has flattened umbilical side attached to substrate.
- Final stage biserial 171
 Final stage uniserial 173
171. Aperture areal, may have rounded lip or short neck 172
 Aperture interiomarginal Dorothisia† .. C275 [184]
 Note: *Textulariella*† C299 [202] has labyrinthic interior. *Morulaeplecta* C251 [163, 164] has initial streptospiral coil.
172. Chambers few, elongate, overhanging laterally, in polymorphine spiral; sutures at high angle 98
 Chambers many, not elongate or strongly overhanging; sutures usually at low angle Karreriella† .. C277 [186]
173. Uniserial portion short (about six chambers); aperture terminal, cribrate Cribrogoesella .. C273 [184]
 Note: *Goesella*† C281 [188] has rounded aperture with tooth. *Liebusella*† C287 [195] has labyrinthic interior.
- Uniserial portion long, giving narrow, very elongate test; aperture variously aligned slits with bordering lips Multifidella .. C277 [185]
 Note: *Martinottiella*† C282 [188] has single slit-like or arcuate aperture.

VALVULININAE (C279)

174. With initial trochospiral coil 175
 Uniserial throughout; no initial coil Dusenburyina .. C281 [188]
 Note: *Cylindroclavulina*† C281 [188] has an initial coil which is often very difficult to see.
175. Initial portion triserial; usually triangular in section 176
 Initial portion not triserial or triangular in section; usually more than three chambers in the initial whorl 179
176. Triserial throughout Valvulina .. C279 [187]
 Only initial stage triserial 177
177. Later portion in loose spiral; five or more chambers to whorl; large plate with series of openings around edge covering umbilical region Cribrobulimina .. C279 [187]
 Later portion uniserial 178
178. Initial portion triangular in section Clavulina .. C279 [187]
 Initial portion rounded in section Cylindroclavulina† .. C281 [188]

179. Reducing to true uniserial development 180
 Reducing to loosely biserial *Plectina* C283 [189]
 Note: Some species may tend to develop a single terminal (uniserial) chamber in gerontic specimens.
180. Short uniserial development; aperture terminal,
 rounded with projecting tooth *Goesella* C281 [188]
 Long uniserial development giving narrow elongate
 test; aperture terminal elongate slit, commonly
 arcuate with bordering lip or short neck *Martinottiella*† C282 [188]
 Note: *Cylindroclavulina*† C281 [188] has three chambers in initial coil.

ATAXOPHRAGMIINAE (C283)

181. Aperture interiomarginal; final stage biserial 183
 Aperture areal; final stage uniserial 182
182. Wall medium to coarsely arenaceous; aperture ir-
 regular or multiple *Liebusella*† C287 [195]
 Wall finely arenaceous with distinct pores; aperture
 circular with ring of teeth *Tritaxilina* C291 [197]

PAVONITINIDAE (C291)

PAVONITININAE (C295)

183. *Textulariella*† C299 [202]

KEY TO RECENT GENERA OF SUBORDER MILIOLINA
 (Loeblich and Tappan 1964 p. C436)

MILIOLACEA (C436)

184. Unilocular 185
 Multilocular 186
185. Single globular chamber; attached Fam. SQUAMULINIDAE 200
 Undivided tubular chamber enrolled for
 most of its length Fam. FISCHERINIDAE 191
186. Coiled throughout or in annular series 187
 Uncoiling or rectilinear series, or, rarely,
 irregular mass 190
187. Chambers tubular to hemispherical,
 elongate in direction of growth 188
 Chambers not tubular in direction of
 growth 189
188. More than two chambers in planispiral or
 low trochospiral coil Fam. FISCHERINIDAE 191
 Two chambers in planispiral, strepto-
 spirial, or milioline coil Fam. MILIOLIDAE 227
189. Axis of coiling short; discoidal or lenti-
 cular Fam. SORITIDAE 261
 Axis of coiling long; sperical to fusiform Fam. ALVEOLINIDAE 274
190. Interior divided into chamberlets Fam. SORITIDAE 261
 Interior of chambers simple Fam. NUBECULARIIDAE 201



FISCHERINIDAE (C438)

191.	Undivided tubular coiled chamber	..	Subfam. CYCLOGYRINAE	..	192
	Series of tubular chambers in coil	..	Subfam. FISCHERININAE	..	195

CYCLOGYRINAE (C438)

192.	Streptospiral; tube may zigzag as it coils	193
	Planispiral; may uncoil in final stage	194
193.	Simple streptospiral coiling; may be one or many changes of axis and plane of coiling	..	<i>Gordiospira</i>	..	C439 [329]
	Tube zigzagging as it coils	..	<i>Meandrospira</i>	..	C440 [333]
194.	Rounded tubular throughout; uncoiling portion, if present, also rounded	..	<i>Cyclogyra</i> †	..	C438 [329]
	Later stages gradually uncoiling to flattened flabeliform portion	..	<i>Cornuspiroides</i> †	..	C438 [329]
	Later stage uncoiling to flattened, irregular, rectilinear, branching and spreading portion	..	<i>Cornuspirella</i> †	..	C438 [329]

Note: *Cornuspiramia* C447 [338] is attached with later branching portion divided by septa.
Tubinella† C477 [365] has straight unbranched uniserial portion.

FISCHERININAE (C441)

195.	Aperture simple open end of chamber, rounded to slit-like	196
	Aperture with bifid tooth, may be modified to rounded plate with central hole	238
	Aperture with broad flat apertural flap	250
	Aperture cribrate	253
196.	Early quinqueloculine stage	237
	Early trochospiral stage	219
	Early streptospiral stage	214
	Early undivided cyclogyrine coil of up to three volutions	197
197.	Adjacent whorls separated by thin plate	214
	Adjacent whorls in contact	198
198.	Low trochospiral	..	<i>Fischerinella</i> †	..	C443 [334]
	Planispiral	199
199.	No lateral laminae; initial whorls visible	..	<i>Fischerina</i> †	..	C441 [334]
	Laminae from each chamber covering entire test and obscuring initial whorls	..	<i>Planispirinella</i> †	..	C443 [334]

SQUAMULINIDAE (C444)

200.		..	<i>Squamulina</i>	..	C444 [337]
------	--	----	-------------------	----	------------

NUBECULARIIDAE (C445)

201.	Chambers coiled throughout	202
	Chambers uncoiling to a rectilinear series possibly branching, or to an irregular mass	203
	Chambers becoming annular	..	Subfam. DISCOSPIRININAE	..	226
202.	Aperture simple open end of chamber	..	Subfam. OPHTHALMIDIINAE	..	209
	Aperture with tooth or apertural flap	..	Subfam. SPIROLOCULININAE	..	215
	Aperture with cribrate trematophore	251

203.	Attached	**	**	**	**	Subfam. NUBECULARIINAE	**	204
	Free living	**	**	**	**	Subfam. NODOBACULARIINAE	**	218

NUBECULARIINAE (C445)

204.	No initial coil	**	**	**	**			205
	Note: Examine initial end very carefully. Specimens may have to be broken, sectioned, or immersed in aniseed oil to reveal initial coil.							
	With initial coil	**	**	**	**			206
205.	Chambers irregular, elongate; without fimbriate keel	**	**	**	**	<i>Calcituba</i> †	**	C446 [338]
	Chambers more regular, inflated; with fimbriate keel and faint transverse “growth lines”	**	**	**	**	<i>Webbina</i>	**	C448 [338]
206.	True uniserial portion after initial coil . .	**	**	**	**			207
	Group of irregular chambers in roughly coiled mass with no true uniserial development . .	**	**	**	**	<i>Nubecularia</i> † . .	**	C445 [338]
	Note: <i>Parrina</i> C478 [365] is not attached. <i>Glomulina</i> C450 [340] is streptospirally coiled.							
207.	Coil of several chambers; uniserial portion not branching	**	**	**	**			208
	Initial coil of single chamber of almost one whorl in length; later in irregular rectilinear series branching at frequent intervals	**	**	**	**	<i>Cornuspiramia</i>	**	C447 [338]
208.	Attached throughout; aperture against attachment; initial coil with arcuate chambers; later chambers short	**	**	**	**	<i>Nubeculopsis</i>	**	C448 [339]
	Attached by initial end only; aperture not against attachment; initial coil with one to three tubular chambers; later elongate or with elongate neck	**	**	**	**			223

OPHTHALMIDIINAE (C448)

209.	Two chambers to the whorl	**	**	**	**			210
	One to numerous chambers in each whorl but not exactly two	**	**	**	**			214
210.	Globular with chambers streptospirally coiled	**	**	**	**	<i>Glomulina</i>	**	C450 [342]
	Flat in planispiral coil with opposed chambers	**	**	**	**			211
211.	Aperture large rounded with large everted lip	**	**	**	**	<i>Weisnerella</i>	**	C452 [340]
	Aperture does not have large everted lip	**	**	**	**			212
212.	Roughly rounded in outline	**	**	**	**	<i>Ophthalmina</i>	**	C450 [342]
	Ovate to fusiform in outline	**	**	**	**			213
213.	Early coil and many succeeding chambers visible . .	**	**	**	**	<i>Ophthalmidium</i> †	**	C448 [340]
	Only two to four elongate chambers visible on either side	**	**	**	**	<i>Edentostomina</i>	**	C448 [341]
214.	Planispirally coiled throughout; rarely more than three chambers in the final whorl	**	**	**	**	<i>Cornuloculina</i> †	**	C448 [340]
	Early streptospiral coil; later planispiral with numerous chambers per whorl	**	**	**	**	<i>Zoyaella</i>	**	C453 [342]

SPIROLOCULININAE (C453)

215.	Planispiral throughout	**	**	**	**			216
	Final stage in milioline coil	**	**	**	**			217
216.	Two opposed chambers per whorl; most of earlier chambers visible; aperture with narrow single or bifid tooth	**	**	**	**	<i>Spiroloculina</i> †	**	C453 [343]
	Note: <i>Riveroina</i> C477 [363] has labyrinthic interior.							

Two or three chambers in final whorl are only ones visible; aperture with apertural flap . . . *Planispironoides*† C453 [344]

Note: *Flintina* C461 [349] has complex tooth. *Nummoloculina*† C468 [355] has laminae which cover early milioline coil. *Planispirinella*† C443 [334] has no apertural flap.

217. Quinqueloculine adult with rounded plate-like tooth which has central hole and ring of apertures around edge . . . *Cribrolinoidea* . . . C453 [343]
 Triloculine adult with apertural flap . . . *Planispirinoides*† . . . C453 [344]

NODOBACULARIINAE (C455)

218. Test compressed; apertural face long and thin . . . 219
 Test not compressed; chambers and apertural face rounded to oval . . . 220
219. Uniserial portion flaring and flabelliform; aperture row of slits in apertural face; early portion planispiral . . . 267
 Test not flaring; aperture elongate slit along length of apertural face; early portion trochospiral . . . *Vertebralina* . . . C456 [346]
220. Later chambers regular, in rectilinear series; single aperture . . . 221
 Later chambers irregular, in irregular uniserial development; typically with more than one aperture in final chamber . . . 259
221. Initial coil of one or several tubular chambers, may be difficult to see . . . 222
Initial coil of many non-tubular chambers . . . 267
222. Chambers pyriform or rounded with tubular neck; some agglutinated material; often attached by initial end . . . 223
 Chambers tubular or rounded without distinct neck; no agglutinated material; never attached . . . 224
223. Aperture rounded without lip . . . *Nodobacularia*† . . . C455 [345]
 Aperture with phialine lip and inwardly projecting teeth . . . *Nubeculina* . . . C456 [346]
224. With well developed initial milioline coil (usually quinqueloculine) . . . 260
 No initial milioline coil; planispiral second chamber of approximately half coil in length . . . 225
225. Chambers distinct; definite sutures and septa; rounded to slit-like, triradiate or cruciform aperture with lip . . . *Nodophthalmidium* . . . C456 [345]
 Chambers indistinct; vestigial septa visible in transmitted light; aperture simple open end of tube . . . 260

DISCOSPIRININAE (C457)

226. With well developed initial cyclogyrine coil . . . *Discospirina* . . . C457 [348]

MILIOLIDAE (C458)

227. No uniserial development . . . 228
 Uniserial development from milioline coil Subfam. TUBINELLINAE . . . 259
228. Interior complex; each chamber divided into chamberlets . . . Subfam. FABULARIINAE . . . 258
 Interior simple . . . 229

229.	Aperture simple open end of tube ..	Subfam. QUINQUELOCULININAE ..	230
	Aperture with bifid tooth or modification of it, e.g., (1) rounded plate with central hole. (2) chevron, triradiate, cruciform, or dendritic slit ..	Subfam. QUINQUELOCULININAE ..	230
	Aperture with apertural flap ..	Subfam. MILIOLINELLINAE ..	246
	Aperture cribrate trematophore ..	Subfam. MILIOLINAE ..	251

QUINQUELOCULININAE (C458)

230.	Exterior porcellaneous calcite ..	231	
	Exterior with arenaceous covering ..	244	
231.	Streptospiral ..	210	
	Planispiral adult ..	232	
	Biloculine or sigmoidal adult ..	242	
	Triloculine adult ..	241	
	Quinqueloculine adult ..	239	
232.	Two chambers, rarely up to two and a half chambers per whorl ..	233	
	Three chambers per whorl ..	238	
233.	With bifid, rarely single tooth ..	234	
	No tooth present ..	236	
234.	Close-coiled; exactly two chambers per whorl ..	235	
	Loose coil of about two and a half chambers per whorl or uncoiling ..	<i>Ptychomiliola</i> .. C465 [353]	
235.	Planispiral throughout ..	216	
	Initial quinqueloculine coil ..	<i>Massilina</i> † .. C462 [350]	
236.	Initial quinqueloculine coil ..	237	
	Planispiral throughout ..	211	
	Note: Specimens may have to be broken, sectioned, or immersed in aniseed oil to reveal the initial coil.		
237.	With network of fine canals in wall ..	<i>Pseudomassilina</i> .. C463 [352]	
	Wall smooth ..	<i>Pateoris</i> † .. C462 [350]	
	Note: This may be a junior synonym of <i>Wellmanella</i> C466 [353].		
238.	Aperture with simple bifid tooth on elongate neck ..	<i>Ptychomiliola</i> .. C465 [353]	
	Apertural plate with central hole developing from early bifid tooth; bar loops over to support plate; no neck ..	<i>Flintina</i> .. C461 [349]	
239.	Apertural plate with central hole developing from early bifid tooth ..	217	
	Aperture with simple bifid tooth ..	240	
240.	Test flattened with final chambers at 180° ..	<i>Massilina</i> † .. C462 [350]	
	Test not flattened; chambers at 144° throughout ..	<i>Quinqueloculina</i> † .. C458 [349]	
241.	Aperture areal, triradiate, cruciform or dendritic ..	<i>Cruciloculina</i> † .. C458 [349]	
	Aperture rounded or elongate with simple or bifid tooth ..	<i>Triloculina</i> † .. C466 [353]	
242.	Internal sigmoidal development; viewed from above, suture is sigmoidal ..	<i>Sigmoilina</i> † .. C465 [353]	
	Biserial throughout or quinqueloculine changing rapidly to biserial, penultimate chamber visible from one side only; keel and suture straight when viewed from above ..	243	
243.	Aperture with bifid tooth or low flat plate with lateral extensions ..	<i>Pyrgo</i> † .. C465 [352]	
	Aperture of areal slits, may be straight, chevron-shaped or almost dendritic ..	<i>Pyrgoella</i> † .. C465 [352]	
	Aperture of radiating slits between curved bars which meet in ring around small central hole ..	255	

244. Sigmoidal development of chambers; small tooth *Sigmoilopsis*† .. C466 [353]
 Quinqueloculine throughout; bifid tooth 245
245. Aperture with crenulate margin .. *Dentostomina* .. C458 [350]
 Aperture with smooth, often porcellaneous margin ..
 and tooth .. *Siphonaperta*† .. C466 [351]
- Note: See Fam. RZEHAKINIDAE Couplet 99.

MILIOLINELLINAE (C466)

246. Two chambers per whorl .. 247
 Three to six chambers per whorl in planispiral coil 250
247. Planispiral and flat; many chambers visible .. 249
 Biloculine; two chambers visible .. *Biloculinella*† .. C467 [355]
 Note: *Pyrgo*† C465 [352] may have “flap” but with slight lateral extensions.
- Triloculine; three chambers visible .. 248
 Quinqueloculine; five chambers visible .. *Scutularis*† .. C468 [356]
 Note: *Cribrolinoides* C453 [343] has apertural plate with central hole and apertures around its edge.
248. With initial cyclogyrine coil .. 217
 With initial quinqueloculine or triloculine coil .. *Miliolinella*† .. C466 [355]
 Note: *Flintina* C461 [349] has flap with central hole and bar curving over to support it.
249. Chambers with internal oblique secondary septa .. 258
 Chambers not labyrinthic .. 216
250. Laminae from each chamber covering most of previously formed exterior of test and obscuring sutures .. *Nummoloculina*† .. C468 [355]
 Chambers without laminae obscuring sutures .. 248

MILIOLINAE (C468)

251. Porcellaneous surface .. 252
 Arenaceous outer layer .. 257
252. Three to six chambers per whorl in planispiral coil 253
 Two chambers per whorl in milioline coil 255
253. Rows of sutural pores and retral processes giving *Elphidium*-like appearance .. *Polysegmentina* .. C472 [359]
 Without sutural pores or retral processes .. 254
254. Compressed; lenticular to discoidal; evolute .. *Hauerina*† .. C470 [357]
 Globular; involute; only three chambers visible .. *Involvohauerina* .. C472 [359]
 Note: *Flintina* C461 [349] has flap with central hole and holes around edge.
255. Test pyriform; aperture as radiate slits between curved bars which meet in ring around small central hole .. *Nevillina* .. C472 [359]
 Test globular, as *Pyrgo*; aperture cribrate trematophore .. 256
256. Interior simple, not divided into chamberlets .. *Cribropyrgo* .. C470 [357]
 Interior labyrinthic .. 258
257. Flat planispiral; early milioline stage; trematophore at peripheral angle .. *Ammomassilina* .. C470 [357]
 Milioline, but chambers added in more than five planes about vertical axis; trematophore in apertural face .. *Schlumbergerina* .. C472 [360]
- Note: See Fam. RZEHAKINIDAE Couplet 99.

FABULARIINAE (C473)

258. Biloculine with cribrate aperture *Fabularia* C473 [362]
 Planispiral with two opposed chambers per whorl;
 aperture areal, terminal, curved slit *Riveroia* C477 [363]

TUBINELLINAE (C477)

259. Later chambers in rectilinear series 260
 Later chambers irregularly uniserial; commonly
 more than one asymmetrically placed aperture *Parrina*† C478 [365]
 Note: *Nubecularia* C445 [338] is attached with early cyclogyrine coil.
260. Vestigial milioline coil; simple proloculus with
 next chamber reversing direction of growth;
 typically lacks heavy longitudinal costae *Tubinella*† C477 [365]
 Well developed milioline coil; usually heavy longi-
 tudinal costae *Articulina* C478 [365]
 Note: *Ptychomiliola* C465 [353] has small aperture with bifid tooth and no everted margin.

SORITIDAE (C482)

261. Chambers (in coiled, rectilinear, or flaring
 growth) divided into chamberlets, or
 many small chambers in annular series 262
 Chambers (in coiled, uniserial rectilinear
 or flaring growth) not divided .. Subfam. PENEROPLINAE 264
262. Uniserial rectilinear throughout .. Subfam. RHAPYDIONININAE 269
 Planispiral or annular; test discoidal 263
 Spherical mass of chamberlets .. Subfam. KERAMOSPHAERINAE 273
263. Small chambers of many shapes which
 alternate more or less regularly with
 those of adjacent series; planispiral
 stage, if any, minute Subfam. SORITINAE 272
 Chambers divided into rectangular cham-
 berlets which do not alternate regularly
 with those of adjacent series; flaring
 planispiral with very curved almost
 spiral sutures; often final annular
 portion Subfam. ARCHAIASINAE 270

PENEROPLINAE (C482)

264. Close-coiled throughout 265
 Uncoiling becoming uniserial 267
265. Aperture open end of tube; possibly simple slit up
 apertural face; no tooth 196
 Aperture with apertural flap 260
 Aperture dendritic slit *Dendritina* C482 [370]
 Aperture cribrate or row of pores or slits 266
266. With rows of sutural pores and retral processes;
 early milioline stage 253
 Without sutural pores, retral processes or early
 milioline stage; may have ridges between sutures *Peneroplis* C482 [369]
 Note: *Archaias* C494 [382] has chambers divided into many rectangular chamberlets and has more swept back sutures.



267. Test rounded in section; aperture single, rounded 268
 Test very compressed and commonly flaring;
 aperture multiple, usually in row or rows .. *Peneroplis* .. C482 [369]
 Note: *Cornuspiroides*† C438 [329] may have growth lines, but no internal septa.
268. Aperture without neck or lip; tooth-like projections
 extending into opening .. *Spirolina* .. C484 [371]
 Aperture with slight neck or phialine lip; no teeth
Monalysidium .. C484 [369]
 Note: *Vertebralina* C456 [346] has early trochospiral coil.

RHAPYDIONININAE (C493)

269. Attached; slightly narrower above basal expansion *Craterites* .. C493 [381]
 Free; conical .. *Ripacubana* .. C493 [380]

ARCHAIASINAE (C494)

270. Planispiral with spiralling septa and sutures; no
 transverse sutures .. *Archaias* .. C494 [382]
 Planispiral early stage, usually becoming very
 flaring, and finally annular; with transverse
 sutures .. 271
271. Initial cyclogyrine coil of several whorls; aperture
 single row of slits around periphery .. 226
 No initial cyclogyrine coil but well developed
Archaias-type planispiral stage with spiralling
 sutures; aperture double row of pores .. *Cyclorbiculina* .. C495 [383]

SORITINAE (C496)

272. Single row of apertures and chambers round peri-
 phery .. *Sorites* .. C496 [385]
 Note: *Discospirina* C457 [348] has initial cyclogyrine coil.
- Double row of apertures and double row of
 alternating chambers from peripheral view .. *Amphisorus* .. C496 [385, 386]
 Numerous minute pores scattered in peripheral
 depression; test several chamberlets thick at
 periphery .. *Marginopora* .. C498 [385]

KARAMOSPHAERINAE (C501)

273. .. *Karamosphaera* .. C501 [390]

ALVEOLINIDAE (C503)

274. Spherical ball of chambers; simple growth outwards
 from central point; irregular to vermiform sutures 273
 Planispiral; elongate axis of coiling, straight or
 gently curved sutures .. 275
275. Spherical to fusiform; single row of apertures .. *Borelis* .. C505 [393, 394]
 Elongate fusiform; several rows of apertures .. *Alveolinella* .. C506 [395]

KEY TO RECENT GENERA OF SUBORDER ROTALIINA (Loeblich and Tappan 1964a, C511) WITH CLASSIFICATION OF SUPERFAMILIES

CLASSIFICATION OF SUPERFAMILIES OF SUBORDER ROTALIINA

I	Test Calcite					
A	Radial wall					
	1 Monolamellar:					
	a. All forms with radiate aperture	Superfam. NODOSARIACEA	343
	b. All forms with apertural tooth or internal tooth plate	Superfam. BULIMINACEA	409
	c. Slit-like, rounded, or cribrate aperture without tooth-plate:					
	i. Planispiral coil, uniserial or single chamber	Superfam. NODOSARIACEA	343
	ii. High trochospiral, biserial (may be enrolled) or uniserial	Superfam. BULIMINACEA	409
	iii. Low trochospiral coil	Superfam. DISCORBACEA	463
	2 Rotaliid septa	Superfam. ROTALIACEA	526
	3 Bilamellar septa:					
	a. Pelagic; typically with globular and/or hispid chambers	Superfam. GLOBIGERINACEA	564
	b. Benthonic; chambers not typically globular or hispid; wall usually thick	Superfam. ORBITOIDACEA	586
B	Granular wall	Superfam. CASSIDULINACEA	646
C	Wall of single crystal or few large crystals	Superfam. SPIRILLINACEA	515
D	Wall of fusiform calcareous spicules	Superfam. CARTERINACEA	723
II	Test aragonite	Superfam. ROBERTINACEA	724

KEY TO FAMILIES OF SUBORDER ROTALIINA

276.	Unilocular	277	
	Multilocular	278
277.	Single rather globular chamber	Fam. GLANDULINIDAE	393
	Non-septate tube in planispiral or low trochospiral coil	Fam. SPIRILLINIDAE	516
278.	Aperture radiate, possibly only as crenulate rim; test glossy	Superfam. NODOSARIACEA	343
	Aperture not radiate	279	
279.	Exterior of single chamber only	Fam. GLANDULINIDAE	393
	Exterior of two chambers only	280	
	Exterior of more than two chambers	281	
280.	Aperture crescentic slit	Fam. NONIONIDAE	676
	Aperture absent or at ends of tubular canals piercing plug	Fam. ROTALIIDAE	529
281.	Uniserial series; may be initial biserial or coiled stage	282	
	Biserial series; may be initial coil	289	
	Coil, spiral, flat sheet or ball of chambers	295	
282.	Single globular chambers <i>irregularly</i> joined to one another, often by elongate stolons	Fam. POLYMORPHINIDAE	382
	Chambers in uniserial rectilinear or curved series	283	
283.	Uniserial throughout	285	
	Early biserial stage	Fam. BOLIVINITIDAE	415
	Early coiled stage	284	

284.	Early planispiral or low trochospiral coil; axis of growth of uniserial stage perpendicular to axis of coiling of initial stage	..	298		
	Early high trochospiral coil; often triserial; axis of growth of uniserial stage continuation of initial axis	330		
285.	Test elongate series of chambers	..	286		
	Test flat cone; chambers appear annular from apex	..		Fam. ANNULOPATELLINIDAE	657
286.	Aperture rounded; often with tooth, lip, or neck	287		
	Aperture slit-like	..	288		
	Aperture cribrate	..		Fam. NODOSARIIDAE	350
287.	Free-living		Fam. EOUVIGERINIDAE ..	432
	Attached	..		Fam. ROTALIIDAE	529
288.	Aperture straight in plane of compression	..		Fam. NODOSARIIDAE	350
	Aperture C- or V-shaped in rounded test slight hood on one side	..		Fam. PLEUROSOMELLIDAE	654
289.	Test elongate; double series of chambers which may be straight enrolled, or twisted; without flattened terminal face of final two chambers	..	290		
	Test conical; flattened umbilical face of final two chambers	..		Fam. SPIRILLINIDAE	516
290.	Early trochospiral stage, soon becoming biserial	291		
	Without early trochospiral stage; may be early enrolled or twisted biserial stage	..	292		
291.	Early low trochospiral coil; biserial stage growing away at tangent to initial coil	..		Fam. CIBICIDIDAE	609
	Early high trochospiral coil; biserial stage grows up axis of initial coil	..	330		
292.	Planispirally enrolled biserial series, at least in early stage; curved axis of growth	..	293		
	Twisted or flat biserial series; straight axis of growth	..	294		
293.	Toothplate extending inwards from aperture to previous foramen; wall radial	..		Fam. ISLANDIELLIDAE	429
	No toothplate visible; wall granular	..		Fam. CASSIDULINIDAE	670
294.	Wall radial; plane of addition of chambers flat throughout	..		Fam. BOLIVINITIDAE	415
	Wall granular; typically with plane of growth twisted at least in early stages	..		Fam. CAUCASINIDAE	658
295.	Chambers in planispiral, streptospiral, or trochospiral coil	296		
	Chambers annular or encrusting or mass of chamberlets growing into ball or twig-like form	..	335		
296.	Chambers in planispirally enrolled biserial series, alternating on either side of plane of coiling; may uncoil	..	293		
	Chambers in simple planispiral, streptospiral or trochospiral coil; may uncoil	..	297		
297.	Surface hispid or finely cancellate; usually hundreds of long thin delicate spines on living specimens; chambers typically globular; pelagic		Superfam. GLOBIGERINACEA	564

	Surface not hispid or finely cancellate and without the many long delicate spines in living specimens; benthonic	298		
298.	Chambers in symmetrical planispiral coil	299		
	Chambers in streptospiral coil; plane of coiling changing	302		
	Chambers in asymmetrical planispiral or trochospiral coil	304		
299.	Chambers divided into chamberlets	..	Fam. NUMMULITIDAE	559
	Chambers not divided	300		
300.	Aperture areal slit in plane of compression or cribrate, at or near peripheral angle	..	Fam. NODOSARIIDAE	350
	Aperture low interiomarginal slit or cribrate at base of, or in, apertural face	301		
301.	Aperture row of pores at base of, or rarely in, apertural face; usually with sutural pores; spiral ridges between raised sutures or solid bars over depressed sutures; wall radial	..	Fam. ELPHIDIIDAE	549
	Aperture low interiomarginal arch; no sutural pores, spiral ridges or sutural bars; wall granular	..	Fam. NONIONIDAE	676
302.	Aperture single	303		
	Aperture multiple	..	Fam. DISCORBIDAE	468
303.	Aperture areal circular on slight neck with lip	..	Fam. SIPHONINIDAE	505
	Aperture interiomarginal arch, with or without tooth	..	Fam. SPHAEROIDINIDAE	414
304.	Test trochospiral, completely evolute spiral side and involute umbilical side; may later uncoil or become annular	310		
	Test asymmetrical planispiral, or trochospiral with partly involute spiral side and/or partly evolute umbilical side	305		
305.	With aperture extending up or isolated in apertural face; may also have interiomarginal arch	306		
	Aperture low interiomarginal arch only, or rarely absent	307		
306.	Test aragonite	..	Fam. ROBERTINIDAE	729
	Test calcite	..	Fam. EPISTOMARIIDAE	509
307.	Series of secondary chambers wedging between primary chambers on one side	..	Fam. AMPHISTEGINIDAE	608
	Without umbilical series of secondary chamberlets	308		
308.	Aperture interiomarginal between periphery and umbilicus	..	Fam. DISCORBIDAE	468
	Aperture interiomarginal at periphery	309		
	Aperture absent or at ends of tubular canals piercing plug	..	Fam. ROTALIIDAE	529
309.	Discoidal; both sides of test flattened with truncate periphery	..	Fam. CIBICIDIDAE	609
	Not discoidal	..	Fam. ANOMALINIDAE	703
310.	Two chambers per whorl	..	Fam. SPIRILLINIDAE	516
	One to many chambers per whorl but not regular arrangement of two per whorl	311		

311.	Ornamentation of radial costae or radial lines of pustules on flattened umbilical surface; commonly two specimens fused by umbilical surfaces . .	Fam. GLABRATELLIDAE	..	502
	No radial lines or pustules on umbilical surface; two specimens rarely fused by umbilical surfaces	312	
312.	High trochospiral coil	..	326	
	Low trochospiral coil	..	313	
313.	Ornamentation of either bars bridging depressed sutures or spiral ridges linking raised sutures	Fam. ELPHIDIIDAE	..	549
	No spiral ridges or bars across chambers	..	314	
314.	Aperture sutural pores only	Fam. DELOSINIDAE	..	669
	Aperture interiomarginal or areal	..	315	
315.	Aperture oval on neck with phialine lip, commonly with fimbriate keel	Fam. SIPHONINIDAE	..	505
	Aperture without neck and phialine lip	316	
316.	Slits or elongate patches of clear shell material in lateral peripheral position	Fam. CERATOBULIMINIDAE	..	725
	Without lateral peripheral slits or elongate patches of clear shell material	..	317	
317.	Six to thirty thick, blunt, longitudinally striated peripheral spines	Fam. CALCARINIDAE	..	543
	No thick blunt peripheral spines	..	318	
318.	Test composed of secreted fusiform calcareous spicules embedded in calcareous ground mass	Fam. CARTERINIDAE	..	723
	Test not of fusiform calcareous spicules	319	
319.	With secondary septa	..	324	
	Without secondary septa	..	320	
320.	With umbilical plug or pillars, or pustulose area in centre of umbilical region	Fam. ROTALIIDAE	..	529
	Without umbilical plug, pillars or pustulose area	321	
321.	With umbilical flaps projecting into umbilicus; umbilical side may be covered by final globose chamber	Fam. DISCORBIDAE	..	468
	Without umbilical flaps at any stage	..	322	
322.	Primary aperture single slit up apertural face; extending from base or areal	Fam. OSANGULARIIDAE	..	698
	Primary aperture low interiomarginal only; may be supplementary apertural pores on spiral or umbilical side	..	323	
323.	Planar or very low domed spiral side and elevated umbilical side	Fam. CIBICIDIDAE	..	609
	Biconvex or elevated spiral side	Fam. EPONIDIDAE	..	596
324.	Test calcite; aperture low interiomarginal arch between periphery and umbilicus	Fam. ASTERIGERINIDAE	..	507
	Test aragonite; aperture umbilical extending as slit up apertural face	..	325	
325.	Aperture extending as slit up apertural face	Fam. ROBERTINIDAE	..	729
	Aperture umbilical arch without slit up apertural face	Fam. CERATOBULIMINIDAE	..	725
326.	Attached at initial end and growing upwards in loose spiral	Fam. ROTALIIDAE	..	529
	Not attached at initial end	..	327	

327.	One to two chambers per whorl through- out	334		
	Triserial, 2½–3½ chambers per whorl; may later become biserial or uniserial	330		
	Four or more chambers per whorl	328		
328.	Sutures raised and granulate ..	Fam. ROTALIIDAE	529	
	Sutures not raised	329		
329.	With secondary septa; test aragonite	Fam. ROBERTINIDAE	729	
	No secondary septa; test calcite	Fam. TURRILINIDAE	413	
330.	Single primary aperture	331		
	Multiple aperture, possible sutural	333		
331.	Aperture rounded on neck or with pro- nounced lip, areal typically terminal ..	Fam. UVIGERINIDAE	451	
	Aperture slit-like, arched, or loop-shaped up apertural face, typically interio- marginal	332		
332.	With internal chamber partitions; test aragonite	Fam. ROBERTINIDAE	729	
	Without internal partitions; test calcite ..	Fam. BULIMINIDAE	437	
333.	Aperture multiple in final chamber	Fam. BULIMINIDAE	437	
	Aperture sutural pores only	Fam. DELOSINIDAE	669	
334.	Aperture loop-shaped up apertural face; test aragonite; chamber arrangement fairly regular	Fam. ROBERTINIDAE	729	
	Aperture ovate, subterminal; test calcite; chamber arrangement very irregular ..	Fam. ISLANDIELLIDAE	429	
335.	Annular or encrusting; growth outwards from central point in two dimensions horizontally; test thin sheet or cone ..	336		
	Growth outwards from central point in three dimensions; each concentric layer of chambers covers whole of unattach- ed area of test; domed to spherical forms	342		
	Growth upwards away from attachment; twig or bryozoan-like forms	Fam. HOMOTREMATIDAE	640	
336.	Flat or conical; only final annulus visible on lower surface ..	337		
	Growth of chambers in flat sheet; similar number of chambers visible above and below	339		
337.	Each annulus only partially divided by radial tubules; growth actually com- pressed uniserial	Fam. ANNULOPATELLINIDAE	657	
	Each annulus consisting of many distinct chambers	338		
338.	Test of secreted fusiform calcareous spicules	Fam. CARTERINIDAE	723	
	Test of granular calcite	Fam. CYMBALOPORIDAE	636	
339.	Each annulus a true tubular circle which may be divided into chamberlets	340		
	Each annulus of distinct contemporary chambers not forming complete or regular circle	341		
340.	Annular chambers undivided; distinct sutural apertures	Fam. CIBICIDIDAE	609	
	Annular chambers divided into chamber- lets; aperture not sutural	Fam. NUMMULITINIDAE	559	

341.	With single central aperture; chambers in ring around it	Fam. HOMOTREMATIDAE	..	640
	With one or two apertures on each of peripheral series of chambers	Fam. PLANORBULINIDAE	..	624
	No opening except mural pores	Fam. ACERVULINIDAE	..	626
342.	With thick blunt spines round periphery of lenticular forms or over whole of spherical types	Fam. CALCARINIDAE	..	543
	Without spines; domed (attached) or spherical	Fam. ACERVULINIDAE	..	626

NODOSARIACEA (C511)

343.	Single chamber only	Fam. GLANDULINIDAE	..	393
	Multilocular	344		
344.	Uniserial throughout, regular or irregular	345		
	Biserial at least in early stages	347		
	Coiled at least in early stages	349		
345.	Irregular uniserial development; may branch, and/or one or more apertures on each chamber	Fam. POLYMORPHINIDAE	..	382
	Regular uniserial development with single aperture	346		
346.	With short entosolenian tube	Fam. GLANDULINIDAE	..	393
	Without entosolenian tube	Fam. NODOSARIIDAE	..	350
347.	Test compressed; periphery angular; uniserial chambers when present are chevron-shaped	Fam. NODOSARIIDAE	..	350
	Test rounded to compressed with well rounded periphery; chambers always globular to ovate	348		
348.	With entosolenian tube	Fam. GLANDULINIDAE	..	393
	Without entosolenian tube	Fam. POLYMORPHINIDAE	..	382
349.	Planispiral coil, rarely low trochospiral; may later uncoil	Fam. NODOSARIIDAE	..	350
	High trochospiral or sigmoidal coil; may later become biserial or uniserial (polymorphine coil)	348		

NODOSARIIDAE (C511)

350.	Biserial at least in early stages	Subfam. PLECTOFRONDICULARIINAE		376
	No biserial stage	351		
351.	Aperture elongate slit only	Subfam. LINGULININAE	..	378
	Aperture rounded, cribrate, or radiate with or without additional small slit	Subfam. NODOSARIINAE	..	352

NODOSARIINAE (C512)

352.	Test of single chamber or final chambers enveloping all previous chambers	353		
	Exterior of many chambers	354		
353.	Unilocular	<i>Lagena</i> †	..	C518 [404]
	Internal uniserial development but last chamber enveloping all previous chambers	<i>Lagenoglandulina</i>	..	C518 [404]
354.	Enrolled throughout	355		
	Uncoiling or rectilinear	356		

355.	With cribrate aperture	<i>Cribrorobulina</i>	..	C516 [405]
	With radiate aperture	<i>Lenticulina</i> †	C518 [406]
	Note: New Zealand forms with an enlarged slit in the apertural face are referred by Eade (1967) to <i>Robulus</i> .						
356.	Uncoiling			357
	Straight or uniformly curved uniserial series; no early coiled stage			365
357.	Initial stage completely enrolled and involute with sudden change to rectilinear uniserial stage, usually with sutures near horizontal			358
	Early stage coiled but not usually tightly enrolled; uncoiling gradually; uniserial stage with sutures sloping down towards proloculus, although may be horizontal sutures between final few chambers			361
358.	Uniserial portion triangular in section	<i>Saracenaria</i> †	C524 [405]
	Uniserial portion rounded in section			359
	Uniserial portion compressed			360
359.	Initial coiled stage rounded periphery; aperture on neck	<i>Dimorphina</i>	C516 [403]
	Initial coiled stage with angular or carinate periphery; aperture not on neck	<i>Marginulinopsis</i> †	C521 [403]
	Note: <i>Vaginulinopsis</i> † C524 [410] has neither initial angular periphery nor apertural neck. <i>Amphicoryna</i> † C513 [401] has both, and initial stage which is not tightly enrolled.						
360.	With chevron-shaped sutures; very compressed	<i>Palmula</i>	C522 [407]
	With straight or slightly curved sutures; ovate in section	<i>Vaginulinopsis</i> †	C524 [410]
361.	Triangular in section	<i>Saracenaria</i> †	C524 [408]
	Rounded in section			362
	Compressed			363
362.	Aperture centrally situated on distinct neck with phialine lip; sutures in later stages horizontal	<i>Amphicoryna</i> †	C513 [401]
	Aperture at peripheral angle and maybe somewhat produced, but without true neck and lip; sutures oblique	<i>Marginulina</i> †	C520 [406]
	Note: <i>Marginulinopsis</i> † C521 [403] has tightly enrolled initial coil.						
363.	With straight or curved sutures			364
	With chevron-shaped sutures	<i>Palmula</i>	C522 [407]
	Note: <i>Plectofrondicularia</i> † C525 [411] has initial biserial stage in at least some members of a population.						
364.	Later chambers ovate in transverse section with rounded periphery	<i>Astacolus</i> †	C514 [401]
	Later chambers thin and flat with angular or carinate periphery	<i>Planularia</i> †	C522 [407]
365.	Aperture radiate			369
	Aperture rounded (beware of damaged specimens), sometimes with teeth			368
	Aperture cribrate or ring of pores (modified radial)			366
366.	Triangular in section			446
	Rounded to quadrangular or compressed in section			367
367.	Aperture three to six pores in circle round central plate formed by fusion of radial ribs of a radiate aperture; microspheric form biserial			377
	Aperture of many randomly arranged pores; microspheric form not biserial	<i>Chrysalogonium</i>	C514 [402]
	Note: <i>Ripacubana</i> C493 [380] has interior divided into chamberlets.						
368.	Aperture circular, smooth, without projecting teeth or internal tooth plate	<i>Orthomorpha</i> †	C522 [407]
	Aperture with indentation or crenulation of lip and internal tooth plate			436

369. Rounded in section throughout 370
Compressed, at least in early stages 374
370. Chambers near spherical; aperture with lip at end
of long cylindrical neck usually with rings around
it; later chambers often separated by stolons;
initial uncoiling stage in microspheric form .. *Amphicoryna*† C513 [401]
Chambers rarely spherical; aperture rarely with lip,
may be produced but not on cylindrical neck;
chamber addition similar throughout in micro-
and megalospheric forms 371
371. Symmetrical; aperture central; sutures horizontal;
test straight 372
Asymmetrical, aperture eccentric and/or sutures
oblique and/or test arcuate 373
372. Chambers, even in early stages, not strongly
embracing *Nodosaria*† C512 [400]
Note: *Entolingulina* C539 [422] has entosolenian tube.
Chambers, at least in early stages, strongly em-
bracing *Pseudonodosaria*† C522 [408]
Note: *Lagenoglandulina* C518 [404] has final chamber completely embracing all earlier chambers.
373. Chambers, at least in early stages, strongly em-
bracing *Pandoglandulina* C522 [408]
Chambers, even in early stages, not strongly
embracing *Dentalina*† C516 [403]
Note: *Phlegeria* Loeblich and Tappan 1963, p. 213 Fig. 2, and *Tomaculoides* Loeblich and Tappan
1963, p. 213 Fig. 2, have short entosolenian tube.
374. Compressed throughout 375
Early chambers only compressed with chevron-
shaped sutures, later chambers rounded in
section *Alfredosilvestris* C512 [401]
Note: *Amphimorphina* C525 [411] has ribs of radial aperture fusing centrally.
375. Aperture eccentric; sutures straight or slightly
curved *Vaginulina*† C524 [410]
Aperture central; sutures chevron-shaped *Frondicularia*† C516 [404]
Note: *Plectofrondicularia* C525 [411] has initial biserial portion in at least some members of a
population.

PLECTOFRONDICULARIINAE (C525)

376. Biserial throughout *Bolivinella* C526 [411]
Uniserial with or without initial biserial stage 337
377. Compressed with angular to carinate margins *Plectofrondicularia*† C525 [411]
Note: Recorded by Eade (1967) as *Proxifrons* a name given by Vella (1963) to the compressed flaring
forms of *Plectofrondicularia*.
Rounded or (rarely) quadrangular in section *Amphimorphina* C525 [411]
Note: *Paradentalina* C533 [417] has a normal radiate aperture, not the ring of pores typical of this
group.

LINGULININAE (C528)

378. Unilocular; aperture slit down one edge *Rimulina* C529 [413]
Multilocular; uniserial at least in later part 379
379. Early stage planispirally enrolled *Lingulopsis* C528 [412]
Uncoiled uniserial throughout 380
380. With keeled margins and very elongate chambers *Mucronina* C528 [414]
Rounded margins; chambers somewhat embracing 381
381. Without entosolenian tube *Lingulina* C528 [412]
With entosolenian tube 396

POLYMORPHINIDAE (C530)

382. Attached ** ** ** ** Subfam. WEBBINELLINAE ** 390
 Test free ** ** ** ** 383
383. Chambers in high trochospiral coil to biserial to regular uniserial series; single aperture; chambers overlapping Subfam. POLYMORPHININAE ** 384
 Chambers in irregular uniserial series commonly branching; commonly several apertures per chamber and/or chambers joined by stolons ** ** Subfam. RAMULININAE . . ** 391

POLYMORPHININAE (C530)

384. Biserial in early stages; may later become uniserial 385
 Sigmoidal in early stages; may later become biserial 386
 Two and a half chambers per whorl (chambers at 144°) in early stages; may later become biserial 388
 Triserial (chambers at 120°) in early stages; may later become biserial ** ** ** ** *Pyrula*† ** ** C533 [416]
 Note: Initial coil form best seen if test is balanced on its aperture in thick gum tragacanth.
385. Biserial becoming uniserial ** ** ** *Paradentalina* . . ** C533 [417]
 Biserial throughout . . ** ** *Polymorphina* . . ** C530 [415]
386. Chambers reach base of test ** ** ** *Sigmoidella*† ** ** C533 [418]
 Chambers overhanging but do not reach base each time ** ** ** 387
387. Sigmoidal throughout *Sigmomorphina*† ** C533 [418]
 Sigmoidal becoming flat biserial *Polymorphina* . . ** C530 [415]
388. Later biserial stage ** ** ** *Pseudopolymorphina*† ** C533 [415]
 Five chambers per whorl throughout ** ** 389
389. Globular or ovate with sutures flush *Globulina* ** ** C530 [416]
 Elongate or ovate with sutures depressed *Guttulina*† ** ** C531 [416]

WEBBINELLINAE (C535)

390. Irregular uniserial growth, attached at initial end 542
 Early polymorphine coil surrounded by flange-like chambers and spreading over substrate ** *Webbinella* ** ** C535 [420]
 Note: *Acervulina*† C694 [564] has few globular chambers with coarse perforations.

RAMULININAE (C537)

391. Chambers irregular and somewhat overlapping ** 392
 Chambers near spherical, joined by stolons; apertures at ends of stolons or long necks ** *Ramulina*† ** ** C537 [420]
 Note: Uniserial members of *Nodosariinae* C512 may have stolons but are regular with only single, usually radiate, aperture. *Siphouvigerina*† C571 [449] has initial coil and internal tooth plate.
392. With many radiate apertures, several per chamber *Sporadogenerina* ** C537 [420]
 With single subterminal aperture ** ** 429

GLANDULINIDAE (C537)

393. Unilocular or final chamber completely embracing all previous chambers ** ** 394
 Chambers in loose high trochospiral coil to biserial or uniserial ** ** Subfam. GLANDULININAE ** 395
394. Unilocular . . ** ** Subfam. OOLININAE ** 403
 Multilocular, final chamber embracing all earlier ones ** ** Subfam. SEABROOKIINAE ** 400

GLANDULININAE (C537)

395.	Uniserial throughout	396		
	Biserial in early stages; may later become uniserial	398		
	Sigmoidal		<i>Laryngosigma</i> †	C539 [421]
	High trochospiral with approximately three chambers per whorl	399		
396.	Test straight, compressed; sutures not oblique		<i>Entolingulina</i>	C539 [422]
	Test arcuate; rounded in section; sutures oblique	397		
397.	Symmetrically added chambers; sutures not overlapping on one side of test; increasing gradually in size		<i>Phlegeria</i> (Loeblich & Tappan 1963 p. 212 Fig. 1)	
	Asymmetrically added chambers; sutures overlapping on one side; test of nearly constant breadth		<i>Tomaculoides</i> (Loeblich & Tappan 1963 p. 213 Fig. 2)	
398.	Biserial becoming uniserial		<i>Glandulina</i> †	C537 [421]
	Biserial throughout		<i>Esosyrinx</i>	C539 [421]
399.	Short free entosolenian tube		<i>Globulotuba</i>	C539 [421]
	Long entosolenian tube attached to inner wall of final chamber		<i>Siphoglobulina</i>	C539 [421]

SEABROOKIINAE (C540)

400.	Compressed ovate		<i>Seabrookia</i>	C540 [425]
	Spherical or nearly so	401		
401.	Aperture rounded; internal uniserial development	353		
	Aperture absent but large pores usually seen	402		
402.	Surface cancellate or with long spines in living specimens; wall thin; internal trochospiral coil	583		
	Surface with beads or pustules of clear shelly material; wall thick; internal group of few chambers	541		
	Surface with many short thick spines; test composed of mass of small chambers	548		

OOLININAE (C540)

403.	No true aperture; many large mural pores; test perfectly spherical	583		
	One or several apertures; test globular to compressed	404		
404.	Aperture rounded or radiate; circular in section	405		
	Aperture slit-like to oval or rounded in fissure-like cavity; test compressed	407		
405.	Two or more stolons protruding from various points around test; asymmetrical	391		
	Single aperture or two symmetrically placed at opposite ends of test	406		
406.	With entosolenian tube; with or without neck		<i>Oolina</i> †	C540 [425]
	Without entosolenian tube; with neck	353		
407.	Aperture slit along periphery	408		
	Aperture subterminal, on one side of test with projecting hood		<i>Parafissurina</i> †	C541 [425]
408.	Aperture at apex; with entosolenian tube		<i>Fissurina</i> †	C540 [425]
	Aperture extending from apex half way down one side; without entosolenian tube	378		



BULIMINACEA (C543)

409.	Uniserial rectilinear series	Fam. EOUVIGERINIDAE ..	432
	Biserial rectilinear series; may become uniserial in later stages	Fam. BOLIVINITIDAE ..	415
	Enrolled, at least in early stages	410	
410.	Planispirally enrolled biserial series	Fam. ISLANDIELLIDAE	429
	Streptospirally enrolled series ..	Fam. SPHAEROIDINIDAE	414
	High trochospiral coil; may become biserial or uniserial	411	
411.	More than three chambers per whorl	Fam. TURRILINIDAE ..	413
	Less than three chambers per whorl	412	
412.	Aperture loop-shaped in apertural face; no neck	Fam. BULIMINIDAE ..	437
	Aperture rounded terminal with neck	Fam. UVIGERINIDAE ..	451

TURRILINIDAE (C543)

TURRILININAE (C543)

413.	Numerous chambers per whorl; chambers very high and narrow; sutures near vertical; aperture loop-shaped in apertural face	<i>Buliminella</i> † ..	C543 [426]
	About five chambers per whorl; chamber not high and narrow; sutures oblique; aperture umbilical	<i>Buliminoides</i> †	C544 [426]

Note: *Angulodiscorbis* C589 [466] has very low broad crescentic chambers; umbilical face perpendicular to axis of growth.

SPHAEROIDINIDAE (C547)

414.	Chambers many; regular trochospiral coil with final chamber covering umbilical area		578
	Chambers few; no early trochospiral coil visible, axis of coiling changing many times	<i>Sphaeroidina</i> †	C547 [432]

Note: *Sphaeroidinella*† C673 [541] has cancellate surface with fimbriate flange around each chamber. *Allomorphina* C743 [611] is involute planispirally coiled with asymmetrical aperture.

BOLIVINITIDAE (C548)

415.	Biserial becoming uniserial		416
	Biserial throughout ..		422
416.	Uniserial chambers divided into many chamberlets		433
	Uniserial portion not divided into chamberlets		417
417.	Aperture single, rounded, or slit-like; internal tooth plate or tube		418
	Aperture absent or multiple; row or ring of pores (modified radial aperture); without internal tooth or tube		421
418.	Aperture subterminal slit with projecting hood on one side ..		655
	Aperture terminal or subterminal, rounded, without projecting hood		419
419.	Wall granular; uniserial stage short with cuneate chambers		667
	Wall radial; uniserial stage well developed with rounded chambers		420

420. Internal twisted tooth plates of successive chambers alternating in position i.e. in planes 180° apart; megalospheric form often uniserial throughout
Internal twisted tooth plates of successive chambers added in plates 120° apart; microspheric form typically triserial in early stages 462 *Rectobolivina*† .. C553 [438]
421. Aperture absent or row of pores across terminal face of final compressed arcuate chamber .. 450
Aperture modified radial aperture; projecting radial bars fusing centrally leaving ring of rounded or elliptical openings 377
422. Aperture areal 427
Aperture interiomarginal, may be indistinct .. 423
423. Aperture slit-like, loop-shaped or high arch in apertural face; without radiating lines of pustules 424
Aperture indistinct; low arch with pustules radiating in lines from apertural area 376
424. Rectangular in transverse section, four angles with well developed keels *Bolivinita*† .. C548 [434]
Not rectangular in section 425
425. Wall granular 665
Wall radial 426
426. Lower margins of chambers frilled and overlapping to give few broad to many small retral processes; test rounded or compressed *Bolivina*† .. C549 [434]
Lower margin of chambers straight or gently curved without crenulations or retral processes; test compressed, commonly keeled *Brizalina*† .. C552 [434, 436]
- Note: *Stainforthia* C561 [442] has large Ω shaped aperture partly closed by tooth plate leaving only slit at one side. *Stainforthia* and *Virgulinopsis* C572 [449] have initial, twisted biserial, or triserial stage.
427. Aperture subterminal slit with projecting hood on one side 655
Aperture terminal or subterminal, rounded, without projecting hood 428
428. Test granular 667
Test radial 454

ISLANDIELLIDAE (C556)

429. Extremely irregular; chambers in smooth-sided high spiraling column; test elongate and slightly arcuate *Orthoplecta* .. C556 [439]
Planispirally enrolled biserial series, later stages may uncoil 430
430. Initially enrolled, later uncoiling 431
Enrolled throughout *Islandiella*† .. C556 [439]
Note: *Cassidulina*† C737 [604] has granular wall.
431. Uncoiling portion rounded or compressed in plane of coiling of initial coil; two sides of test similar shape *Cassidulinoidea*† .. C556 [439]
Uncoiling portion compressed perpendicular to plane of coiling of initial coil; concave and convex sides of very different appearance *Reissia* Loeblich and Tappan 1964b, p. 28.
- Note: *Ehrenbergina*† C738 [604] has granular wall.

EOUVIGERINIDAE (C556)

432. Regular undivided globular chambers in regular uniserial series .. 434
 Chambers divided into chamberlets or irregular chambers in irregular uniserial development .. 433
433. Chambers divided into honeycomb pattern .. *Millettia* ... C558 [441]
 Chambers very irregular; sutures irregular; test smooth arcuate .. 429
434. Uniserial rectilinear throughout; chambers globose typically without longitudinal costae .. 435
 Early coil, biserial stage or eccentric second chamber in at least some members of a population; chambers typically compressed with longitudinal costae .. 420
435. Aperture with circular smooth lip, without internal tooth plate .. 368
 Aperture with indentation or crenulation of phialine lip, and internal tooth plate .. 436
436. Aperture with smooth lip and small tooth or indentation of phialine lip .. *Stilostomella*† ... C559 [440]
 Aperture on slight neck with crenulate or dentate phialine lip and distinct apertural tooth .. *Siphonodosaria* ... C556 [440]
- Note: *Amphicoryna*† C513 [401] does not have internal tooth plate.

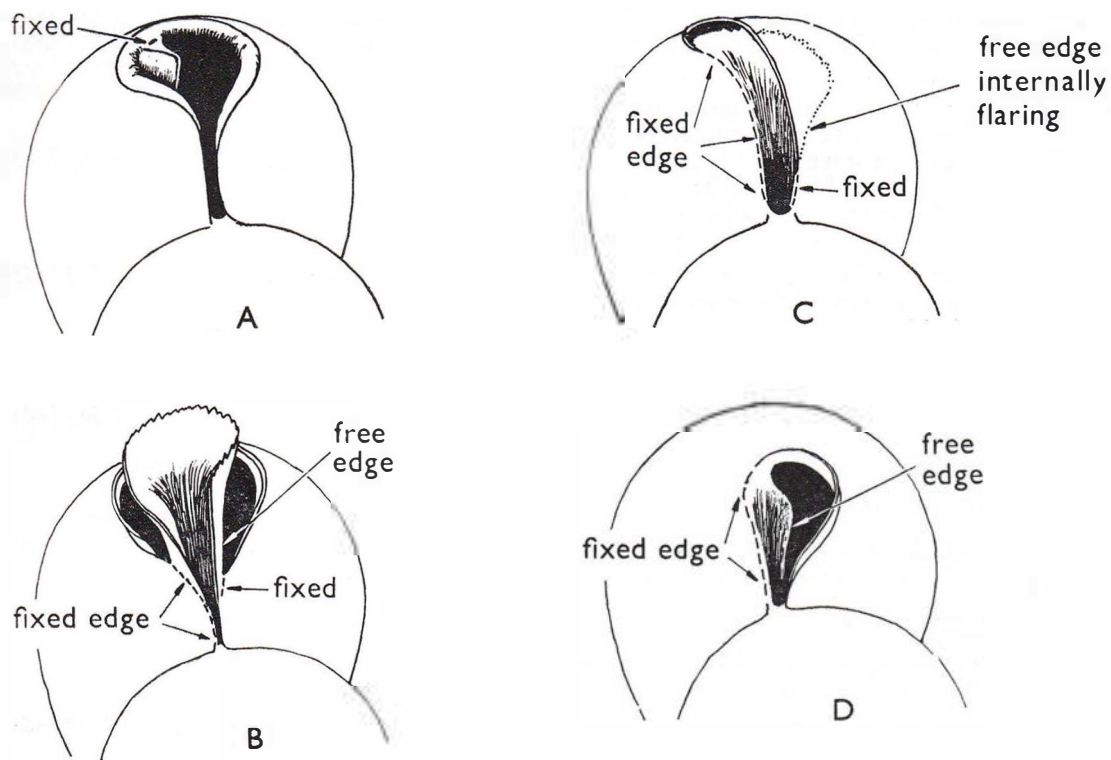
BULIMINIDAE (C559)

437. Aperture single .. 438
 Aperture multiple, or mural pores only.. Subfam. PAVONININAE .. 446
438. Aperture loop-shaped or slit-like up apertural face typically with tooth plate; may be areal, almost terminal .. Subfam. BULIMININAE .. 439
 Aperture low arch, interiomarginal, or areal paralleling base of final chamber Subfam. PAVONININAE .. 446

BULIMININAE (C559)

439. Triserial throughout; final chamber may tend to uniseriality .. 443
 Initially triserial becoming biserial, possibly twisted biserial .. 440
440. Chambers very compressed and cuneate in side view; aperture elongate slit occupying almost whole of distal face .. 449
 Chambers more rounded; aperture slit-like or loop-shaped in apertural face .. 441
441. Aperture more slit-like, commonly with lip; tooth plate distinct and only touching aperture at one point .. 442
 Aperture large \cap shaped with tooth plate passing across and leaving only narrow slit or loop-shaped opening to chamber interior .. *Stainforthia* ... C561 [442]
- Note: *Virgulopsis*† C561 [442] has pustulose surface and is recorded from the Recent of New Zealand.
442. Wall radial .. 456
 Wall granular .. 668





TEXT FIG. 1. Diagrammatic sketches of aperture: A. *Uvigerinella*; B. *Globobulimina*; C. *Praeglobobulimina*; D. *Bulimina*.

- | | | | |
|--------------------|--|-----|---------------------------------------|
| 443. | Internal tooth plate fused to final chamber along whole of one side of loop-shaped or elongate aperture; aperture usually without elevated rim or collar . . . | 444 | |
| | Internal tooth plate fused to final chamber only at single point; open rounded aperture, typically with elevated rim or collar (Text fig. 1A) | 460 | |
| 444. | Curved fan-like portion of tooth plate projecting from aperture which is usually nearly terminal; chambers typically strongly embracing (Text fig. 1B) | | <i>Globobulimina</i> † C559 [442] |
| | No fan-like projection from aperture; tooth plate may flare internally; chambers rarely embracing | 445 | |
| 445. | Aperture elongate extending up toward apex; free edge of tooth plate normally flaring internally and joined to lower edge of the aperture (Text fig. 1C) | | <i>Praeglobobulimina</i> † C561 [442] |
| | Aperture loop-shaped near base of apertural face; free edge of tooth plate not joined to lower margin of aperture (Text fig. 1D) | | <i>Bulimina</i> † C559 [442] |
|
 | | | |
| PAVONININAE (C561) | | | |
| 446. | Triserial throughout | 447 | |
| | Triserial to biserial only | 448 | |
| | Triserial, possibly via biserial, to uniserial | 450 | |
| | Uniserial throughout | | <i>Acostina</i> C563 [444] |

447.	Aperture interiomarginal arch	..	<i>Reussella</i> †	C563 [445]
	Aperture areal slit paralleling basal margin	..	<i>Trimosina</i>	C565 [445]
	Aperture cribrate	..	<i>Fijiella</i>	C563 [444]
448.	Aperture in two parts, basal oval and nearly terminal rounded part; chambers globose, each with a single spine	<i>Mimosina</i>	C563 [445]
	Aperture single slit or arch; chambers usually somewhat compressed with or without spine . .	449				
449.	Aperture elongate slit up one side of compressed cuneate chamber; interior simple	..	<i>Valvobifarina</i>	C565 [445]
	Aperture low arch in flattened chamber; interior labyrinthic	..	183			
450.	Uniserial stage triangular or quadrate in section	..	<i>Chrysalidinella</i>	C563 [444]
	Uniserial stage compressed with low broad arcuate chambers	..	<i>Pavonina</i>	C563 [444]

UVIGERINIDAE (C565)

451.	With well developed biserial stage	..	452			
	No biserial stage, triserial and/or uniserial	..	457			
452.	Initial stage only biserial, later well developed uniserial stage	..	462			
	Mainly biserial, may be initial triserial coil and/or final chamber or two may tend to uniseriality . .	453				
453.	With angular periphery and twisted plane of growth	..	<i>Compressigerina</i>	C565 [446]
	Test well rounded; plane of growth twisted or straight . .	454				
454.	Aperture rounded, terminal	..	455			
	Aperture elongate extending up from base of chamber . .	456				
455.	Aperture well removed from basal margin with phialine lip on slight neck	<i>Hopkinsina</i> †	C567 [446]
	Aperture close to basal margin with lip but without neck	..	<i>Sagrina</i>	C569 [448]
456.	Aperture in imperforate apertural face; penultimate aperture not visible; toothplate narrow, not flaring, part of free edge with fimbriate margin	..	<i>Virgulinopsis</i>	C572 [449]
	No imperforate apertural face; usually outer portion of penultimate aperture also visible; toothplate internally wide and flaring with fimbriate margin	<i>Sagrina</i>	C569 [448]
457.	Triangular in section	<i>Trifarina</i> †	C751 [450]
	Rounded in section	458			
458.	Later chambers loosely attached, with very deep sutures, or may be separated by stolons; apertural neck perforate	..	<i>Siphouvigerina</i> †*	C571 [449]
	Chambers somewhat overlapping throughout; apertural neck, if present not perforate	459				

*New Zealand species referable to *Siphouvigerina* are recorded as *Neouvigerina* by Eade (1967).

459.	Almost completely triserial, may tend to uniseriality in final chamber or two	..	460			
	Predominantly uniserial usually with early coiled or biserial stage	..	462			
460.	Aperture oval to slit-like extending up from base of chamber, may have rim but no neck or phialine lip	..	<i>Uvigerinella</i>	C572 [449]
	Aperture rounded, on neck with phialine lip	..	461			

461.	Straight and narrow tooth plate, base of which is attached to margin of previous foramen	<i>Euvigerina</i> †	C566 [447]
	Tooth plate as above but with broad wing-like projection from one side	<i>Uvigerina</i> †	C565 [446]
462.	Triserial to uniserial	<i>Rectouvigerina</i> †	C569 [448]
	Biserial to uniserial	<i>Siphogenerina</i> †	C569 [449]

Note: *Siphogenerina* has initial triserial stage in microspheric specimens *Rectobolivina*† C553 [438] has tooth plates of successive chambers in planes 180° apart instead of 120° and is uniserial only in megalospheric specimens.

DISCORBACEA (C572)

463.	Aperture interiomarginal				464
	Aperture areal				465
464.	Aperture umbilical with radial grooves or radial rows of pustules on umbilical side only	Fam. GLABRATELLIDAE	502
	Aperture extraumbilical-umbilical				466
465.	With pronounced neck and phialine lip	Fam. SIPHONINIDAE	505
	Without neck or phialine lip				466
466.	Without internal partitions; commonly with umbilical chamber flaps	Fam. DISCORBIDAE	468
	With internal partitions, usually forming ring of chamberlets on umbilical side				467
467.	With rosette of alternating chamberlets	Fam. ASTERIGERINIDAE	507
	Internal partitions confused or dividing off umbilical part of chamber	Fam. EPISTOMARIIDAE	509

DISCORBIDAE (C572)

468.	Coiled test of two to four chambers	Subfam. BAGGININAE	498
	More than four chambers in trochospiral or asymmetrical planispiral coil				469
469.	Without imperforate area above aperture	Subfam. DISCORBINAE	470
	With clear imperforate area above aperture	Subfam. BAGGININAE	498

DISCORBINAE (C572)

470.	With two chambers per whorl, i.e., conical biserial, in later stages				497
	Not regular arrangement of two chambers per whorl				471
471.	With single chamber covering whole of umbilical side at least in megalospheric adult	<i>Tretomphalus</i> †	C585 [459]
	Without chamber covering umbilical side				472
472.	Test minute (usually less than 64 μ); aperture rounded umbilical with umbilical teeth				525
	Adult usually retained in 64 μ (240 mesh) sieve; aperture extraumbilical-umbilical				473
473.	True trochospiral, completely evolute spiral side and involute umbilical side; may have open umbilicus				474
	Trochospiral, somewhat involute on spiral side and/or evolute on umbilical side				491
474.	Biconvex or elevated spiral side				475
	Flattened spiral side, elevated umbilical side				485
475.	Without umbilical plug				476
	With umbilical plug				488

476.	Without supplementary apertures on spiral side ..	477			
	With apertural slits which are sutural or paralleling suture on spiral side ..	489			
477.	Without pustulose or granulate coating of umbilical region ..	478			
	With pustulose or granulate umbilical region ..	490			
478.	Chambers without secondary septa ..	479			
	Chambers with secondary septa ..	509			
479.	Aperture low interiomarginal arch ..	480			
	Aperture slit-like up apertural face ..	<i>Epistominella</i> †	..	C578	[453]
480.	Periphery angular, sometimes carinate ..	481			
	Periphery well rounded ..	501			
481.	Test auricular; imperforate area above aperture ..	501			
	Test rounded; no imperforate area above aperture ..	482			
482.	Chambers not lunate; many to whorl ..	483			
	Chambers lunate with final chamber occupying much of the periphery ..	<i>Neoconorbina</i> †	..	C582	[457]
483.	Without very coarse perforation on umbilical side ..	484			
	Final chamber with very coarse perforation (cribrate aperture) on umbilical side ..	604			
484.	Umbilical flaps coalescing over umbilicus ..	<i>Discorbis</i> †	..	C572	[451]
	Umbilical flaps not coalescing leaving open umbilicus ..	<i>Rosalina</i> †	..	C584	[459]
	Note: <i>Lamarkina</i> † C769 [631] has umbilical aperture and test of aragonite.				
485.	Periphery keeled; wall radial ..	486			
	Periphery not keeled, usually rounded; wall granular ..	694			
486.	Bilamellar; usually coarsely perforate, aperture low interiomarginal arch extending on to high domed umbilical side ..	615			
	Monolamellar; usually finely perforate; aperture peripheral or extending on to spiral side; test usually compressed ..	487			
487.	Aperture low arch on flattened spiral side ..	<i>Planodiscorbis</i> †	..	C583	[458]
	Aperture peripheral on flattened spiral side ..	<i>Discorbinella</i> †*	..	C575	[453]
	*Some New Zealand species referred to <i>Discorbinella</i> by Loeblich and Tappan 1964a are recorded under <i>Discopulvinulina</i> by Eade (1967).				
488.	Without umbilical flaps ..	<i>Gavelinopsis</i> †	C578	[456]
	With umbilical flaps ..	<i>Discorbis</i> †	..	C572	[451]
	Note: <i>Pararotalia</i> C612 [486] has internal "tooth plate" with areal intercameral foramen. <i>Cibicidoides</i> C757 [621] has granular wall.				
489.	Supplementary apertures slit-like, along sutures on both sides of test ..	<i>Helenina</i>	..	C580	[457]
	Apertures on spiral side are areal, close to and paralleling spiral sutures, apertures on umbilical side perpendicular to radial sutures ..	514			
490.	With supplementary sutural apertures near periphery on umbilical side; primary aperture without pustulose lip ..	<i>Buccella</i> †	..	C575	[451]
	Without supplementary sutural apertures; primary aperture with thick pustulose lip ..	603			
491.	With double keel or keel plus angular peripheral ridge; both sides flat to concave ..	495			
	With single keel or angular periphery; planoconvex ..	492			
	Without keel, periphery well rounded; biconvex ..	496			

492.	With broad peripheral keel; high-domed chambers with deep sutures between	<i>Laticarinina</i> †	C580 [457]
	Keel not broad and flat; chambers not high-domed with deep sutures	493	
493.	Convex side completely evolute	<i>Earlmyersia</i>	C577 [455]
	Convex side partly evolute to involute	494	
494.	Umbilical to sutural flaps with alar projections; tips of projections are fused to test near periphery leaving openings in front and behind; umbo on convex side	<i>Lamellodiscorbis</i>	C580 [456]
	Umbilical flaps not modified with alar projections; without umbo on convex side	<i>Discorbinella</i> †*	C575 [453]
*Some New Zealand species referred to <i>Discorbinella</i> by Loeblich and Tappan 1964a are recorded under <i>Discopulvinulina</i> by Eade (1967).			
495.	With double keeled periphery; aperture peripheral ovate opening with lip	<i>Planulinoides</i> †	C584 [458]
	Truncate periphery between marginal keel and unkeeled peripheral ridge; aperture slit-like beneath chamber flaps	<i>Bronnimannia</i> †	C574 [451]
496.	Involute on both sides; aperture peripheral slit up apertural face	<i>Stetsonia</i>	C585 [459]
	Involute umbilical side, partly evolute spiral side; aperture umbilical	500	
497.	With initial undivided spiral coil	522	
	With initial triserial or biserial development	<i>Patellinella</i> †	C582 [457]

BAGGININAE‡(C586)

498.	Two to four subglobular chambers arranged in coil	499	
	More than four chambers; trochospiral	500	
499.	Wall thicker with rough appearance due to many knobs and ridges; aperture interiomarginal arch divided by bars	<i>Rugidia</i>	C587 [462]
	Wall thin and smooth; aperture low interiomarginal arch	<i>Physalidia</i>	C587 [462]
Note: <i>Allomorphina</i> C743 [611] has asymmetrical aperture.			
500.	Without umbilical flaps; partly involute on spiral side	<i>Baggina</i>	C586 [462]
	With umbilical flaps extending over umbilical area; evolute spiral side	501	
501.	Chamber rapidly enlarging, text auricular; commonly with keel	<i>Cancriis</i> †	C586 [462]
	Chambers increasing gradually in size; test and periphery rounded	<i>Valvulineria</i> †	C587 [462]

GLABRATELLIDAE (C587)*

502.	Low trochospiral	503	
	High trochospiral	504	
503.	Aperture large ovate opening in umbilical area of final chamber; limbate sutures on spiral side	<i>Heronallenia</i> †	C589 [464]
	Aperture inconspicuous, restricted to umbilicus; sutures on spiral side depressed	<i>Glabratella</i> †	C588 [464]
Note: Umbilical surfaces are often dissolved during plastogamy so specimens appear to have an oversized umbilical aperture.			

*This family is now placed in the ORBITOIDACEA (Loeblich and Tappan 1964b).

504. With flat umbilical face perpendicular to axis of growth, chambers low *Angulodiscorbis* .. C589 [466]
 With umbilical face at low angle to axis of growth; chambers high so test increases rapidly in size . . 413

SIPHONINIDAE (C591)

505. Test subglobular without fimbriate keel; commonly with random blunt spines or pustules *Siphoninoides* C591 [468]
 Test compressed with fimbriate keel 506
 506. Coiled throughout *Siphonina*† C591 [468]
 Uncoiling in later stages *Siphoninella* C591 [468]

ASTERIGERINIDAE (C592)

507. Rosette of chamberlets wedged between primary chambers around umbilicus; aperture interiomarginal arch only; periphery typically angular 508
 Secondary septae or plates extending from apertural face to septa; aperture interiomarginal, areal or sutural; periphery typically rounded 509
 508. Star-like rosette of diamond or leaf-shaped secondary chamberlets on high-domed umbilical side *Asterigerina* C592 [469]
 Ring of rounded secondary chamberlets on almost flat umbilical side *Asterigerinata* C592 [469]

EPISTOMARIIDAE (C592)

509. Primary aperture in terminal face either extending up from the base, or areal 510
 Primary aperture low interiomarginal slit between umbilicus and periphery, often difficult to see . . 513
 510. Aperture divided by vertical partition into two parallel slits or rows of perforations *Palmerinella* C598 [473]
 Aperture simple slit or rounded 511
 511. Periphery rounded 512
 Periphery with single keel or angular 492
 Periphery with double keel and truncate 495
 512. Without secondary septa; aperture elongate slit extending from base diagonally across apertural face 496
 With secondary septa dividing off umbilical series of chamberlets; aperture loop-shaped in apertural face, or rarely areal *Eponidella* C595 [472]
 Note: *Cerobertina*† C781 [641] and *Ceratobulimina*† 766 [630] have test of aragonite.
 513. With bars of clear shell material bridging sutures . . *Epistomaroides* C594 [473]
 Without bars of shell material bridging sutures . . 514
 514. With supplementary slit-like apertures on spiral side near to, and paralleling spiral sutures; tooth plate, if present, vertically across chamber between spiral and umbilical sides *Pseudoeponides* C598 [474]
 Without supplementary apertures on spiral side, chambers divided by numerous secondary septa projecting inwards from periphery and apertural face *Torrosina* C598 [474]



528.	Planispiral or annular; typically compressed	Fam. NUMMULITIDAE	559
	Trochospiral; not compressed	Fam. ROTALIIDAE	529

ROTAIIDAE (C605)

529.	One to four inflated chambers visible, arranged side by side or embracing previous chambers	Subfam. PEGIDIINAE	541
	Many chambers visible in trochospiral coil, possibly becoming uniserial	530	
530.	Not attached; low to fairly high trochospiral coil with umbilical plug or pustulose area	Subfam. ROTALIINAE	531
	Attached by initial end, later growing upwards in high trochospiral coil; without umbilical plug or pustulose area	Subfam. RUPERTININAE	542

ROTAIINAE (C605)

531.	With three slender spines radiating from test; spines continuous from umbilicus	<i>Asterorotalia</i>	C608 [482]
	Without elongate spines	532	
532.	Test very globose; relatively high trochospiral coil with strongly convex spiral and umbilical sides; chambers well rounded	<i>Asanoina</i>	C607 [481]
	Test not so globose and/or with angular periphery	533	
533.	With areal foramen; commonly chambers pointed or each with short peripheral spine or scattered pustules	<i>Pararotalia</i>	C612 [486]
	Interiomarginal foramen	534	
534.	With double row of sutural canals and pores on both sides of test, may be difficult to see around granular thickening of sutures	<i>Pseudorotalia</i>	C613 [487]
	Without sutural pores or canals on spiral side	535	
535.	With central glassy plug in open umbilicus; plug may be divided by fissures into many pillars, but is continuous to proloculus	536	
	With central pustulose area, pustules or pillars not continuous from one whorl to next	537	
536.	Septa double; periphery typically rounded	<i>Ammonia</i> †	C607 [479]
	Note: <i>Cibicoides</i> C756 [621] has granular wall.		
	Septa single; periphery typically angular	488	
537.	With thickened pustulose apertural lip, pustulose area is visible remains of previous lips, typically well elevated spiral side	603	
	Without thickened pustulose apertural lip	538	
538.	Test granular; perforations in pustulose area lead to cavity beneath	692	
	Test radial; no cavity under pustulose area	539	
539.	Monolamellar septa; finely pustulose or granulate umbilical region; small sutural slits near periphery on umbilical side	490	
	Rotallid septa; umbilical region coarsely pustulose or with pillars or plug; may have sutural slits	540	



540. With single glassy plug often divided by fissures in adult *Ammonia*† C607 [479]
 No early stage with undivided glassy plug; umbilical pillars not continuous from one whorl to the next *Rotalia*† C606 [479, 480]

PEGIDIINAE (C625)*

541. Test sublenticular; two to four chambers in apposition or coil; pustulose side separated from smooth side by broad keel *Pegidia* C625 [498]
 Test globular; final chamber pustulose and strongly embracing *Sphaeridia* C626 [498]
 Note: *Orbulina* C675 [541] has cancellate surface without large pustules.

*This group has been transferred by Loeblich and Tappan (1964b) to the Orbitoidacea as the Family PEGIDIIDAE.

RUPERTININAE* (C627)

542. Aperture interiomarginal slit with lip *Rupertina* C627 [499]
 Aperture rounded, terminal, with lip or neck of non-perforate calcite *Biarritzina*† C628 [499]

*This group has been transferred by Loeblich and Tappan (1964b) to the Orbitoidacea as a subfamily of the CIBICIDAE.

CALCARINIDAE (C628)

543. Lenticular; spines around periphery 544
 Spherical; spines randomly distributed 548
 544. Trochospiral throughout 545
 Early coiled stage; later covering of small chambers in layers over whole test 547
 545. Three long slender smooth spines radiating from proloculus 531
 Three or more spines which are usually thick with striations, not continuing to proloculus 546
 546. Foramen areal, restricted by internal tooth plate; one spine on each chamber 533
 Foramen interiomarginal, without internal tooth plate; spine growth somewhat irregular around periphery *Calcarina* C628 [500]
 547. Early minute planispirally coiled stage *Baculogypsina*† C629 [501]
 Early well developed trochospiral stage *Baculogypsinoidea* C629 [501, 502]
 548. Minute initial coil; later concentric layers of small chambers *Schlumbergerella* C629 [503]
 Well developed trochospiral stage with few concentric layers of small chambers *Baculogypsinoidea* C629 [501, 502]

ELPHIDIIDAE (C631)

549. Trochospiral Subfam. FAUJASININAE 557
 Planispiral and uncoiling Subfam. FAUJASININAE 550
 550. With vertical ridges up apertural face; spiral ridges somewhat irregular; minute anastomosing canals passing through wall and umbilical plug Subfam. FAUJASININAE 557
 Without ridges up apertural faces; spiral ridges, if present, straight; without anastomosing canals in walls, canals in septa and umbilical plug simple Subfam. ELPHIDIINAE 551

ELPHIDIINAE (C631)

551. Whole of apertural face covered with pores;
test porcellaneous 266
With row of pores at base of apertural face possibly
with few additional pores in apertural face 552
552. With retral processes 553
Without retral processes but may have solid bars
crossing sutural depression 555
553. Later stage uncoiling *Ozawaia* C640 [510]
Close coiled throughout 554
554. Septa double in upper half, single in lower half;
umbilical plug, if any, small; sutural pores
single *Elphidium*† C631 [505, 506]
Septa completely double; umbilical plug often very
large, often double row of sutural pores *Cellanthus* C635 [507]
555. With double row of sutural pores; commonly with
grooves extending from pores onto chamber wall
on either side *Elphidiella* C638 [508]
With single row of pores, may be difficult to see 556
556. With single opening or row of pores at base of aper-
tural face plus one or more areal pores in aper-
tural face *Cribroelphidium*† C635 [508]
With basal slit or row of pores; no areal aperture *Cribrononion* C637 [509]
- Note: Forms with granular walls are referred to the Fam. NONIONIDAE, Couplet 676.

FAUJASININAE (C640)

557. Planispiral *Parrellina* C642 [513, 515]
Trochospiral 558
558. With supplementary umbilical chamberlets; aper-
ture low interiormarginal arch, areal slit may be
visible; ornamentation solid pillars across de-
pressed sutures 513
Without supplementary chamberlets; aperture row
of pores near base of apertural face; ornamenta-
tion of spiral ribs between elevated sutures *Polystomellina*† C642 [513]
- Note: Hornibrook (1967) and Eade (1967) prefer to retain the name *Notorotalia* for South Pacific species.

NUMMULITIDAE (C643)

559. Simple planispiral; not divided into
chamberlets Subfam. NUMMULITINAE 560
Annular or flaring planispiral; chambers
divided into chamberlets Subfam. CYCLOCLYPEINAE 561

NUMMULITINAE (C645)

560. *Nummulites*† C645 [518, 519]

CYCLOCLYPEINAE (C647)

561. Flaring planispiral chambers divided into chamber-
lets, possibly final few chambers annular 562
Annular series of chamberlets; initial flaring plani-
spiral stage may rarely be seen 563

562. Test perforate, radial; aperture interiomarginal arch, single row of terminal slits, or absent .. *Heterostegina* C650 [518, 521]
 Test imperforate, porcellaneous; aperture single or double row of pores on peripheral face of last chamber .. 270
563. Test perforate radial; mural pores but no proper apertures .. *Cycloclypeus* .. C647 [518, 521]
 Test imperforate porcellaneous; peripheral apertures .. 272

GLOBIGERINACEA (652)

564. Adult planispiral; aperture equatorial .. Fam. HANTKENINIDAE .. 566
 Adult trochospiral or streptospiral, aperture umbilical to extraumbilical-umbilical; rarely sutural or areal .. 565
565. Aperture umbilical in adult or with areal or sutural pores; may have bulla .. Fam. GLOBIGERINIDAE .. 573
 Aperture extraumbilical-umbilical in adult .. Fam. GLOBOROTALIIDAE .. 569

HANTKENINIDAE (C663)

HASTIGERININAE (C663)

566. Test finely hispid all over .. 567
 Chambers with few broad trigonally prismatic spines concentrated at periphery .. 568
567. Chambers radially elongate .. *Boliella* .. C665 [531]
 Chambers rounded to ovate .. *Hastigerina*† .. C663 [531 No. 1]
 Note: Parker (1962) would prefer to retain the name *Globigerinella* for this group.
568. Chambers rounded to ovate; planispiral throughout .. *Hastigerina*†* C663 [531 No. 2-4]
 Chambers radially elongate; later part streptospirally coiled .. 577

*Bé (1965) thinks that the type species of *Hastigerina* and *Hastigerinella* are merely the adult and gerontic stages of the same species. Parker (1962) "would like to see the combination of *Hastigerina* and *Hastigerinella* into one genus rather than *Hastigerina* and *Globigerinella*." Banner and Blow (1960) are convinced that the types are not conspecific or congeneric, and retain *Hastigerina* and *Hastigerinella* in the same sub-family. Loeblich and Tappan (1964b) refer the two "genera" to different families.

GLOBOROTALIIDAE (C666)

GLOBOROTALIINAE (C666)

569. With angular imperforate periphery, usually with keel .. *Globorotalia*† .. C667 [533, 534]
 With more rounded perforate periphery, no keel .. 570
570. Chamber radially elongate .. 577
 Chambers not radially elongate .. 571
571. With single chamber covering umbilical region .. 584
 Without chamber covering umbilicus .. 572
 Chambers well rounded and/or with apertural lip or teeth .. 581
572. Somewhat compressed spiral side and elevated umbilical side; without apertural lip .. *Turborotalia*† .. C668 [533]
 Note: Eade (1967) includes species of *Turborotalia* in the genus *Globorotalia*.

GLOBIGERINIDAE (C669)

573. With large umbilical (to slightly extra-umbilical) aperture, clearly visible in adult; may be additional sutural apertures .. ** ** Subfam. GLOBIGERININAE .. 576
 Without large umbilical aperture in typical adult; aperture areal or sutural pores, or around edge of umbilical bulla .. 574
574. With umbilical bulla in typical adult .. Subfam. CATAPSYDRACINAE ... 584
 Without umbilical bulla .. 575
575. Test thick; aperture sutural below frilled flange ** ** ** Subfam. SPHAERODINELLINAE .. 582
 Test thin-walled; aperture multiple areal or small sutural .. ** ** Subfam. ORBULININAE .. 583

GLOBIGERININAE (C669)

576. Chambers radially elongate .. ** 577
 Chambers globular .. ** 578
577. Test finely hispid all over .. ** *Beella*† .. C669 [537]
 Test with few broad trigonally prismatic spines concentrated at periphery .. ** *Hastigerinella*†* .. C671 [539]
- *See footnote to couplet 568
578. Without final embracing umbilical chamber; bullae do not have thickened glassy lip .. 579
 Final chamber embracing entire umbilical side; aperture interiomarginal with thickened glassy lip; surface smooth .. ** *Pulleniatina*† .. C671 [539]
 Note: *Tretomphalus*† C585 [459] has cribrate aperture.
579. With secondary sutural apertures on the spiral side *Globigerinoides*† .. C670 [536]
 Without secondary sutural apertures .. 580
580. Aperture umbilical throughout life; no apertural lip or flaps .. ** *Globigerina*† .. C669 [536]
 Aperture extraumbilical-umbilical at least in young stage; commonly with apertural lip or flap .. 581
581. Chambers increasing rapidly in size and strongly inflated; surface coarsely cancellate; apertural lip .. ** *Subbotina* .. C673 [539]
 Chambers not increasing so rapidly in size; surface hispid, smooth, or cancellate; aperture becoming umbilical in adult; commonly with lip or umbilical teeth .. ** *Globorotaloides* .. C671 [540]
 Note: *Globoquadrina* †C670 [539], which keys out here, is regarded as a Recent genus by Eade (1967).

SPHAERODINELLINAE (C673)

582. .. ** *Sphaeroidinella*† .. C673 [541]
 Note: Bé (1965) infers that this form is merely the gerontic stage of *Globigerinoides sacculifera*.

ORBULININAE (C675)

583. Single spherical chamber, usually with internal Globigerina-like coil .. ** *Orbulina*† .. C675 [541]
 High trochospiral coil with small sutural apertures almost completely surrounding later chambers *Candeina*† .. C675 [541]
 Note: *Globigerinoides* C670 [536] has few large sutural openings.



CATAPSYDRACINAE (C676)

584.	Final chamber globose and enveloping entire umbilical side; single aperture with thick glassy lip	**	**	**	**	**	578
	With umbilical bulla or bulla-like extension of final chamber, aperture usually multiple around edge of bulla; without thickened lips	**	**	**	**	**	585
585.	With bulla-like extension of final chamber extending across umbilical region	**	**	**	**	<i>Globigerinita</i> †	C676 [543]
	With true umbilical bulla	**	**	**	**	<i>Tinophodella</i> ..	C678 [543]

Note: Most pelagic species have occasional specimens with a bulla, but *Tinophodella* includes those forms in which the vast majority of adult specimens have a bulla.

ORBITOIDACEA (C678)

586.	Test free; in low trochospiral or planispiral coil, or may be strongly embracing ..	**	**	**	**	587
	Test attached; commonly annular or encrusting or growing upwards from attachment	**	**	**	**	590
587.	Many chambers visible in spiral series ..	**	**	**	**	588
	One or two chambers visible which embrace earlier chambers	**	**	**	**	Subfam. PEGIDIINAE 541
588.	Spiral side flattened ..	**	**	**	**	Fam. CIBICIDIDAE 609
	Test biconvex	**	**	**	**	589
589.	With rosette of chamberlets around umbilicus	**	**	**	**	Fam. AMPHISTEGINIDAE 608
	Without umbilical series of chamberlets ..	**	**	**	**	Fam. EPONIDIDAE 596
590.	Coil or at least initial coiled portion, with flattened spiral side	**	**	**	**	591
	Initial coil, if present, with spiral side not flattened ..	**	**	**	**	592
591.	Low coil not growing away from attachment; with umbilical aperture	**	**	**	**	Fam. HOMOTREMATIDAE 641
	Coiled portion with peripheral aperture, may later spread over substrate or grow upwards	**	**	**	**	Fam. CIBICIDIDAE 609
592.	Encrusting forms	**	**	**	**	593
	Growth upwards from attachment; may branch	**	**	**	**	595
593.	All chambers visible above and below ..	**	**	**	**	594
	Only peripheral chambers visible on lower surface	**	**	**	**	Fam. CYMBALOPORIDAE 636
594.	Apertures at periphery, one or two per chamber, each with lip	**	**	**	**	Fam. PLANORBULINIDAE 624
	No apertures except mural pores	**	**	**	**	Fam. ACERVULINIDAE .. 626
595.	Test multitude of small chambers; several pores per chamber; bryozoan-like forms	**	**	**	**	Fam. HOMOTREMATIDAE 641
	Test of few to several dozen chambers in spiral series, with single aperture	**	**	**	**	Fam. CIBICIDIDAE 609

EPONIDIDAE (C678)

596.	Test attached by umbilical surface; later chambers spreading over substrate	**	**	**	**	607
	Test free	**	**	**	**	597

597.	Test globular with high spiral and umbilical sides, sutures raised and granulate	**	**	532		
	Test not globular with raised granulate sutures	..		598		
598.	Aperture umbilical; test aragonite	**	**	727		
	Aperture equatorial to extraumbilical-umbilical; test calcite	**	**	599		
599.	Aperture interiomarginal at periphery	..		717		
	Aperture between periphery and umbilicus	**	**	600		
600.	Supplementary apertural pores on spiral side or both sides	**	**	606		
	Supplementary apertural pores on umbilical side only	**	**	604		
	Without supplementary apertural pores	**	**	601		
601.	Wall granular	**	**	693		
	Wall radial	..	**	602		
602.	Wall monolamellar	**	**	480		
	Wall bilamellar	**	**	603		
603.	High-domed spiral side; apertural lip much thickened	**	**		<i>Neoeponides</i> †	** C680 [547]
	Lenticular; apertural lip not greatly thickened	**	**		<i>Eponides</i> †	** C678 [544]
	Note: <i>Globorotalia</i> † C667 [533] usually has a cancellate or hispid surface and is pelagic.					
604.	Supplementary areal pores scattered over umbilical surface of final chamber	**	**		<i>Poroeponides</i> ..	** C683 [546]
	Supplementary apertures in row paralleling periphery	**	**	605		
605.	With umbilical pustulose area	..	**	490		
	Without umbilical pustulose area	**	**		<i>Paumotua</i> ..	** C682 [546]
606.	Wall granular; sutures on umbilical side typically sinuate	**	**	695		
	Wall radial; sutures on umbilical side typically almost radial	**	**	514		
607.	Spiral side (upper) pustulose	**	**		<i>Planopulvinulina</i>	** C682 [546]
	Spiral side not pustulose	**	**	638		

AMPHISTEGINIDAE (C685)

608.	Not symmetrical; unequally biconvex and/or chamberlets clearly visible on one side	**	**		<i>Amphistegina</i> ..	** C685 [549]
	Symmetrical; equally biconvex without rosette of chamberlets round umbo	**	**	560		

CIBICIDIDAE (C685)

609.	Partly evolute or evolute on both sides; biplanar or biconcave with truncate periphery	**	**		Subfam. PLANULININAE	** 610
	Flattened spiral side with domed involute umbilical side at least in early stages	..	**		Subfam. CIBICIDINAE ..	** 612

PLANULININAE (C686)

610.	Primary aperture low equatorial interiomarginal arch	**	**	611		
	Primary aperture areal and somewhat oblique, or absent	**	**	495		
611.	Supplementary apertures beneath umbilical flaps on less evolute side	**	**		<i>Planulina</i> †	** C686 [552]
	Apertures beneath umbilical flaps on both sides	..	**		<i>Hyalinea</i>	** C686 [552]
	Note: <i>Paromalina</i> † C763 [627] has perforate periphery and granular wall.					

CIBICIDINAE (C687)

612. Close-coiled throughout 613
 Early coiled stage, later uncoiling or annular .. 621
613. Aperture at periphery, may extend slightly on to
 umbilical and spiral sides 614
 Aperture not peripheral; may reach umbilicus .. 618
614. With supplementary sutural apertures either at
 periphery or on umbilical side 615
 No supplementary sutural apertures at periphery or
 on umbilical side 616
615. With supplementary apertures at proximal margin
 of each chamber on periphery *Caribbeanella* C688 [555]
 With supplementary apertures at proximal margin
 of each chamber near umbilicus giving effect of
 umbilical flaps projecting into umbilicus *Montfortella* (Loeblich and
 Tappan 1963: 213, Figs.
 7-9b)
- Note: *Helenina* C580 [457] has rounded periphery, aperture extending to umbilicus and monolamellar wall.
616. Wall radial 617
 Wall granular 716
617. Wall monolamellar; test compressed 487
 Wall bilamellar; test not typically compressed *Cibicides*† C688 [554]
618. Aperture extending to umbilicus 619
 Aperture not reaching umbilicus 620
619. Wall radial; usually attached by flat spiral side;
 peripheral keel 645
 Wall granular; not attached; usually without keel .. 699
620. Aperture between periphery and umbilicus on
 high-domed umbilical side 694
 Aperture on spiral side; umbilical side low-domed .. 487
621. Later stages biserial, possibly to uniserial 622
 Later stages annular *Cyclocibicides* C690 [554]
 Later stages irregular or spreading network 623
622. Wall granular; typically uniserial but sutures may
 be oblique; finely perforate 704
 Wall radial; more typically biserial or staggered
 uniserial; coarsely perforate *Dyocibicides*† C690 [556]
623. Later chambers closely arranged around initial
 coil; single large aperture per chamber *Cibicidella*† C690 [554]
 Note: *Carpentaria*† C707 [580] has chambers arranged around single central chamber.
 Later chambers in loose network spreading over
 attachment; several apertures per chamber *Planorbulooides* C691 [558]

PLANORBULINIDAE (C692)

624. Later chambers in open network; apertures on
 projecting necks round edge of each chamber
 against attachment 623
 Compact throughout; one or two peripheral
 interiomarginal apertures to each chamber of
 outer series 625
625. Regular annular series; outer row alternating with
 previous ring of chambers *Planorbulinella* C694 [563]
 Irregular series *Planorbulina*† C693 [561]
 Note: *Cibicidella* C690 [554] has large initial *Cibicides*-type coil with few later irregular chambers.

ACERVULINIDAE (C694)

626.	Chambers numerous (hundreds)	627		
	Chambers few (up to several dozen)	633		
627.	Thin encrusting sheet; single layer of chambers, or may irregularly encrust its own early growth	..		628		
	Test of several layers of chambers; usually with one layer alternating with those of row below; typically growing into domed or spherical mass			629		
628.	Chambers globular		<i>Acervulina</i> † C694 [564]
	Chambers vermiform		<i>Planogypsina</i> C698 [568]
	Note: <i>Miniacina</i> † C705 [577] and <i>Sporadotrema</i> C705 [578] (couplet 644) have pores of two types.					
629.	Test spherical	630		
	Test a dome of chambers, possibly irregularly protruding in places	631		
630.	Chambers rounded, rectangular, or polygonal with many pores per chamber		<i>Sphaerogypsina</i> C698 [569]
	Chambers irregular with only single pore at margin of each chamber	273		
631.	Surface with pores of two sizes	644		
	Surface with pores of only one size; commonly with limbate sutures	632		
632.	Interior with layers of chambers		<i>Gypsina</i> † C694 [566, 567]
	Interior with large passages and irregular hollows due to resorption	643		
633.	Initial polymorphine coil with radiate aperture	..		390		
	Initial coil not polymorphine with radiate aperture	..		634		
634.	Initial coil with flattened spiral side as <i>Cibicides</i>	..		623		
	Initial coil not like <i>Cibicides</i>	..		635		
635.	Chambers globose; with deep sutures; surface with many fairly large perforations		<i>Acervulina</i> † C694 [564]
	Chambers not globose or with deep sutures; upper surface pustulose	607		

CYMBALOPORIDAE (C698)

636.	Test conical; chambers alternating in annular series				<i>Cymbaloporetta</i> † C701 [570]
	Test discoidal or flattened; chambers in annular series or irregular	637		
637.	Aperture several series of pores in vertical lines on peripheral face, commonly sutural; initial coil obscured		<i>Cymbaloporella</i> C699 [570]
	Aperture on umbilical side; commonly a series of areal and/or sutural pores; initial coil usually obvious	638		
638.	Aperture series of pores commonly in rows and sutural	639		
	Aperture umbilical with sutural slit or slits	484		
639.	Surface smooth		<i>Pyropilus</i> C702 [575]
	Surface pustulose	607		

HOMOTREMATIDAE (C702)

640.	Trochospiral; attached by spiral side; single central aperture		Subfam. VICTORELLINAE 645
	Growing upwards away from attachment or encrusting; many bryozoan-like forms; aperture multiple small pores		Subfam. HOMOTREMATINAE 641

HOMOTREMATINAE (C702)

641.	With single rounded or slit-like terminal aperture	542		
	With aperture of pores only	642		
642.	Test conical or club-shaped; pores only on broad terminal face	269		
	Test encrusting or twig-like, commonly branching; pores over whole test	643		
643.	Aperture of cribrate plates surrounded by imperforate rims		<i>Homotrema</i>	C702 [577]
	Aperture cribrate without imperforate rims	644		
644.	Two series of pores, one much larger than the other		<i>Miniacina</i> †	C705 [577]
	Pores of single size over most of test, larger pores only at tips of branches		<i>Sporadotrema</i>	C705 [587]

VICTORIELLINAE (C705)

645.			<i>Carpentaria</i> †	C707 [580]
------	--	--	----------------------	------------

CASSIDULINACEA (C725)

646.	Essentially uniserial	647		
	Essentially biserial, includes enrolled twisted and embracing forms	648		
	Essentially trochospiral or planispiral	649		
647.	Test elongate rectilinear series		Fam. PLEUROSOMELLIDAE	654
	Test depressed, conical; chambers appear annular from apex		Fam. ANNULOPATELLINIDAE	657
648.	Chambers alternating at 180°, and embracing all except penultimate chamber		Fam. NONIONIDAE	676
	Elongate biserial series; straight or twisted about elongate axis		Fam. CAUCASINIDAE	658
	Biserial enrolled		Fam. CASSIDULINIDAE	670
649.	High trochospiral with sutural pores only		Fam. DELOSINIDAE	669
	Low trochospiral or planispiral; primary aperture interiomarginal	650		
650.	Monolamellar (these forms are now referred to the Superfam. Nonionacea)*	651		
	Bilamellar (these forms are now referred to the Superfam. Anomalinea)*	653		
*Loeblich and Tappan 1964b.				
651.	Planispiral, may be asymmetrical		Fam. NONIONIDAE	676
	Trochospiral	652		
652.	Less than five chambers per whorl		Fam. NONIONIDAE	676
	Five or more chambers per whorl		Fam. ALABAMINIDAE	692
653.	Aperture equatorial interiomarginal		Fam. ANOMALINIDAE	703
	Aperture either extraumbilical-umbilical, interiomarginal or areal slit		Fam. OSANGULARIIDAE	698

PLEUROSOMELLIDAE (C725)

654.	Aperture loop-shaped in glassy apertural face; test actually high trochospiral with about one chamber per whorl; test radial aragonite	731		
	Aperture curved slit in subterminal position, with hood on one side; true uniserial development; test granular calcite	655		

655. Early biserial or staggered uniserial stage or biserial throughout *Pleurostomella* .. C725 [594]
 No biserial stage; sutures horizontal throughout 656
656. Chambers not greatly overlapping; sutures constricted *Nodosarella* .. C730 [594]
 Chambers strongly overlapping; sutures not greatly constricted *Ellipsoglandulina* .. C728 [595]

ANNULOPATELLINIDAE (C730)

657. *Annulopatellina*† .. C730 [599]

CAUCASINIDAE (C731)

658. Initial stage high trochospiral, later biserial Subfam. CAUCASININAE .. 668
 Initial stage biserial, possibly very twisted, may later remain biserial or become uniserial Subfam. FURSENKONININAE .. 659

FURSENKONININAE (C731)

659. Angular periphery with keel or spines 660
 Rounded periphery without keel or spines 662
660. Aperture terminal ovate; chambers low; without spine 661
 Aperture extending up from base of chamber, possibly divided into two; chambers more cuniform in side view; usually with spines 448
661. Test radial 453
 Test granular *Sigmavirgulina* .. C733 [601]
662. Aperture extending up from base of apertural face; biserial throughout 663
 Aperture areal, terminal or subterminal; may tend to uniseriality 666
663. Aperture large \cap shaped with tooth passing across and leaving only narrow loop or slit to chamber interior 441
 Aperture more slit-like without tooth passing across 664
664. Test granular 665
 Test radial 456
665. Initial portion very twisted biserial (may appear high trochospiral); chambers greater in height than breadth throughout; tooth plate with crenulate margin *Fursenkoina*† .. C731 [600]
 Initial portion not so greatly twisted; early chambers greater in breadth than height; tooth plate with smooth margin *Cassidella* .. C732 [600]
666. Aperture curved subterminal slit with projecting hood on one side 655
 Aperture rounded and terminal or nearly so 667
667. Wall radial; usually thickened apertural lip or neck 455
 Wall granular; apertural lip thin and without neck *Coryphostoma* .. C733 [600]

Note: New Zealand species that key out as *Coryphostoma* are recorded under *Loxostomum* by Eade (1967).

CAUCASININAE (C734)

668. Aperture very elongate slit extending from base across top and half way down opposite side, one margin projecting slightly above other; without apertural tooth plate *Francesita* (Loeblich and Tappan 1963, p. 215, Figs. 3-6)
- Aperture not such an elongate slit, lips of similar height; usually with apertural tooth 659

DELOSINIDAE (C735)

669. Canals from sutural pores lead to spongy cribrate area in apertural face, wall granular *Delosina* C735 [602]
- Without subsutural canals; sutural pores only; wall radial 583

CASSIDULINIDAE (C736)

670. Uncoiling 671
Close-coiled throughout 673
671. Uncoiling portion compressed in plane of coiling of initial coil; two sides of test of similar appearance 431
Uncoiling portion compressed perpendicular to plane of coiling of initial coil; concave and convex sides of very different appearance (test shape reminiscent of crayfish tail) 672
672. Wall radial 431
Wall granular *Ehrenbergina*† C738 [604]
673. Surface smooth 674
Surface ornamented by reticulate ridges *Favocassidulina*† C738 [604]
674. With internal tooth plate; wall radial; aperture elongate interiomarginal partly closed by free edge of tooth plate 430
Without internal tooth plate; wall granular; aperture slit-like up apertural face, usually paralleling distal margin 675
675. Test lenticular; periphery angular *Cassidulina*† C737 [604]
Test globular; periphery broadly rounded *Globocassidulina*† C738 [604]

NONIONIDAE (C742)

676. Total of two to four chambers only 499
Test of more than four chambers, although early ones may be covered in embracing or involutely coiled forms 677
677. Aperture equatorial; planispiral or trochospiral, usually more than four chambers per whorl Subfam. NONIONINAE 680
Aperture extraumbilical-umbilical or umbilical; asymmetrical planispiral or trochospiral, two to four chambers per whorl; forms with two chambers per whorl may appear to be embracing biserial Subfam. CHILOSTOMELLINAE 678



CHILOSTOMELLINAE (C742)

678. Three or four chambers in final whorl; obviously in asymmetrical planispiral or trochospiral coil 679
 Two chambers to whorl; each ovate chamber at 180° to previous chamber and embracing almost all except penultimate chamber *Chilostomella*† .. C742 [611]
 Note: *Globobulimina*† C559 [442] has tooth plate and radial wall.
679. Only final whorl (usually of three chambers) visible externally *Allomorphina* .. C743 [611]
 All chambers visible on spiral side; usually three to four chambers in final whorl *Quadriformina* .. C744 [611]

NONIONINAE (C742)

680. Without final enveloping chamber 681
 Final chamber globose and enveloping most of test; fimbriate margin at base of apertural face *Chilostomellina* .. C746 [613]
 Adult without projection of final chamber over one side only 682
681. Adult with projection of final chamber over umbilicus on one side; opposite side partly evolute *Nonionella*† C748 [613]
 Note: Trochospiral forms lacking well formed umbilical chamber projections are recorded as *Zeaflorius* by Eade (1967).
682. With sutural flaps attached to proximal edge of chamber and partly covering preceding suture and possibly umbilicus 683
 Without sutural and umbilical flaps 684
683. Flaps very large extending across apertural face obscuring aperture *Bisaccium* .. C746 [612]
 Flaps not extending across apertural face *Astrononion*† .. C746 [612]
684. Test aragonite; with areal slit paralleling interior-marginal arch 735
 Test calcite; without aperture in apertural face 685
685. Chambers increasing gradually in size; test fairly rounded in side view 686
 Chambers increase rapidly in size; test flaring; height and breadth of chambers increase approximately threefold or more in final whorl 690
686. Periphery angular test usually large; compressed or lenticular with very large number of chambers per whorl 560
 Periphery well rounded; test size moderate; rarely more than 20 chambers per whorl 687
687. Wall radial; with fine sutural pores, sometimes difficult to see 555
 Wall granular; without sutural pores 688
688. Umbilicus filled with granulate calcite material or solid boss *Nonion*† C746 [612]
 Not with filled umbilicus 689
689. Closed umbilicus; three to six rarely to nine chambers per whorl; wall monolamellar *Pullenia*† C748 [613]
 Deep open umbilicus; usually more than six chambers per whorl; wall bilamellar 715
690. Early trochospiral stage with extension over one side of umbilicus; adult planispiral *Nonionellina* C748 [613]
 Note: The species recorded as *Florilus flemingi* by Eade (1967) is a *Nonionellina* (Lewis and Jenkins, 1969).
 Without early stage with single umbilical projection 691

691. Wall radial; with fine sutural pores, sometimes difficult to see 555
 Wall granular; usually without sutural pores *Florilus*† C746 [612]
 Note: Trochospiral forms recorded as *Zeaflorius* by Eade (1967).

ALABAMINIDAE (C748)

692. Umbilical side not obscured by secondary growth material 693
 Umbilical side covered by plate of secondary shelly material *Trichohyalus* C750 [614]
 Note: *Trichohyalus* is transferred by Loeblich and Tappan 1964b to the Fam. ANOMALINIDAE.
693. Spiral side flattened; umbilical side elevated and umbilicate 694
 Spiral side and umbilical side low-domed 695
694. Short low interiomarginal aperture near midline of apertural face; secondary openings below umbilical flaps *Gyroidina*† C750 [614]
 Long low interiomarginal aperture; extending from periphery to umbilicus 699
695. With small supplementary apertures on spiral side at junction of spiral and radial sutures; periphery carinate *Oridorsalis*† C750 [614]
 Without supplementary apertures on spiral side; periphery sub-angular to rounded 696
696. Aperture not in recessed apertural face, interiomarginal or on umbilical side with lip or flap 679
 Aperture in recessed apertural face formed by projection of final chamber along the periphery, interiomarginal at the base of this face or extending slightly up face 697
697. Finely perforate; surface smooth *Alabamina* C748 [614]
 Coarsely perforate; surface pustulose *Svratkina* C750 [614]

OSANGULARIIDAE (C752)

698. Aperture low interiomarginal arch from periphery to umbilicus 699
 Aperture extending up apertural face or isolated in apertural face 700
699. Without umbilical flaps extending into umbilicus.. . 719
 With umbilical flaps extending over umbilicus *Gyroidinoide*s† C753 [615]
 Note: *Gyroidina*† C750 [614] has aperture confined to middle of apertural face.
700. Test aragonite; aperture umbilical extending as slit up umbilical face of final chamber 734
 Test calcite; aperture extending from extraumbilical position up apertural face or isolated in apertural face 701
701. Chambers with secondary septa giving umbilical series of chamberlets 512
 Without secondary septa 702
702. Aperture elongate slit up apertural face near to and paralleling periphery; wall radial 479
 Aperture partly basal, partly extending at oblique angle up apertural face; wall granular *Osangularia* C752 [615]

ANOMALINIDAE (C753)

703.	With supplementary apertures or strips of clear shell material around peripheral margin	Subfam. ALMAENINAE	..	721
	Without supplementary apertures or strips of clear shell material round peripheral margin	Subfam. ANOMALININAE	..	704

ANOMALININAE (C753)

704.	Coiled throughout			705
	Early coiled portion, later uncoiling to uniserial or irregularly biserial development	<i>Karrerria</i> †	..	C760 [623]
705.	With very broad flat keel, commonly almost sheet-like with growth lines			492
	Keel, if present, not broad and flat			706
706.	With umbilical flaps having apertural openings beneath			707
	Without umbilical flaps, but may have final chambers overhanging umbilicus			711
707.	With umbilical flaps on both sides	<i>Paromalina</i> †	..	C763 [627]
	With umbilical flaps on one side only			708
708.	Wall radial			486
	Wall granular			709
709.	Side with umbilical flaps high-domed convex			694
	Side with umbilical flaps flat or concave			710
710.	Periphery angular; upper side involute	<i>Hanzawaia</i>	..	C759 [623]
	Periphery broadly rounded; upper side partly involute	<i>Discanomalina</i> †	..	C757 [622]
711.	With granulate shell material filling umbilicus and possibly covering whole umbilical side			712
	Without umbilical granulate shell material			713
712.	Granulate shell material confined to umbilicus and possibly extending along sutures			691
	Plate of granulate shell material covering entire umbilical side; hollow beneath granulate plate			692
713.	Completely involute (may have open umbilicus)			714
	At least partly evolute on one side			716
714.	Three ovate chambers per whorl; aperture not equatorial			679
	More than four chambers per whorl; aperture equatorial			715
715.	Periphery angular; umbilici closed; wall radial			608
	Periphery rounded; umbilici open and deep; wall granular	<i>Melonis</i> †	..	C761 [627]
716.	Periphery angular			717
	Periphery rounded			718
717.	Aperture short, interiomarginal at periphery	<i>Cibicidoides</i>	..	C757 [621]
	Aperture elongate interiomarginal from half way to umbilicus, across periphery on to spiral side	<i>Heterolepa</i>	..	C759 [623]
	Aperture extending to umbilicus or not extending to periphery			694
718.	Final chamber overhanging umbilical region on one side			681
	Final chamber not overhanging umbilicus			719

719.	Apertureinteriomarginal at peripherycontinuous on to spiral side	<i>Anomalinoides</i> †	..	C755 [619]
	Aperture not extending on to spiral side	720		
720.	Nearly planispiral with umbo on spiral side and depressed umbilicus	<i>Anomalina</i> †	..	C754 [618]
	Trochospiral with flattened spiral side and high- domed umbilical side	694		
ALMAENINAE (C763)					
721.	Apertures on proximal margins of chambers at periphery	615		
	With lateral peripheral apertures or areas of clear shell material	722		
722.	Involute on both sides; test calcite	<i>Anomalinella</i>	..	C764 [628]
	At least partly evolute on one side; test aragonite	728		

CARTERINACEA (C764)

CARTERINIDAE (C764)

723.	<i>Carterina</i>	C765 [629]
------	------------	------------------	----	----	------------

ROBERTINACEA (C766)

724.	Foramen formed by resorption of the septa; not homologous with aperture; mainly low trochospiral forms	Fam. CERATOBULIMINIDAE	..	725
	Foramen homologous with part of primary aperture; mainly high trochospiral forms	Fam. ROBERTINIDAE	..	729

CERATOBULIMINIDAE (C766)

725.	Aperture umbilical, possibly extending as slit up apertural face	Subfam. CERATOBULIMININAE	..	726
	Apertures or strips of clear shell material in lateral peripheral position	Subfam. EPISTOMININAE	..	728

CERATOBULIMININAE (C766)

726.	Aperture continuing as slit up apertural face	<i>Ceratobulimina</i> †	..	C766 [630]
	Aperture low umbilical arch	727		
727.	Each chamber completely divided into two; division marked by sutures on spiral and umbilical sides; anterior half of chamber non- perforate	<i>Rubratella</i>	..	C770 [633]
	Chambers not completely divided; partial secondary septa do not form sutures at surface; perforate throughout	<i>Lamarckina</i> †	C769 [631]
	Note: <i>Baggina</i> C586 [462] has imperforate area above aperture.				

EPISTOMININAE (C771)

728.	Trochospiral with lateral peripheral aperture on one side only	<i>Hoeglundina</i> †	..	C775 [636]
	Partly evolute on both sides; lateral peripheral apertures or strips of clear shell material on both sides	<i>Mississippina</i> †	..	C776 [637]



ROBERTINIDAE (C777)

729.	High trochospiral to almost uniserial	**	**	731		
	Low trochospiral to planispiral	**	**	730		
730.	Completely evolute spiral side, involute umbilical side	**	**	733		
	Asymmetrical planispiral or planispiral . .	**	**	735		
731.	Single chamber to whorl, parallel sides . .	**	**	<i>Ungulatella</i>	**	C782 [642]
	Several chambers to whorl, test expanding	**	**	732		
732.	Aperture slit up apertural face only	**	**	<i>Robertina</i> †	**	C777 [641]
	Aperture slit up apertural face plus low interiomarginal or sutural slit	**	**	<i>Robertinoides</i> †	**	C782 [642]
733.	Sutures of secondary septa visible on spiral side only or absent; test close-coiled throughout	**	**	734		
	Sutures of secondary septa visible on umbilical side only; test uncoiling	**	**	<i>Cerobertina</i> †	**	C781 [641]
	Note: <i>Eponidella</i> C595 [472] has calcite test.					
734.	Partial secondary septa; chambers not divided by sutures; aperture umbilical extending as slit up apertural face	**	**	726		
	Complete secondary septa with sutures on spiral side; aperture two divergent slits, one up apertural face, one low interiomarginal sutural	**	**	<i>Pseudobulimina</i> †	**	C782 [641]
735.	Symmetrical with areal slit-like aperture parallel to equatorial interiomarginal arch	**	**	<i>Cushmanella</i>	**	C781 [641]
	Asymmetrically developed; aperture areal oblique plus interiomarginal arch	**	**	<i>Alliatina</i>	**	C778 [641]
	Note: <i>Eponidella</i> C595 [472] has calcite test.					

ILLUSTRATED GLOSSARY

agglutinated: composed of foreign particles bound together by cement. (Fig. 1) e.g., *arenaceous* (Fig. 8) *argillaceous* (Fig. 9)

alar projection: wing-like extension. e.g. *alar projections from umbilical flaps* (Fig. 3)

anastomosing tubes: system of intercommunicating tubes (Fig. 2)

angular periphery: sharp or acutely angled outer edge giving clear distinction between two sides of test, possibly with keel. (Figs. 16, 31, 39, 57) (cf. *rounded periphery*)

annular: circular or arranged in rings round central point. (Figs. 4, 5)

apertural face: flattened, usually distal, portion of final chamber which contains main opening to exterior. (Figs. 13, 24, 32A, 46A, 51B, 53C, 58, 74, 75A, 78, 82, 83)

apertural flap: flattened, broadly rounded projection into principal opening to exterior. (Fig. 6)

aperture: major opening(s) of test to exterior, through which protoplasm projects in life. e.g., *single interiomarginal aperture* (Fig. 32A) *multiple interiomarginal aperture* (Fig. 51A) *single areal aperture* (Fig. 77) *multiple areal aperture* (Fig. 24) (*secondary*) *sutural aperture* (Figs. 51A, B; 90)

aposition: placing side by side; usually refers to two chambers or two sets of enveloping chambers. (Fig. 63)

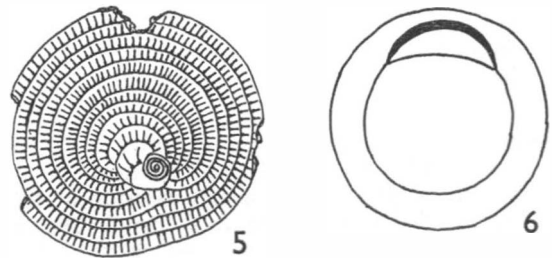
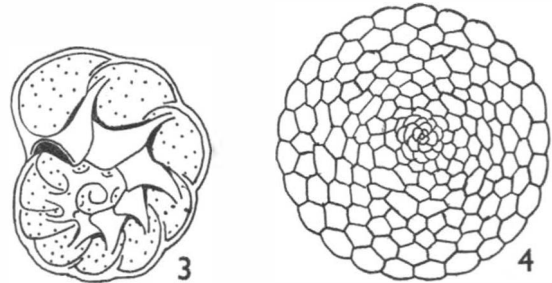
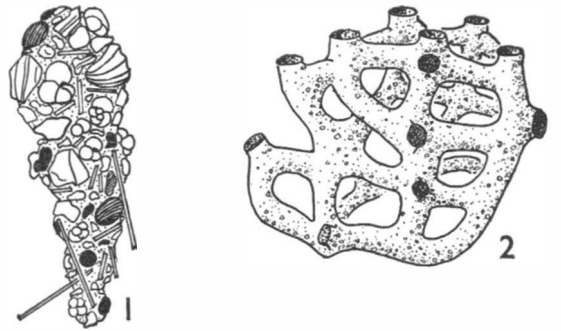
aragonite: metastable orthorhombic form of calcium carbonate. (See Identification Techniques 4, p. 81)

arch: curved or domed opening, typically interiomarginal. (Figs. 3; 32A; 53A, C; 58; 85A)

arcuate: bent, curved. e.g., *arcuate chambers* (Fig. 37) *arcuate test* (Fig. 41) *arcuate aperture* (Fig. 44)

areal: describing opening(s) in chamber wall not bounded at any point by wall of earlier chamber. In some species all that separates opening from earlier chamber is thin lip. e.g., *single areal aperture* (Figs. 8, 25, 77) *multiple areal aperture* (Figs. 24, 82) *areal foramen*. (Fig. 39) (cf. *interiomarginal*)

arenaceous: composed of inorganic sand grade material. (Figs. 8, 54, 91)



1. *Reophax*. 2. *Syringamina*. 3. *Lamellodiscorbis*.
4. *Sorites*. 5. *Discospirina*. 6. *Biloculinella*. 7. *Chilostomella*. 8. *Alveolophragmium*. 9. *Pelosphaera*.

GLOSSARY—continued

argillaceous: composed of silt or clay grade material. e.g., *argillaceous with few large sand grains* (Fig. 9)

asymmetrical planispiral: having chambers coiled in single flat plane, not spiralling. Usually similarly evolute or involute on both sides, but chambers or apertures are not symmetrical about central plane. (Figs. 10A, B, C; 13)

auricular: ear-shaped. e.g., *auricular test* (Fig. 11)

axis: imaginary line around which chambers are arranged or coiled, or along which chambers appear to grow. (Figs. 12, 14, 15) e.g., *axis of coiling* (Fig. 12) *axis of growth* (Figs. 58Y, 87)

axis of coiling: imaginary line around which planispiral or trochospiral forms are coiled. (Figs. 12, 57x)

axis of growth: imaginary line passing symmetrically through test from proloculus to apertural region in biserial and uniserial forms; line along which test appears to grow. (Figs. 57Y, 58Y, 87)

bar: rod of solid shell material forming a link, between two parts of test. e.g., *sutural bars* (Figs. 51A, B)

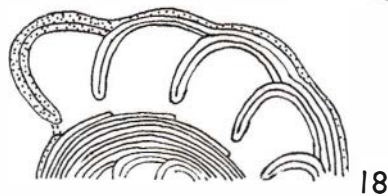
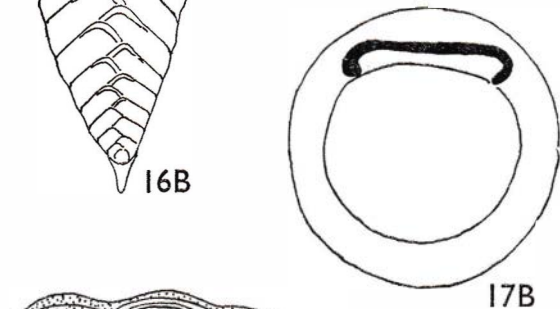
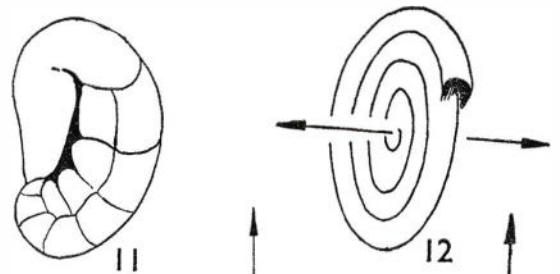
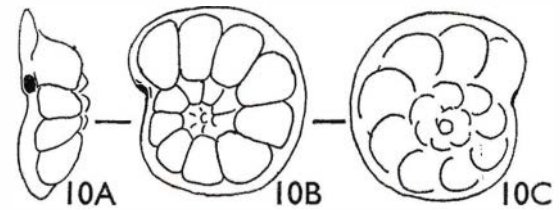
bifid tooth: projection into test opening divided into two minor projections, typically Y-shaped. (Figs. 14; 17A, B; 66c)

bilamellar: having chamber wall primarily of two layers. Outer layer of each chamber covers whole of previously formed exterior of test. Septa do not incorporate canal systems between the two layers, and wall of final chamber consists of two layers. (Fig. 18, equatorial section)

biloculine: milioline, with each chamber added in plane at 180° to previous chamber, “embracing” so that only final two chambers visible. (Figs. 6; 17A, B)

biserial: with two rows of chambers alternating on either side of test, not typically “embracing” (Figs. 16A, B, 58) e.g., *twisted biserial* (Figs. 86, 87) *enrolled biserial* (Figs. 30A, B, C; 31A, B; 57)

bullae: blister-like structure partially or completely covering primary or secondary apertures; independent of primary chambers; typically covers umbilical region; may have one or more accessory interiomarginal apertures. (Fig. 19)



10. *Laticarinina*. 11. *Cerobertina*. 12. *Spirillina* (axis of coiling). 13. *Stetsonia*. 14. *Quinqueloculina* (axis of coiling). 15. *Goesella* (axis of growth and coiling). 16. *Brizalina*. 17. *Pyrgo*. 18. Diagrammatic section through bilamellar test.

GLOSSARY—continued

calcite: stable hexagonal-rhombohedral form of calcium carbonate (see Identification Techniques 4, p. 81)

cancellate: having surface with very fine regular latticework of raised ridges. (Fig. 20)

carinate: having keel or flange. (Figs. 10A, B, C; 46A, B; 59)

chamberlet: subdivision of chamber produced by secondary septa. (Figs. 5, 11)

chevron: Λ -shaped. e.g., *chevron suture* (Fig. 21) *chevron aperture* (Fig. 45A)

close-coiled: completely and tightly enrolled, not uncoiling; may be involute or evolute. (Figs. 20, 22, 26, 46A, B) (cf. *uncoiling*)

closed umbilicus: inner margins of chambers meet at centre of umbilical side and do not leave central depression. (Figs. 22, 55) (cf. *open umbilicus*)

costa: ridge or rib. (Figs. 47, 48)

costate: having ridges. (Figs. 47, 48)

crenulate: having edge finely notched or scalloped. e.g., *crenulate margin of aperture* (Figs. 66A, B, C)

cribrate aperture: multiple areal aperture of rounded holes grouped together. (Figs. 24, 54, 82)

cruciform aperture: opening consisting of four divergent slits; an x-shaped opening. (Fig. 25)

cunifform: wedge-shaped. e.g., *cunifform chambers* (Figs. 27, 45B)

cyclogyrine: having undivided tubular planispiral test. (Figs. 12, 26)

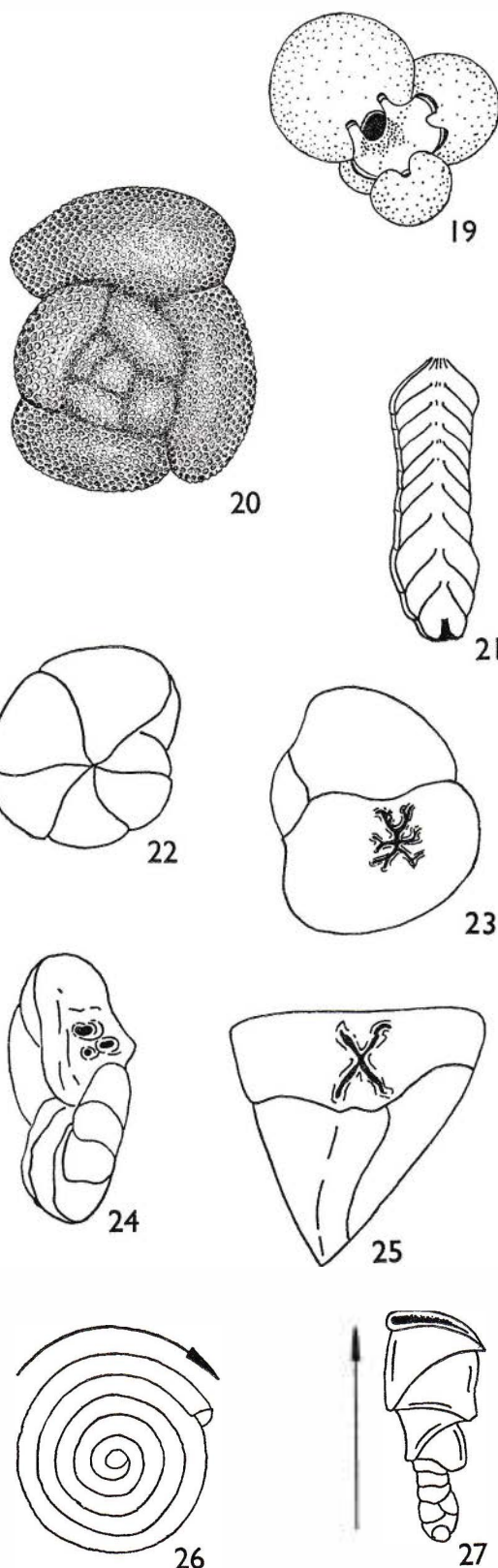
dendritic aperture: opening or openings which branch irregularly. (Fig. 23)

dichotomous branching: repeated division into two. (Fig. 28)

discoidal: round and flat, like a coin. (Figs. 85A, B)

distal: direction away from proloculus in direction of growth. (Figs. 26, 27)

embracing: with later chambers partly covering or completely surrounding earlier chambers. e.g., *partly embracing* (Figs. 7, 79) *completely embracing* (Fig. 29)



19. *Tinophodella*. 20. *Globoquadrina* (del. N. de B. Hornibrook). 21. *Frondicularia*. 22. *Alabama*. 23. *Cruciloculina*. 24. *Jadammina*. 25. *Cruciloculina*. 26. *Cyclogyra* (arrow pointing distally). 27. *Valvobifarina* (arrow pointing distally).

GLOSSARY—continued

enrolled biserial: with two rows of chambers planispirally enrolled. Chambers alternate on opposite sides of test giving Y-shaped sutures when viewed from either side. Test may be uncoiling to give a biserial form with curved axis of growth. (Figs. 31A, B) e.g., *uncoiling biserial, compressed in plane of coiling* (Fig. 57) *uncoiling biserial, compressed perpendicular to plane of coiling* (Figs. 30A, B, C)

entosolenian tube: internal tube-like extension from aperture. (Figs. 33, 64)

equatorial aperture: symmetrical opening of planispiral test, may be interiomarginal or areal. (Figs. 8, 32B)

evolute: with each whorl of enrolled forms not embracing earlier whorls, all whorls visible. (Fig. 26) e.g., *completely evolute spiral side of trochospiral forms* (Figs. 20, 50A, 53B, 75C) *partly evolute* (Figs. 3, 61) (cf. *involute*)

extraumbilical aperture: opening in final chamber not extending to umbilicus, commonly interiomarginal midway between periphery and umbilicus. (Figs. 53A, C, 59, 90)

extraumbilical-umbilical aperture: interiomarginal opening in final chamber that extends from umbilicus to point outside umbilical region, possibly to periphery. (Fig. 34)

fimbriate: having fringe-like margin. (Figs. 35A, B)

fistulose chamber: final irregular chamber with tubular extensions commonly seen in Polymorphinidae and Glandulinidae. (Fig. 36)

fixed edge: side of tooth plate fixed to apertural lip or border. (Text figs. 1b, c, d, p. 43) (cf. *free edge*)

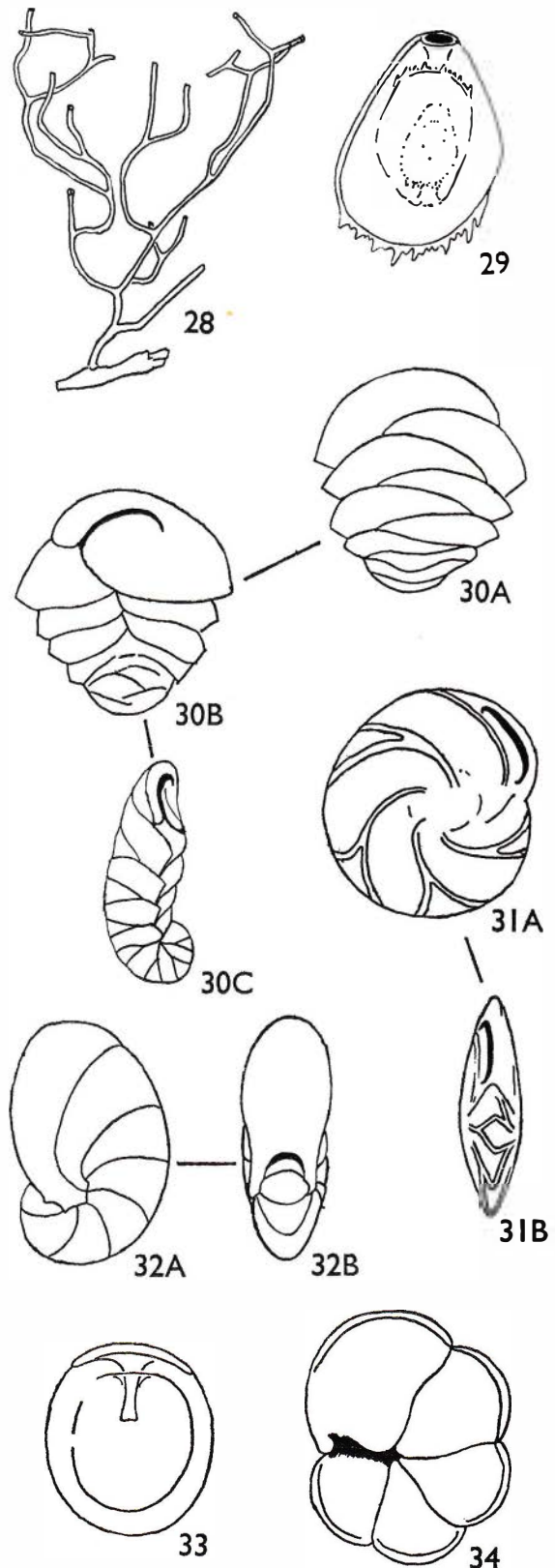
flabelliform: shaped like a fan. (Figs. 37, 38)

flap: flattened projection from chamber wall. e.g., *apertural flap* (Fig. 6) *umbilical flap* (Fig. 53c) *sutural flap* (Fig. 81)

flaring: rapidly increasing in size. Usually refers to test, i.e. each chamber is considerably larger than previous one. (Figs. 11, 32B) e.g., *flaring with final arcuate chambers* (Fig. 37)

flask-shaped: rounded with long narrow neck. (Fig. 40)

foramen: opening in septum between two chambers; may represent a previous aperture or be formed secondarily. (Fig. 39)



28. *Dendrophrya*. 29. *Seabrookia*. 30. *Ehrenbergina*.
31. *Cassidulina*. 32. *Nonionellina*. 33. *Fissurina*.
34. *Globorotalia*.

free edge: side of tooth plate not fused to apertural lip or border. In rare instances free edge may be fused at one point. (Text fig. 1B, C, D, p. 43) (cf. *fixed edge*)

fusiform: spindle-shaped, tapering at each end. (Fig. 42)

gerontic: pertaining to senile stage in life history. Often growth form is different from that in ordinary adult giving whole test very different appearance.

granular: having test composed of randomly orientated minute crystals of calcite; seen between crossed Nicols as a multitude of tiny flecks of colour (see *Identification Techniques 3*, p. 81)

granulate: with small prominences; having roughened surface as if sprinkled with grains. (Figs. 85A, B)

high trochospiral: spirally coiled, height of spire being greater than breadth of terminal face. (Figs. 78, 83.) e.g., *high trochospiral with embracing chambers* (Fig. 79)

hispid: covered with fine hair-like spines. (Fig. 41)

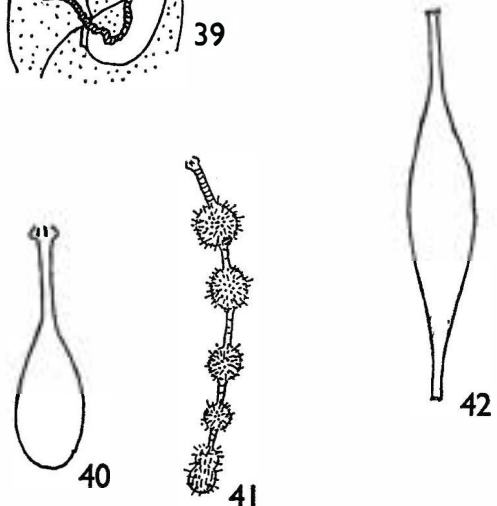
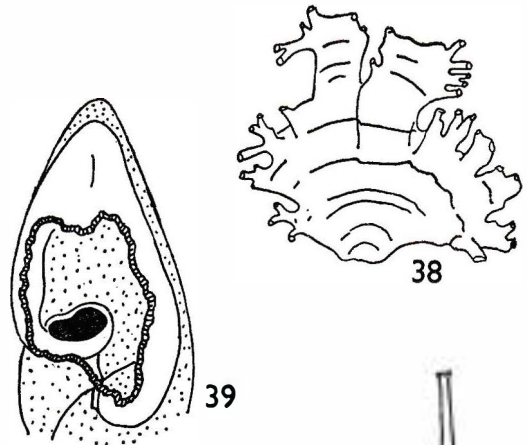
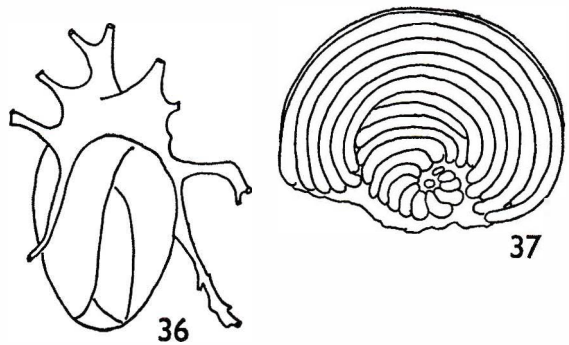
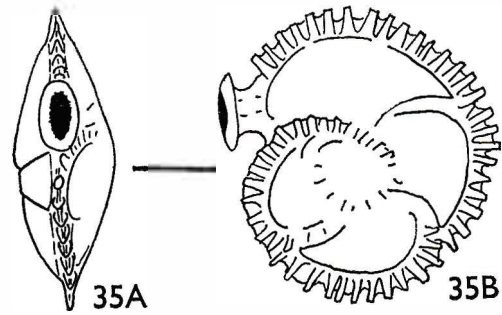
hood: terminal or marginal portion of chamber projecting above aperture. (Figs. 45A, B)

hyaline: colourless, glassy; in the strict sense, transparent, but term usually extended to all transparent or translucent forms which appear colourless, white, or pale blue in transmitted light (see *Identification Techniques 3*, p. 81) (cf. *porcellaneous*)

interiomarginal aperture: opening in final chamber touching margin of that chamber so that part of opening is bounded by wall of earlier chamber. e.g., *loop-shaped interiomarginal* (Figs. 78, 79, 83) *extraumbilical interiomarginal* (Figs. 53A, c; 59; 89) *extraumbilical-umbilical* (Fig. 34) *umbilical* (Fig. 88) *multiple interiomarginal* (Figs. 51A, B; 75A) (cf. *areal aperture*)

involute: each whorl of enrolled forms completely embracing earlier whorls; only final whorl visible. (Figs. 32A, B; 46A, B) e.g., *completely involute umbilical side of trochospiral forms* (Figs. 22, 34, 50B, 53C, 59, 75B) *partly involute* (Figs. 3, 61) (cf. *evolute*)

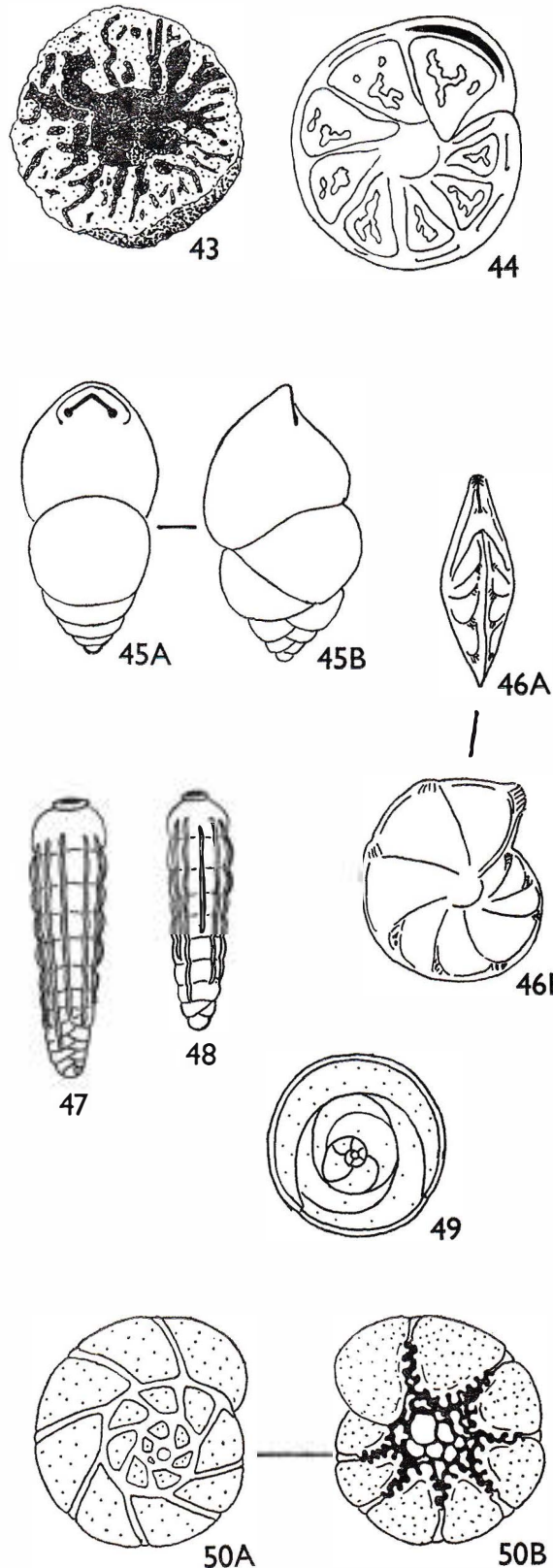
keel: peripheral shell thickening. (Figs. 10A, B, C; 46A, B; 59)



35. *Siphonina*. 36. *Sigmomorphina*. 37. *Peneroplis*. 38. *Julienella*. 39. *Pararotalia* (final chamber broken). 40. *Lagena*. 41. *Amphicoryna*. 42. *Lagena*.

GLOSSARY—continued

- labyrinthic:** having complex spongy wall with interlaced dendritic channels perpendicular to test surface. (Fig. 43)
- lamella** or **lamina:** thin plate-like layer of aragonite or calcite which partly or completely covers test, one being formed with addition of each new chamber. (Figs. 18; 52; 69B, C; 70)
- lateral peripheral:** near to and paralleling periphery e.g., *lateral peripheral aperture* (Fig. 44)
- lenticular:** lens-shaped, approximately equally bi-convex with angular periphery. (Figs. 46A, B)
- limbate suture:** thickened or elevated line of union between two chambers or two whorls. (Figs. 31A, B; 50A)
- lip:** thickened, elevated, or in some way differentiated, border to aperture. (Figs. 7, 8, 27, 47, 48, 77, 80, 85A, 90) e.g., *phialine lip* (Figs. 35A, B)
- loop-shaped aperture:** ovate or comma-shaped opening in apertural face usually with narrow end extending towards base or umbilical region of apertural face. (Figs. 78, 83)
- low trochospiral:** spirally coiled with one side (spiral side) more evolute than other (umbilical side), height of spire being less than breadth of final whorl. (Figs. 50A, B; 53A, B, C; 75A, B, C)
- lunate:** crescent-shaped. e.g., *lunate chambers* (Fig. 49)
- megalospheric:** having large initial chamber. Commonly represents generation formed by asexual reproduction and usually undergoes sexual reproduction. (Fig. 48) (cf. *microspheric*)
- microspheric:** having small initial chamber. Commonly represents generation formed by sexual reproduction and which undergoes asexual reproduction. (Fig. 47) (cf. *megalospheric*)
- milioline:** with elongate tubular chambers (may be sausage-shaped to hemispherical in external appearance), each half a whorl in length, arranged in various planes from 120° to 180° apart about central axis. This is a rather vague term, but is generally used to exclude planispiral evolute coiling (e.g., Figs. 54, 82), although the elongate tubular chambers may still be described as milioline. e.g. *biloculine* (Figs. 6; 17A, B) *triloculine* (Figs. 23; 25; 84A, B, C) *quinqueloculine* (Figs. 14; 66A, B, C)



43. *Oryctoderma*. 44. *Hoeglundina*. 45. *Pleurostomella*.
46. *Lenticulina*. 47. *Rectobolivina* (microspheric form).
48. *Rectobolivina* (megalospheric form). 49. *Neconorbina*. 50. *Ammonia*.

monolamellar: having chamber primarily of single layer which continues as thin lamella to cover whole exterior of test; septa composed of single layer throughout. (Fig. 52)

multilocular: test of many chambers. (Figs. 51A, B; 52; 53A, B, C; 54; 55; 56) (cf. *unilocular*)

multiple aperture: two or more openings of similar type constituting major opening to exterior. e.g., *multiple areal* (Figs. 24; 51A, B; 54; 82) *multiple interiomarginal at base of apertural face* (Figs. 51A, B; 75A) *multiple sutural* (Figs. 51A, B; 69A)

mural pores: minute openings, general over test of many foraminifers; have pseudochitinous linings and sieve plates seen only by electron microscope. (Indicated as dots in Figs. 3; 50A, B; 68; 90)

neck: narrow tubular portion connecting aperture with chamber cavity. (Figs. 35A, B, 40, 41, 42)

open umbilicus: central depression, often continuous to proloculus, between inner margins of chambers in involute coil; previous whorls not visible in depression. e.g., *open umbilicus with umbilical plug* (Figs. 50B, 59) *open umbilicus with umbilical flaps* (Fig. 53c)

ovate: egg-shaped, usually in outline only. e.g., *ovate chambers* (Fig. 61)

oviform: three-dimensional egg-shaped body. e.g., *oviform chambers* (Fig. 56)

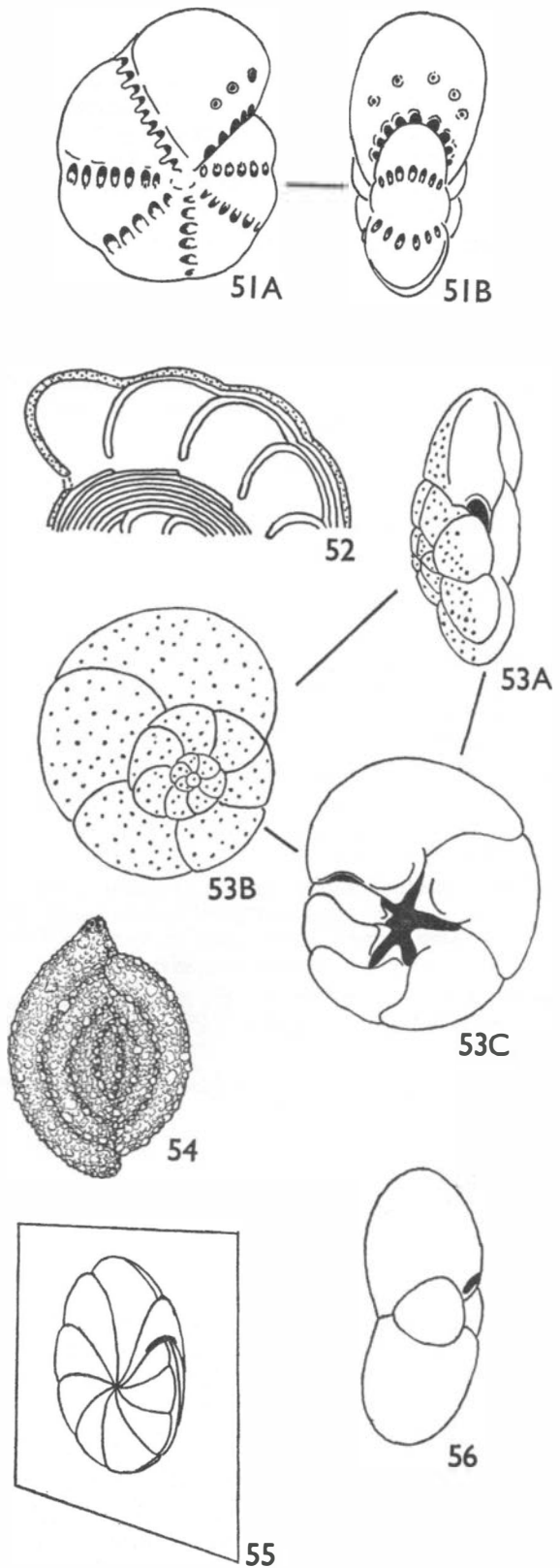
peripheral angle: region where periphery meets terminal or apertural face in coiled forms. e.g., *aperture at peripheral angle* (Figs. 46A, B; 54)

periphery: outer edge as seen when specimen is lying in its most stable position. e.g., *angular periphery* (Figs. 16; 31A, B; 39; 57) *rounded periphery* (Figs. 32A, 51B, 58) *periphery with keel* (Figs. 10A, B, C; 46A, B) *aperture at periphery* (Figs. 33, 54, 61, 85A)

phialine lip: everted rim of aperture, commonly on neck. (Figs. 35A, B)

pillar: column of solid shell material, commonly formed by subdivision of umbilical plug. (Fig. 50B)

plane of coiling: imaginary plane surface through coiled or spiral forms perpendicular to axis of coiling. (Figs. 55, 57a)



51. *Criboelphidium*. 52. Diagrammatic section through monolamellar test. 53. *Rosalina*. 54. *Ammomassilina*. 55. *Anomalinella* (plane of coiling). 56. *Cystamina*.

plane of growth: imaginary plane surface symmetrically through two rows of chambers of biserial forms; in some genera may be twisted or curved. e.g., *flat plane of growth* (Fig. 58) *twisted plane of growth* (Fig. 87) *curved plane of growth* (Figs. 57b; 30A, B, C)

planispiral: coiled in single plane; similarly involute or evolute on both sides. (Figs 12; 32A, B; 46A, B; 54; 85A, B) see also *asymmetrical planispiral* (Figs. 10A, B, C) *planispirally enrolled biserial* (Figs. 31A, B)

plastogamy: fusion of adults by umbilical surfaces at time of sexual reproduction ensuring fertilisation of gametes. (Fig. 60)

plug: filling of solid shell material: e.g., *umbilical plug* (Figs. 50B, 59)

polymorphine coil: high trochospiral to biserial or sigmoidal series; chambers overlapping; sutures at large angle to horizontal; aperture terminal. (Figs. 64; 71A, B) e.g., *polymorphine coil with final fistulose chamber* (Fig. 36)

porcellaneous: white, opaque, or slightly translucent in reflected light; fragments usually appear amber to chestnut in transmitted light (see Identification Techniques 3, p. 81) (cf. *hyaline*)

pore: minute rounded opening in chamber wall. e.g., *sutural pores* (Figs. 51A, B; 69A) *areal pores* (Figs. 51A, B; 82) *mural pores* (Figs. 3; 50A, B; 68; 90)

primary aperture: main opening of test to exterior. e.g., *primary interiomarginal aperture* (Fig. 90) *primary multiple interiomarginal aperture* (Figs. 51A, B) (cf. *supplementary aperture*)

proloculus: initial embryonic chamber, usually well rounded. (Conspicuous in Figs. 20, 48, 61, 62, 64, 71A)

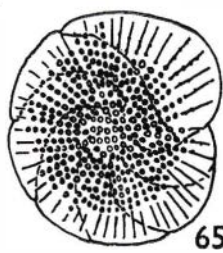
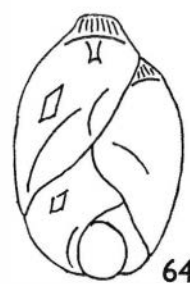
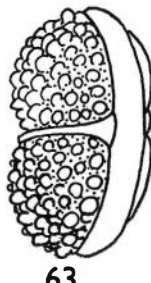
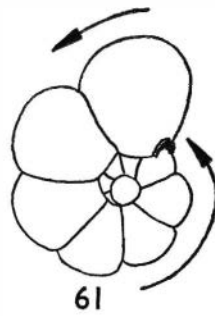
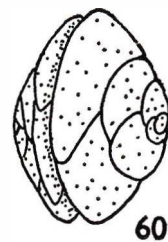
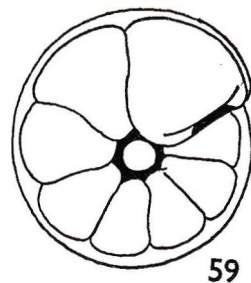
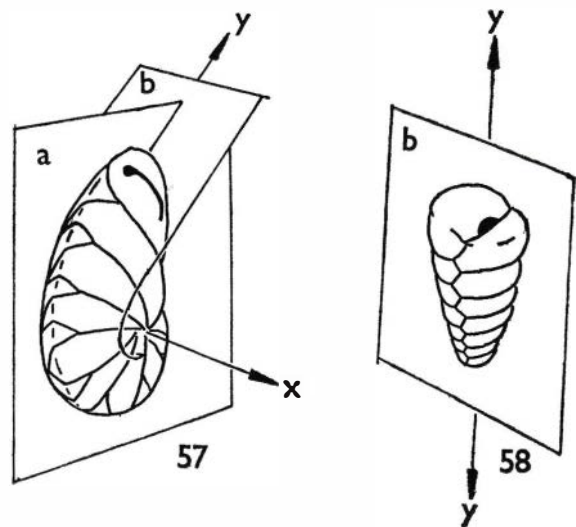
proximal: nearer to proloculus in direction of growth. (Figs. 61, 62)

pustule: small rounded elevation, hollow or solid. (Figs. 63, 65)

pustulose: having surface ornamented with pustules. (Figs. 63, 65)

pyriform: pear-shaped. e.g., *pyriform chambers* (Fig. 67A)

quinteloculine: milioline; each chamber added in plane at 144° to previous chamber; five chambers visible. (Figs. 14; 66A, B, C)



57. *Cassidulinoides* (a = plane of coiling, b = plane of growth, x = axis of coiling, y = axis of growth). 58. *Textularia* (b = plane of growth, y = axis of growth). 59. *Gavelinopsis*. 60. A plastogamic pair of *Glabrattella*. 61. *Anomalinoidea* (arrow pointing proximally). 62. *Saccorhiza*. 63. *Pegidia*. 64. *Esosyrinx*. 65. *Glabrattella*.

GLOSSARY—continued

radial suture: suture between successive chambers in spiral forms. (Fig. 22) e.g., *limbate radial sutures* (Figs. 50A; 75B, C) (cf. *spiral sutures*)

radial wall: test composed of calcite or aragonite crystals with c-axis perpendicular to surface; seen between crossed Nicols as black cross with concentric rings of colour mimicking negative uniaxial figure. (see Identification Techniques 3, p. 81)

radiate aperture: opening associated with numerous divergent slits. (Figs. 21; 40; 41; 46A, B; 64; 67A, B; 71B)

rectilinear: growing in a straight line; usually refers to uniserial forms, but may be applied to biserial forms. (Figs. 1, 21, 48, 67A)

reticulate: having surface marked by network, usually of raised ornamental ridges, not necessarily regular. (Figs. 68; 75B, C) (Note: Forms with a very fine regular network of ridges are referred to as *cancellate*)

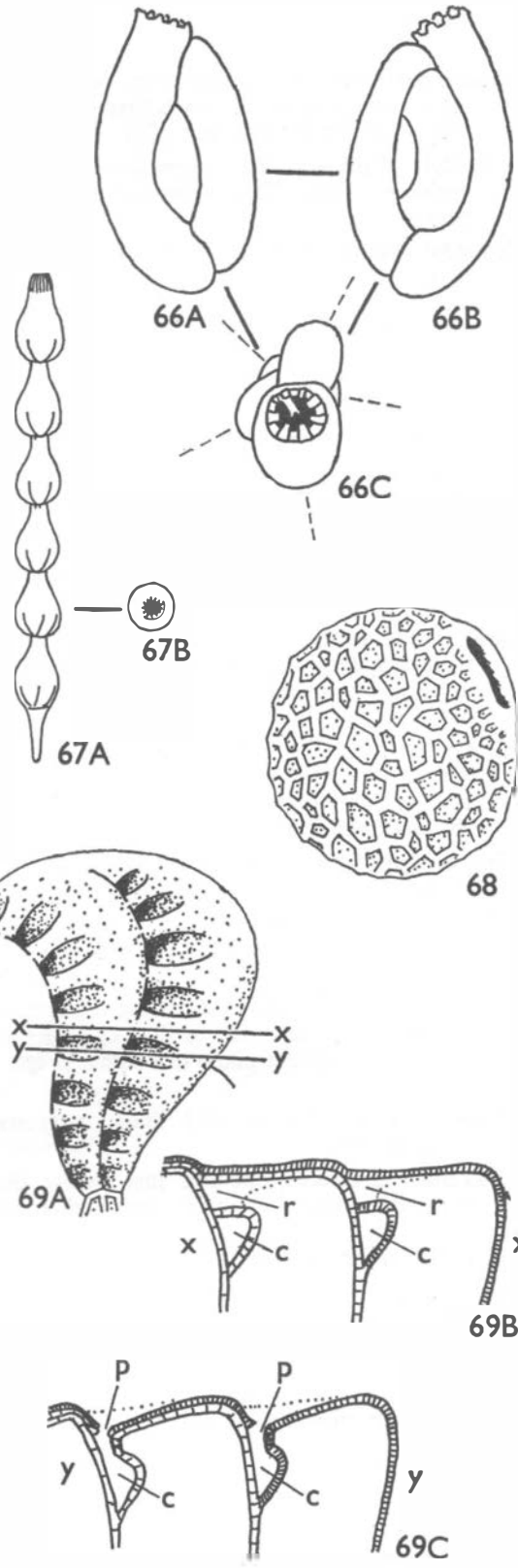
retral processes: proximally projecting extensions of chamber cavity located beneath external ridges on chamber wall (commonly above internal canal system) and ending blindly at face of previous chamber. (Figs. 69A, B, C, r = retral process; c = canal system; p = pore)

rotallid wall: chamber primarily of single layer continuing as thin lamella over whole of previously formed exterior of test. As each new chamber is added a lamella is also added over distal face of previously formed chamber enclosing a canal system between septal layers. Wall of final chamber single; all others double but incorporating a canal system. (Figs. 69B, C; 70)

rounded periphery: smooth transition between two sides of test; outer edge as viewed from stable position not angled. (Figs. 8, 32B, 51B) (cf. *angular periphery*)

secondary septum: plate inside chamber formed simultaneously with, or subsequent to, more distal portion of test, partly or completely dividing chamber into two or more chamberlets; independent of, and typically perpendicular to, true septa. (Fig. 5) (Sutures of secondary septa visible in Fig. 11)

septate: having partitions or septa dividing test into several chambers



66. *Dentostomina*. 67. *Nodosaria*. 68. *Favocassidulina*. 69. *Elphidium*: A part of exterior; B diagrammatic section through a retral process (x-x); C diagrammatic section between retral processes (y-y); r = retral process, c = canal, p = sutural pore.



GLOSSARY—continued

septum: partition between chambers, often consisting of distal face of previously formed chamber. (Figs. 18; 39; 52; 69A, B, C; 70)

sigmoidal: S-shaped. e.g., *sigmoidal suture and periphery* (Fig. 72B) *sigmoidal coiling* (see below)

sigmoidal coiling: chambers added at slightly less than 180° so that two sets of chambers curve outwards in opposite directions from central point producing S-shaped arrangement in median section of miliolids or basal view of polymorphinids. Two to many chambers visible. (Figs. 71A, B; 72A, B, C)

sinuate: having several bends. e.g., *sinuate suture* (Fig. 73)

spinose: having elongate and/or pointed projections from surface of test. (Fig. 74) e.g., *spinose periphery* (Fig. 29) (Note: Forms with a dense covering of very fine spines are referred to as hispid)

spiral ridges: elongate elevations roughly parallel with periphery or spiral suture of coiled forms, usually between adjacent radial sutures. (Figs. 69A; 75B, C)

spiral side: side of trochospiral forms where all whorls are visible, or side which is more evolute than opposite side. (Figs. 50A, 53B, 75C) (cf. *umbilical side*)

spiral suture: suture between adjacent whorls in evolute forms. (Fig. 26) (cf. *radial suture*)

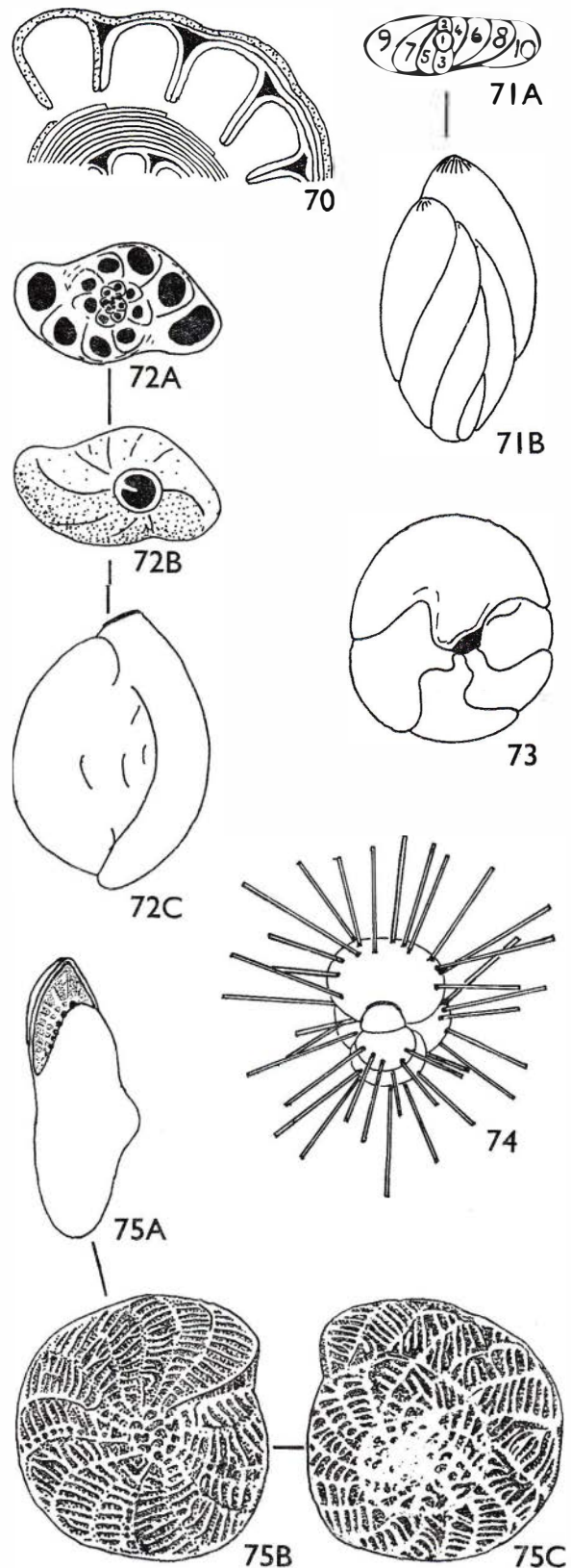
stolon: prolonged extension of test; often tubular portion connecting chambers. (Fig. 41)

streptospiral: coiled like ball of wool. During growth axis and plane of coiling change; may be one change or many or final chamber only may be in different plane of coiling. (Figs. 76, 77)

striate: marked with parallel lines or grooves. (Figs. 14, 78)

subterminal aperture: opening just below distal point of uniserial, biserial, and high trochospiral forms, or to one side of periphery of uniserial forms. (Figs. 45A, B)

supplementary aperture: opening of test to exterior additional to and independent of *primary aperture* (q.v.); in some forms may completely replace primary aperture, e.g., *supplementary areal apertures* (Figs. 51A, B); *supplementary sutural apertures* (Figs. 51A, B, 81, 90)



70. Diagrammatic section through a rotallid wall. 71. *Sigmomorphina*. 72. *Sigmoidina*. 73. *Tiphotrocha*. 74. *Hastigerina*. 75. *Polystomellina* (*Notorotalia*).

GLOSSARY—continued

sutural aperture: opening situated on suture, not on apertural face. (Figs. 51A, B; 69A; 81; 90)

sutural flap: flattened extension of proximal wall of chamber over suture. (Fig. 81)

suture: line of union between two chambers or two whorls. e.g., *limbate suture* (Figs. 31A, B; 50A) *spiral suture* (Fig. 26) *radial suture* (Fig. 22)

tectin: organic substance, basically glycoprotein. Looks like chitin but is distinct chemically. (Synonym of *pseudochitin*)

tectinous: having shell or layer of tectin

terminal: at distal point. e.g., *areal terminal aperture* (Figs. 21, 41, 47, 48, 67A) *interiomarginal terminal aperture* (Fig. 79)

test: agglutinated or secreted shell of animal

tooth: projection into principal opening of test. e.g., *simple tooth* (Fig. 72B) *bifid tooth* (Figs. 14; 17A, B; 66C) *apertural flap* (Fig. 6) *umbilical teeth* (Fig. 90) *tooth plate* (see below)

tooth plate: internal apertural modification consisting of contorted plate that extends from aperture, through chamber, commonly as far as previous foramen. (Fig. 80) e.g., *part of tooth plate projecting through aperture* (Fig. 79)

trematophore: perforate plate over aperture of some members of Miliolacea; sometimes referred to as a sieve plate. (Fig. 82)

triloculine: milioline, each chamber added in a plane at 120° to previous chamber, three chambers visible. (Figs. 84A, B, C)

triradiate aperture: opening consisting of three divergent slits. (Fig. 84A)

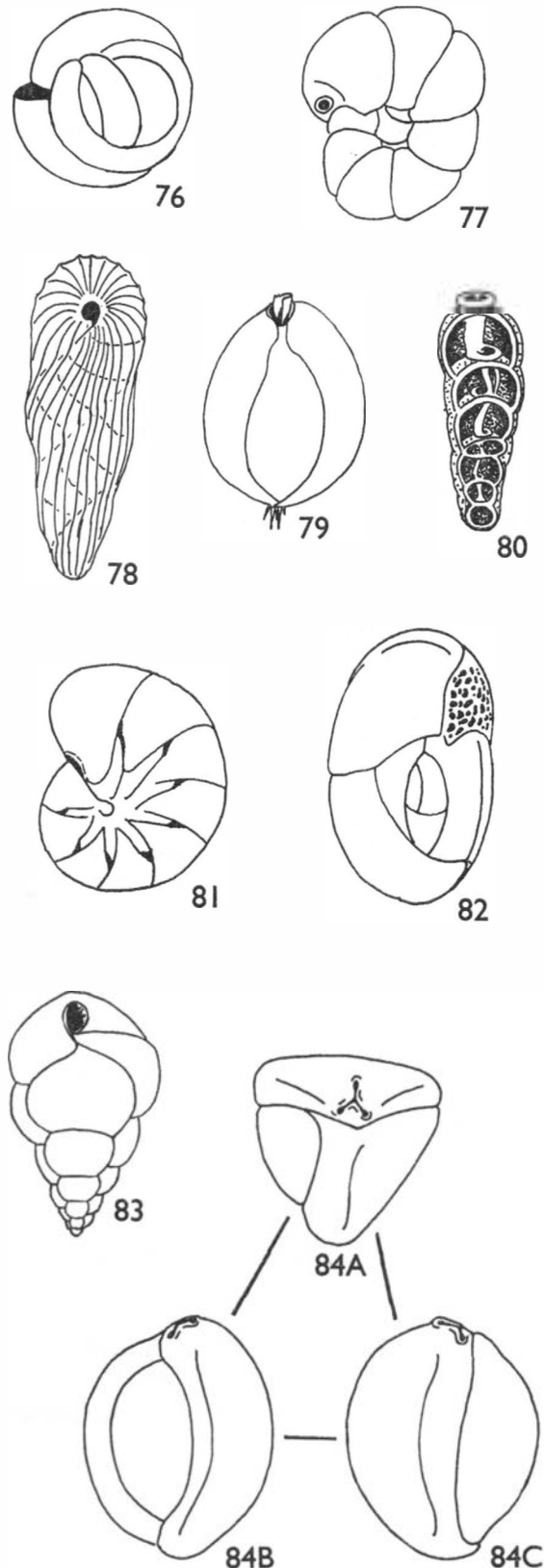
triserial: high trochospiral, with three chambers in each whorl, which results in three columns of chambers. (Fig. 83)

trochospiral: chambers spirally coiled, evolute on one side, involute on opposite side. e.g., *low trochospiral* (Figs. 50A, B; 53A, B, C; 75A, B, C) *high trochospiral* (Figs. 78, 83)

truncate periphery: flattened periphery with sharp angle to two sides. (Figs. 21; 85A, B)

twisted biserial: with chambers in two columns which spiral around central axis; plane of growth twisted about axis of growth. (Figs. 86, 87)

umbilical aperture: arched opening in final chamber; lower boundary defined by umbilicus; does not extend outside umbilical region. (Figs. 88, 89)



76. *Glomospira*. 77. *Recurvoides*. 78. *Buliminoides*. 79. *Globobulimina*. 80. *Rectobolivina*. 81. *Astronion*. 82. *Hauerina*. 83. *Bulimina*. 84. *Cruciloculina*.

GLOSSARY—continued

umbilical bulla: blister-like structure that covers umbilical region. (Fig. 19)

umbilical flap: extension of chamber wall over umbilicus, possibly covering umbilicus. (Figs. 53C, 90)

(Note: when main aperture is umbilical (e.g. Fig. 89), flaps are called umbilical teeth)

umbilical plug: solid shell material occupying open umbilicus. Surface of plug may be subdivided. (Figs. 50B, 59)

umbilical region: area round umbilicus. e.g., *granulate covering of umbilical region* (Fig. 85)

umbilical side: surface of trochospiral forms where only final whorl visible or which is more involute than opposite surface. (Figs. 50B, 53C, 75B) (cf. *spiral side*)

umbilical tooth: plate-like extension of inner margin of chamber into umbilical aperture, usually triangular. (Fig. 89)

umbilicate: having an umbilicus. Usually used to describe forms with a very depressed umbilicus. (Figs. 24; 53A, C)

umbilicus: centre (possibly depressed) of involute planispiral forms, or umbilical (involute) side of trochospiral forms where all chambers of last-formed whorl meet. e.g., *open umbilicus* (Fig. 53C) *closed umbilicus* (Fig. 22)

umbo: dome, cone, or knob of solid shell material in centre of planispiral coil or spiral side of low trochospiral coil; usually only surface thickening. (Figs. 75A, C; 85A, B)

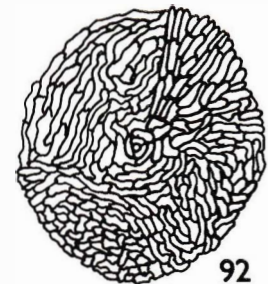
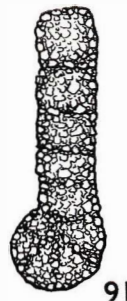
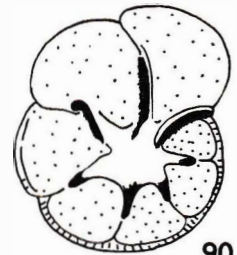
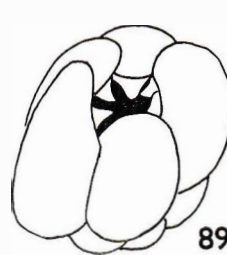
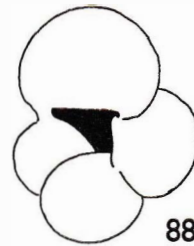
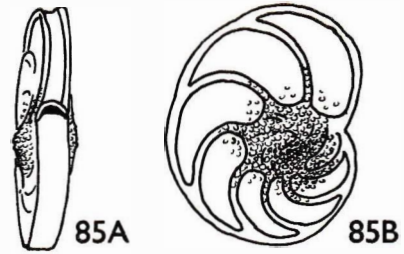
uncoiling: having early chambers enrolled, usually planispirally, later portion biserial or uniserial with axis of growth perpendicular to axis of coiling. e.g., *close-coiled changing suddenly to rectilinear uniserial* (Fig. 91) *gradually uncoiling (biserial) series* (Fig. 57) *uncoiling with later chambers arcuate* (Fig. 37)

unilocular: single chambered, test not divided by septa into chambers. Also used to describe undivided tubular forms which arise from an initial proloculus. e.g., *single globose chamber* (Figs. 9, 33, 40, 42) *essentially tubular chamber* (Figs. 2, 12, 26, 28, 62) cf. multilocular

uniserial: having chambers in single row. (Figs. 1, 21, 41, 67A)

vermiform: long, thin, cylindrical, usually irregularly bent. e.g., *vermiform chambers* (Fig. 92)

whorl: single turn or volition of coiled test through 360°.



85. *Planulina*. 86. *Compressigerina*. 87. *Fursenkoina*.
88. *Globigerina*. 89. *Globoquadrina*. 90. *Discorbis*.
91. *Ammobaculites*. 92. *Planogypsina*.

USEFUL TECHNIQUES

COLLECTING

Pelagic

1. An N70 plankton net (Kemp and Hardy 1929) with fine mesh of about 200 microns (86 meshes per in.) is satisfactory for normal vertical or oblique tows (cf. Bé 1959a).
2. A Clarke-Bumpus sampler (Clarke and Bumpus 1940) is towed at constant depth (horizontal tow) and the volume of water strained is measured by a flow meter. It must be towed slowly through considerable distances.
3. The continuous plankton recorder (Hardy 1936) is suitable for high-speed towing through near-surface waters.
4. The multiple plankton sampler (Bé, Ewing, and Linton, 1959) will take samples at successive depth ranges in a vertical haul. Several nets are opened and closed at the required depth by messengers or in more refined models (Bé 1962) by a pressure gauge.

Benthonic

1. The Phleger corer (Phleger 1951), a short corer with a 1½ in. diameter plastic inner lining, collects a relatively undisturbed sample of known surface area. At the N.Z. Oceanographic Institute an even shorter barrel with 2 in. diameter plastic liner and much wider weights (to prevent deep penetration) has been found satisfactory. Square weights will prevent the equipment rolling on deck in rough weather, and a bayonet-fix cutter described by Willis (1966) is easily fixed and removed but does not slip off when the corer is being used. Care must be taken to keep all apparatus vertical until the sample is removed. The same volume of sediment is removed from the surface of the core on each occasion (about 1 cm depth) and placed in a jar with the water from immediately above the sediment and 90% ethyl alcohol as preservative. The short corer is the normal equipment for sediment and foraminiferal distribution studies at the New Zealand Oceanographic Institute, but it is of little use in areas of pure sand or pebbles.
2. An orange-peel grab modified with a canvas skirt (Phleger 1952; Reish 1959) or with plates at the top of the blades to protect the surface layer will collect a suitable sample of sand or gravel. A piece of plastic liner is pressed into the surface and the upper 1 cm of sediment removed, placed in a jar, and 90% ethyl alcohol added. Alternatively, a shorter corer can be built into the orange-peel grab as illustrated by Willis (1966).

3. If sampling is done in very shallow water from a small boat, lightweight equipment must be used. A short corer with movable weights is particularly suitable (Murray 1961). The line is attached to the weights so that by repeatedly raising them and letting them fall the corer is driven into the sediment. If the investigation is not quantitative, a small snapper grab (Lafond and Dietz 1948) or even simply a piece of tube, one end of which is closed and the opposite end attached by a line to the surface, will collect a sample of the surface sediment.
4. Waves lapping onto a beach commonly leave a faint white line as each one recedes. If this is very carefully removed with a sharp knife or razor blade, it is usually found to be rich in empty tests of the local littoral and sub-littoral foraminiferal population.
5. Living specimens for observations and culture studies can be obtained from intertidal weeds. Around New Zealand the calcareous algae *Corallina officinalis* (Linnaeus) forms a purplish brown carpet in many tide pools and usually provides a varied fauna of living foraminifers (Hedley *et al.*, 1967). The algae can be collected in handfuls and shaken under water over a coarse sieve (about 16 meshes per in., 1.0 mm aperture) with a finer sieve (about 120 meshes per in., 0.125 mm aperture) below it. If the debris from the finer sieve is left to stand overnight in a shallow dish of sea water, many foraminifers, particularly miliolids which are negatively geotropic, will climb to local high points and up the sides of the dish. These can easily be removed for experiments.

If, during calm weather, glass slides are left overnight among weeds in a tide pool, the animals will commonly crawl on to the slide and so can be removed completely unharmed from their natural habitat.

SAMPLE PROCESSING

Pelagic

Shelled planktonic organisms (foraminifers, radiolaria, and pteropods) can be separated from the mass of unshelled organisms by a density separation in saturated sodium chloride solution (Bé 1959b).

Benthonic

1. *Washing*: Wash through a 240 mesh per in. sieve (0.064 mm aperture), which removes alcohol and all material of silt and clay grade, including juvenile and very small foraminifers.

2. *Staining*: Protoplasm is stained satisfactorily by Rose Bengal (Walton 1952).

- (a) Transfer the residue to a beaker containing an aqueous solution of Rose Bengal. The solution should be dark pink; the concentration is not critical.
- (b) Leave for more than 30 minutes, preferably several hours.
- (c) Drain off the stain on a 240 mesh per in. sieve then wash the sample thoroughly to remove all excess stain. The sample is easier to study if it is washed through a nest of sieves and the fractions of similar size are examined separately. Removal of surface staining is most effective if the sample is left to stand in clean water and washed again.
- (d) Wash the sample on to a counting tray.
- (e) The sample is best examined under water at this stage; as the shell wall is usually less opaque when wet very fragile specimens may be broken by further processing and tectinous forms would shrivel beyond recognition when dried. Specimens that were alive when the sample was collected usually have blobs of pink- or red-stained protoplasm in one or many chambers. Empty tests may have an epizoic fauna of bacteria or algae which when stained appears as a patchy pink film on the interior surface.

3. *Dividing large samples*: Large samples may be repeatedly divided into two roughly equal portions by either

- (a) make a mound of the sample, divide it twice at right angles then take opposite quarters, or
- (b) use the microsplitter described by Parker (1948), which consists of a trough bisected by a knife edge, or
- (c) for more accurate work use the microsplitter designed by Otto (1933) which has channels pointing in opposite directions.

The sample can then be spread evenly and sparsely on the counting tray and each specimen in random squares identified. When the required number has been counted the total of each group of foraminifers in the sample can be estimated. For counts of dead or total population, the material is more easily sieved and picked if it is first dried.

4. *Concentrating*: Except for quantitative studies the foraminifers can be concentrated by a density separation in carbon tetrachloride (Cushman 1940).

- (a) The washed and dried sample is poured into a beaker containing carbon tetrachloride.
- (b) The floating portion with the carbon tetrachloride is decanted off into a filter funnel.
- (c) The filtered carbon tetrachloride is retained for further use. The residue in the filter paper, which contains most of the whole, airfilled foraminifers, is dried. The residue in the beaker contains all the mineral grains, and broken and dense foraminifers.

Warning: Carbon tetrachloride vapour is extremely dangerous if inhaled in large quantities, through a lighted cigarette, or after drinking alcohol. It is recommended that concentration be done in the open air or in a room with doors open to create a draught. Fume cupboards are often ineffective in removing the dense vapour.

Bromoform or tetrabromomethane diluted with acetone to a specific gravity of about 2.3 can be used instead of carbon tetrachloride. Both heavy liquids are less volatile and toxic than carbon tetrachloride and float off a far higher proportion of the foraminifers (Gibson and Walker 1967). However, pellets, ash, and glauconite grains may also float, sometimes in large numbers, and thorough washing of the concentrate and residue can be time consuming.

REMOVING SPECIMENS FROM SAMPLE

1. Specimens can be picked from the dry sample with an artist's OO or O sable hair brush. Dipped in water and drawn lightly across a piece of cloth this will produce a very fine point to which dry tests will adhere. Specimens immersed in water can be removed, with practice, by wedging them between the bristles of the brush.
2. A vacuum-needle segregating pick (Stinemeyer 1965) is a useful tool, if large numbers of dry samples are to be picked and sorted.

MOUNTING

1. Cardboard slides with a black-based tray are customarily used for mounting specimens. A thin layer of gum tragacanth is painted on to the tray as an adhesive. Before it is mixed to a thin paste with water, about 5% by weight of mercuric chloride or clove oil should be added to the gum tragacanth powder to inhibit mould growth and attack by mites and silverfish.
2. Plastic slides are now being produced with a photo-paper base to which a damp foraminifer will stick automatically. These are very convenient and cheap. Gum tragacanth must still be used to stick large specimens and the mounts are easily damaged by some oils (not clove oil), repeated moistening, and mercuric chloride.

IDENTIFICATION TECHNIQUES

1. *To reveal surface ornamentation or apertural details*
Blue or green fountain pen ink painted on to the surface usually enhances detail; a solution of methyl blue or malachite green may be useful for very fine structures.

2. *To reveal internal structure*

If the specimen is dampened with water or oil, considerably more internal detail is usually revealed. Oil of cedarwood and aniseed oil are both satisfactory, but aniseed oil damages plastic mounts.

Greater detail can be observed if the specimen is transferred to a glass slide with a central cavity, covered with a few drops of oil, and examined in transmitted light. Glycerine or castor oil may be used to make the mount semi-permanent. If necessary the specimen can be crushed or cut with a razor blade while immersed in oil.

With care the shell wall can be dissected away with the point of a fine brush that has been dipped in very dilute hydrochloric acid.

3. To distinguish optical characters of calcareous wall

The optical characters and arrangement of crystals in the calcareous wall are most readily distinguished if the specimen is transferred to a glass slide, covered with a drop of oil, crushed, and the fragments examined in transmitted light and between crossed Nicols (polaroids) (Table 1).

4. To reveal mineralogical composition of calcareous walls

Calcite and aragonite can be distinguished by using Feigl's solution (Feigl 1954).

To make up Feigl's solution:

- (a) Dissolve 11.8 g of $\text{MnSO}_4 \cdot 4\text{H}_2\text{O}$ in 100 ml of water.
- (b) Dissolve 1 g of AgSO_4 in the MnSO_4 solution.
- (c) Bring to boil.

- (d) Cool.
- (e) Filter off suspension.
- (f) Add 1 or 2 drops of dil. NaOH solution to filtrate.
- (g) Filter after 2 hours.
- (h) Store solution in small dark airtight bottle away from direct sunlight; it will then keep for many months or even years.

To stain with Feigl's solution:

- (a) Transfer specimen to glass slide.
- (b) Etch surface by painting with very dilute H_2SO_4 and leaving for about 1 minute.
- (c) Add several drops of pure alcohol to wash away excess acid, then wait for specimen to dry.
- (d) Paint on Feigl's solution, wait for 5–10 minutes.

If specimen remains unaffected by Feigl's solution it is calcite. If specimen becomes black in Feigl's solution it is aragonite.

TABLE 1. Distinguishing optical characters in calcareous walls

Transmitted light	X Nicols	Type of wall	
amber or reddish brown without pores	no figure, possibly specks of colour	porcellaneous wall	
colourless, milky, or pale blue with pores	black cross with rings of colour mimicking uniaxial figure	radial wall	hyaline wall
	mottled with specks of colour	granular wall	

TABLE 2. Lamellar nature of calcareous walls observed in thin section

Lamella from each chamber continuing over previously formed exterior of test	Septa	Final chamber	Type of wall
absent; test typically of single layer throughout	of single lamella	of single lamella	non-lamellar
present	of single lamella	of single lamella	monolamellar <i>see</i> Glossary Fig. 52
present	double or partly double, with canal system between lamellae	of single lamella	rotallid wall <i>see</i> Glossary Figs. 69 B, c; 70
present	double, without canal system between lamellae	double	bilamellar <i>see</i> Glossary Fig. 18

5. To reveal lamellar nature of calcareous wall

The structure of the shell wall can be readily observed in thin section. Preparation of thin section (Hornibrook, 1968):

- (a) Heat specimen gently on clean slide with very small piece of Lakeside 70c cement. Use hot plate at about 140°C.
- (b) Transfer to binocular microscope and orientate specimen with hot needle.
- (c) Allow to cool.
- (d) Grind on glass plate with aluminium oxide powder mixed to a thin paste with water. 302 grade aluminium oxide is preferable for large specimens, 303 for more delicate forms.
- (e) Wash and examine under microscope at frequent intervals. If necessary reheat so that interiors of chambers are filled with cement.
- (f) When nearing required state of grinding, transfer to microscope stage and continue grinding under observation in reflected light by slowly rotating over the specimen another slide lubricated with

aluminium oxide paste.

- (g) When required position is reached reheat slide and turn specimen over with a hot needle, pressing the flattened face against slide.
- (h) Allow to cool.
- (i) Continue grinding on glass plate.
- (j) Grind final stages under observation, but this time use of transmitted light and fine (304) grade grinding powder is advisable.
- (k) When required thickness is reached wash and dry slide.
- (l) Add drop of fairly fluid Canada balsam/xylene mixture.
- (m) Rest one edge of cover slip in place, then lower it slowly into position.
- (n) Heat for some time on hot plate at 90°–100°C.
- (o) Allow to cool.

Layers (lamellae) making up the walls and septa can be seen by examining the thin section under high magnification on a petrological microscope. (Table 2).

REFERENCES

- BANNER, F. T.; BLOW, W. H. 1960: The taxonomy, morphology and affinities of genera included in the subfamily Hastigeriinae. *Micropaleontology* 6 (1): 19–31.
- BÉ, A. W. H. 1959a: Ecology of Recent planktonic Foraminifera. Pt 1. Areal distribution in Western North Atlantic. *Micropaleontology* 5: 77–100.
- 1959b: A method of rapid sorting of Foraminifera from marine plankton samples. *J. Paleont.* 35 (5): 846–8.
- 1962: Quantitative multiple opening and closing plankton samples. *Deep Sea Res.* 9: 144–51.
- 1965: The influence of depth on shell growth in *Globigerinoides sacculifera* (Brady) *Micropaleontology* 11 (1): 81–97.
- BÉ, A. W. H.; EWING, M.; LINTON, L. M. 1959: A quantitative multiple opening and closing plankton sampler for vertical towing. *J. Cons. perm. int. Explor. Mer.* 25 (1): 36–46.
- CLARKE, G. L.; BUMPUS, D. F. 1940: The plankton sampler—an instrument for quantitative plankton investigations. *Spec. Publs. limnol. Soc. Am.* 5.
- CUSHMAN, J. A. 1940: "Foraminifera; their Classification and Economic Use." 3rd ed. Harvard University Press.
- EADE, J. V. 1967: A Checklist of Recent New Zealand Foraminifera. *Bull. N.Z. Dep. scient. ind. Res.* 182.
- FEIGL, F. 1954: "Spot Tests." Elsevier, London, 518 pp.
- GIBSON, T. G.; WALKER, W. M. 1967: Flootation methods for obtaining foraminifera from sediment samples. *J. Pal.* 41: 1294–9.
- HARDY, A. C. 1936: The continuous plankton recorder. "Discovery" Rep. 11: 457–503.
- HEDLEY, R. H. 1962: *Gromia oviformis* (Rhizopodea) from New Zealand with comment on fossil Chitozoa. *N.Z. Jl Sci.* 5 (2): 121–36.
- HEDLEY, R. H.; HURDLE, C. M.; BURDETT, I. D. J. 1967: The Marine Fauna of New Zealand. Intertidal Foraminifera of the *Corallina officinalis* zone. *Bull. N.Z. Dep. scient. ind. Res.* 180.
- HOLME, N. A. 1955: An improved vacuum grab for sampling the sea floor. *J. mar. biol. Ass. U.K.* 34: 545–51.
- HORNIBROOK, N. de B. 1968: A Handbook of New Zealand Microfossils. *N.Z. Dep. scient. ind. Res. Inf. Ser.* 62.
- HUNT, O. D. 1929: A new method of quantitative sampling of the sea bed. *J. mar. biol. Ass. U.K.* 2: 137–47.
- KEMP, S.; HARDY, A. C. 1929: The *Discovery* investigations, objects, equipment and methods. Pt II. The ships, their equipment and methods used in research. "Discovery" Rep. 1: 151–222 (183–184).
- LAFOND, E. C.; DIETZ, R. S. 1948: New Snapper-type seafloor sediment sampler. *J. sedim. Petrol.* 18: 34–7.
- LEWIS, K. B.; JENKINS, C. 1969: Geographical variation of *Nonionellina flemingi*. *Micropaleontology* 15: 1–12.
- LOEBLICH, A. R., jun; TAPPAN, H. 1961: Suprageneric Classification of the Rhizopoda. *J. Paleont.* 35: 245–330.
- 1963: Four new Recent genera of Foraminifera. *Protozoology* 10 (2): 212–5.
- 1964a: "Treatise on Invertebrate Paleontology." Part C. Protista 2. Sarcodina Chiefly Thecomoebian and Foraminiferida." Vol. 1: C1–C510d. Vol. 2: C591–C900, 652 figs. The Geological Society of America and the University of Kansas Press, New York.
- 1964b: Foraminifera classification and evolution. *J. geol. Soc. India* 5: 5–40.
- METCALF, Z. P. 1954: The construction of keys. *Syst. Zool.* 3: 38–45.
- MURRAY, J. 1961: Ecology and systematics of the benthonic Foraminifera of Christchurch Harbour, Hampshire. *Unpublished thesis, University of London.*
- OTTO, G. H. 1933: Comparative tests on several methods of sampling mineral concentrates. *J. sedim. Petrol.* 3 (1): 30–9.
- PARKER, F. L. 1948: Foraminifera of the continental shelf from the Gulf of Maine to Maryland. *Bull. Mus. comp. Zool. Harv.* 100 (2): 213–41.
- 1962: The planktonic foraminiferal species in Pacific sediments. *Micropaleontology* 8 (2): 219–54.
- PHLEGER, F. B. 1951: Ecology of Foraminifera, north-west Gulf of Mexico Pt. 1. Foraminiferal distribution. *Mem. geol. Soc. Am.* 46: 1–88.
- 1952: Foraminifera ecology off Portsmouth, New Hampshire. *Bull. Mus. comp. Zool. Harv.* 106 (8): 315–90.
- REISH, D. J. 1959: Modification of the Hayward orange-peel bucket for bottom sampling. *Ecology* 40 (3): 502–3.
- STINEMEYER, E. H. 1965: Microfossil vacuum-needle segregating pick. Pp. 276–83. In "Handbook of Paleontological Techniques." Eds B. Kummel and D. Raup. Freeman, London, 852pp.
- WALTON, W. R. 1952: Techniques for recognition of living Foraminifera. *Contr. Cushman Fdn foramin. Res.* 3 (2): 56–60.
- WILLIS, R. P. 1966: A simple joint for attaching core cutters to barrels. *N.Z. Jl Sci.* 9: 77–83.

INDEX

- Acervulina*, 38, 58
Acervulinidae, Fam., 35, 55, 58
Acostina, 43
Adercotryma, 16, 19, 20
Alabamina, 63, 69
Alabaminidae, Fam., 59, 63
Alanwoodia, 49
Alfredosilvestris, 37
Alliatina, 66
Allogromia, 8
Allogromiidae, Fam., 8
Allogromiina, Subord., 8
Allomorphina, 40, 47, 62
Almaeninae, Subfam., 64, 65
Alveolinella, 29
Alveolinidae, Fam., 22, 29
Alveolophragmium, 17, 67
Ammoastuta, 17
Ammobaculites, 17, 78
Ammobaculoides, 17
Ammodiscacea, Superfam., 8
Ammodiscidae, Fam., 8
Ammodiscinae, Subfam., 13
Ammodiscoides, 13
Ammodiscus, 13
Ammoflinitina, 15, 16
Ammolagena, 14
Ammomarginulina, 17
Ammomassilina, 27, 73
Ammonia, 50, 51, 72
Ammopemphix, 12
Ammoscalaria, 17
Ammosphaeroidina, 16, 19
Ammosphaerulina, 16
Ammotium, 17
Ammovertella, 14
Ammovertellina, 13
Amphicoryna, 36, 37, 71
Amphifenestrella, 11
Amphimorphina, 37
Amphisorus, 29
Amphistegina, 56
Amphisteginidae, Fam., 32, 55, 56
Angulodiscorbis, 48
Annulopatellina, 60
Annulopatellinidae, Fam., 31, 34, 59, 60
Anomalina, 65
Anomalinella, 65, 73
Anomalinidae, Fam., 32, 59, 63, 64
Anomalininae, Subfam., 64, 65
Anomalinoides, 65, 74
Archaias, 28, 29
Archaiasinae, Subfam., 28, 29
Arenoparrella, 20
Articulina, 28
Asanoina, 50
Aschemonella, 14
Aschemonellinae, Subfam., 14
Astacolus, 36
Asterigerina, 48
Asterigerinata, 48
Asterigerinidae, Fam., 33, 45, 48
Asterorotalia, 50
Astrammia, 9
Astrononion, 62, 77
Astrorhiza, 9
Astrorhizidae, Fam., 8, 9
Astrorhizinae, Subfam., 9
Ataxophragmiidae, Fam., 14, 20
Ataxophragmiinae, Subfam., 20, 22
Baculogypsina, 51
Baculogypsinoides, 51
Baggina, 47, 65
Baggininae, Subfam., 45, 47
Barbourinella, 20
Bathysiphon, 9, 10
Bdelloidina, 18
Beella, 54
Biarrizina, 51
Bigenerina, 19
Bilocolinella, 27, 67
Bisaccium, 62
Bolivina, 41
Bolivinita, 41
Bolivinitidae, Fam., 30, 31, 40
Borelis, 29
Botellina, 10
Botellininae, Subfam., 9, 10
Brachysiphon, 10, 11
Brizalina, 41, 67
Bronnimannia, 47
Buccella, 46
Bulimina, 43, 77
Buliminacea, Superfam., 30, 40
Buliminella, 40
Buliminidae, Fam., 34, 40, 42
Bulimininae, Subfam., 42
Buliminoides, 40, 77
Calcarina, 51
Calcarinidae, Fam., 33, 35, 49, 51
Calcituba, 24
Cancris, 47
Candaina, 54
Canepaia, 17
Caribbeanella, 57
Carpentaria, 57, 59
Carterina, 65
Carterinacea, Superfam., 30, 65
Carterinidae, Fam., 33, 34, 65
Cassidella, 60
Cassidulina, 41, 61, 70
Cassidulinacea, Superfam., 30, 59
Cassidulinidae, Fam., 31, 59, 61
Cassidulinoides, 41, 74
Catapsydracinae, Subfam., 54, 55
Caucasinidae, Fam., 31, 59
Caucasininae, Subfam., 60, 61
Causia, 11
Cellanthus, 52
Ceratobulimina, 48, 65
Ceratobuliminidae, Fam., 33, 65
Ceratobulimininae, Subfam., 65
Cerobertina, 48, 66, 68
Chilostomella, 62, 67
Chilostomellina, 62
Chilostomellinae, Subfam., 61, 62

Chrysalidinella, 44
Chrysalogonium, 36
Cibicidella, 57
Cibicides, 57, 58
Cibicididae, Fam., 31, 32, 33, 34, 51, 55, 56
Cibicidinae, Subfam., 56, 57
Clavulina, 21
Colonammina, 12
Compressigerina, 44, 78
Conicospirillina, 49
Cornuloculina, 24
Cornuspiramia, 23, 24
Cornuspirella, 23
Cornuspiroides, 23, 28
Coryphostoma, 60
Coscinophragmatinae, Subfam., 16, 18
Craterites, 29
Cribrorigenerina, 19
Cribrbulimina, 21
Cribroelphidium, 52, 73
Cribrogoesella, 21
Cribrolinoides, 25, 27
Cribrononion, 52
Cribropyrgo, 27
Cribrobulimina, 36
Cribrostomoides, 16
Crithionina, 13
Cruciloculina, 26, 69, 77
Cushmanella, 66
Cyclammina, 17
Cyclammininae, Subfam., 16, 17
Cyclocibicides, 57
Cycloclypeinae, Subfam., 52
Cycloclypeus, 53
Cyclogyra, 23, 49, 69
Cyclogyrinae, Subfam., 23
Cyclorbiculina, 29
Cylindroclavulina, 21, 22
Cymbaloporella, 58
Cymbaloporetta, 58
Cymbaloporidae, Fam., 34, 55, 58
Cystammina, 19, 73

Daitrona, 13
Delosina, 61
Delosinidae, Fam., 33, 34, 59, 61
Dendritina, 28
Dendronina, 9
Dendrophryinae, Subfam., 9, 10
Dendrophrya, 10, 70
Dentalina, 37
Dentostomina, 27, 75
Diffusilina, 13
Diffusilininae, Subfam., 11, 13
Dimorphina, 36
Discammina, 16
Discanomalina, 64
Discobotellina, 13
Discopulvinulina, 46
Discorbacea, Superfam., 30, 45
Discorbidae, Fam., 32, 33, 45
Discorbininae, Subfam., 45
Discorbinella, 46, 47
Discorbis, 46, 78
Discospirina, 25, 29, 67
Discospirininae, Subfam., 23, 25
Dorothia, 21
Dusenburyina, 21
Dyocibicides, 57

Earlmyersia, 47
Edentostomina, 24
Eggerella, 21
Ehrenbergina, 41, 61, 70

Ellipsoglandulina, 60
Elphidiidae, Fam., 32, 33, 49, 51
Elphidiinae, Subfam., 51, 52
Elphidium, 52, 75
Entolingulina, 37, 39
Entzia, 20
Eouvigerinidae, Fam., 31, 40, 42
Ephidiella, 52
Epistomariidae, Fam., 32, 45, 48
Epistomaroides, 48
Epistominella, 46
Epistomininae, Subfam., 65
Eponidella, 48, 66
Eponides, 56
Eponididae, Fam., 33, 55, 56
Esosyrinx, 39, 74
Euuvigerina, 45

Fabularia, 28
Fabulariinae, Subfam., 25, 28
Faujasininae, Subfam., 51, 52
Favocassidulina, 61, 75
Fijiella, 44
Fischerina, 23
Fischerinella, 23
Fischerinidae, Fam., 22, 23
Fischerininae, Subfam., 23
Fissurina, 39, 70
Flintina, 25, 26, 27
Florilus, 63
flemingi, 62
Francesita, 61
Fronicularia, 37, 69
Fursenkoina, 60, 78
Fursenkoininae, Subfam., 60

Gaudryina, 21
Gavelinopsis, 46, 74
Glabratella, 47, 74
Glabratellidae, Fam., 33, 45, 47
Glandulina, 39
Glandulinidae, Fam., 30, 35, 38
Gladulininae, Subfam., 38, 39
Globigerina, 19, 54, 78
Globigerinacea, Superfam., 30, 31, 53
Globigerinella, 53
Globigerinidae, Fam., 53, 54
Globigerininae, Subfam., 54
Globigerinita, 55
Globigerinoides, 54
sacculifera, 54
Globobulimina, 43, 62, 77
Globocassidulina, 61
Globoquadrina, 54, 69, 78
Globorotalia, 53, 56
Globorotaloides, 54
Globorotaliidae, Fam., 53
Globorotaliinae, Subfam., 53
Globotextularia, 21
Globotextulariinae, Subfam., 20, 21
Globulina, 38
Globulotuba, 39
Glomospira, 14
Glomulina, 24
Goatapitiqba, 12, 18
Goesella, 21, 22
Gordiospira, 23
Gromia, 8
Guttalina, 38
Gypsina, 58
Gyroidina, 63
Gyroidinoides, 63

Haddonina, 18
Halyphysema, 10
 Hantkeninidae, Fam., 53
Hanzawaia, 64
Haplophragmoides, 16
 Haplophragmoidinae, Subfam., 16
Hastigerina, 53, 76
Hastigerinella, 53, 54
 Hastigerininae, Subfam., 53
Hauerina, 27, 77
Helenina, 46, 57
 Hemisphaerammininae, Subfam., 11, 12
Heronallenia, 47
Heterolepa, 64
Heterostegina, 53
Hippocrepina, 10
 Hippocrepininae, Subfam., 9, 10
Hoeglundina, 65, 72
Homotrema, 59
 Homotrematidae, Fam., 34, 55, 58
 Homotrematinae, Subfam., 58, 59
Hopkinsina, 44
Hormosina, 15
 Hormosinidae, Fam., 14
 Hormosininae, Subfam., 14, 15
Hyalinea, 56
Hyperammina, 10, 11

Involvohauerina, 27
Iridia, 8, 12
Islandiella, 41
 Islandiellidae, Fam., 31, 34, 40, 41

Jaculella, 10
Jadammina, 20
Jullienella, 11, 71

Kalamopsis, 14
Karamosphaera, 29
 Karamosphaerinae, Subfam., 28, 29
Karrerria, 64
Karrerriella, 21

Lagena, 35, 71
Lagenammina, 10, 11
Lagenoglandulina, 35, 37
Lamarckina, 65
Lamelloscorbis, 47, 67
 Larginidae, Fam., 8
Laryngosigma, 39
Laticarinina, 47, 68
Lenticulina, 36, 72
Liebusella, 19, 21, 22
Lingulina, 37
 Lingulininae, Subfam., 35, 37
Lingulopsis, 37
Lituola, 17
 Lituolacea, Superfam., 8, 14
 Lituolidae, Fam., 14, 15
 Lituolinae, Subfam., 16, 17
Lituotuba, 14, 18
Loxostomum, 60

Marginopora, 29
Marginulina, 36
Marginulinopsis, 36
Marsipella, 9

Martinottiella, 21, 22
Massilina, 26
Meandrospira, 23
Melonis, 64
 Miliolacea, Superfam., 22
 Miliolidae, Fam., 16, 22, 25
 Miliolina, Subord., 8, 22
 Miliolinae, Subfam., 26, 27
Miliolinella, 27
 Miliolinellinae, Subfam., 26
Mimosina, 44
Masonella, 13
Miliammina, 15
Millettia, 42
Miniacina, 58, 59
Mississippiina, 65
Monalysidium, 29
Montfortella, 57
Morulæpectra, 18, 21
Mucronina, 37
Multifidella, 21

Neoconorbina, 46, 72
Neoeponides, 56
Neouvigerina, 45
Nevillina, 27
Nodobacularia, 25
 Nodobaculariinae, Subfam., 24, 25
Nodophthalmidium, 25
Nodosarella, 60
Nodosaria, 37, 75
 Nodosariacea, Superfam., 30, 35
 Nodosariidae, Fam., 31, 32, 35
 Nodosariinae, Subfam., 35, 38
Nodosinum, 15
Nonion, 62
Nonionella, 62
Nonionellina, 62, 70
 Nonionidae, Fam., 30, 32, 52, 59, 61
 Nonioninae, Subfam., 61, 62
Normanina, 10, 11
Notorotalia, 52
Nouria, 15
 Nouriidae, Fam., 14, 15
Nubecularia, 24, 28
Nubeculariella, 10
 Nubeculariidae, Fam., 22, 23
 Nubeculariinae, Subfam., 24
Nubeculina, 15, 25
Nubeculopsis, 24
Nummoloculina, 25, 27
Nummulites, 52
 Nummulitidae, Fam., 32, 34, 50, 51
 Nummulitinae, Subfam., 52

Oolina, 39
 Oolininae, Subfam., 38, 39
 Ophthalmidiinae, Subfam., 23, 24
Ophthalmidium, 24
Ophthalmina, 24
 Orbitoidacea, Superfam., 30, 48, 51, 54
Orbulina, 51, 54
 Orbulininae, Subfam., 54
Oridorsalis, 63
Orthomorphina, 36
Orthoplecta, 41
Oryctoderma, 13, 72
Osangularia, 63
 Osangulariidae, Fam., 33, 59, 63
Ozawaia, 52

Palmerinella, 48
Palmula, 36
Pandoglandulina, 37
Paradentalina, 37, 38
Parafissurina, 39
Pararotalia, 50, 71
Paramolina, 56, 64
Parrellina, 52
Parrina, 24, 28
Patellina, 49
Patellinella, 47
Patellininae, Fam., 49
Patellinoidea, 49
Pateoris, 26
Paumotuia, 56
Pavonina, 44
Pavonininae, Subfam., 42, 43
Pavonitiniidae, Fam., 14, 22
Pavonitiniinae, Subfam., 22
Pegidia, 51, 74
Pegidiidae, Fam., 51
Pegidiinae, Subfam., 50, 51, 54
Pelosina, 10, 12
Pelosphaera, 12, 67
Peneroplinae, Subfam., 28
Peneroplis, 28, 29, 70
Phlegeria, 37, 39
Pilulina, 11, 12
Physalidia, 47
Placopsilina, 18
Placopsilininae, Subfam., 16, 18
Planctostoma, 19
Planispirillina, 49
Planispirinella, 23, 25
Planispironoides, 25
Planodiscorbis, 46
Planogypsina, 58, 78
Planopulvinulina, 56
Planorbulina, 57
Planorbulinella, 57
Planorbulinidae, Fam., 35, 55, 57
Planorbulinoides, 57
Planularia, 36
Planulina, 56, 78
Planulininae, Subfam., 56
Planulinoides, 47
Plectina, 22
Plectofrondicularia, 36, 37
Plectofrondiculariinae, Subfam., 35, 37
Pleurostomella, 60, 72
Pleurostomellidae, Fam., 31, 59
Polymorphina, 38
Polymorphinidae, Fam., 30, 35, 38
Polymorphininae, Subfam., 38
Polysegmentina, 27
Polystomellina, 52, 76
Poritextularia, 18
Poroeponides, 56
Praeglobobulimina, 43
Protobotellina, 9, 10, 11
Protoschista, 15
Proxifrons, 37
Psammonyx, 13
Psammophax, 11, 12
consociata, 11, 12
Psammosphaera, 11
Psammosphaerinae, Subfam., 11
Pseudobolivina, 19
Pseudoboliviniinae, Subfam., 18, 19
Pseudobulimina, 66
Pseudoeponides, 48
Pseudomassilina, 26
Pseudonodosaria, 37
Pseudopolymorphina, 38
Pseudorotalia, 50
Pseudowebbinella, 13
Prychomiliola, 26, 28

Pullenia, 62
Pulleniatina, 54
Pyrgo, 26, 27, 68
Pyrgoella, 26
Pyropilus, 58
Pyrula, 38

Quadriformina, 62
Quinqueloculina, 26
Quinqueloculininae, Subfam., 26

Radicula, 9
Ramulina, 38
Ramulininae, Subfam., 38
Recticulophragmium, 17
Rectobolivina, 41, 45, 72, 77
Rectouvigerina, 45
Recurvoides, 16, 20, 77
Reissia, 41
Remaneica, 20
Remaneicinae, Subfam., 19, 20
Reophax, 15, 67
Reussella, 44
Rhabdammina, 9
Rhapydionininae, Subfam., 28
Rhizammina, 9
Rhizammininae, Subfam., 9
Rimulina, 37
Ripacubana, 29
Riveroina, 24, 28
Robertina, 66
Robertinacea, Superfam., 30, 65
Robertinidae, Fam., 32, 33, 34, 65, 66
Robertinoides, 66
Robulus, 36
Rosalina, 46, 73
Rotalia, 51
Rotaliacea, Superfam., 30, 49
Rotaliammina, 20
Rotaliella, 49
Rotaliellidae, Fam., 49
Rotaliidae, Fam., 30, 31, 32, 33, 34, 50
Rotaliina, Subord., 8, 30
Rotaliinae, Subfam., 50
Rubratella, 65
Rudigaudryina, 21
Rugidia, 19, 47
Rupertina, 51
Rupertininae, Subfam., 50, 51
Rzehakinidae, Fam., 14, 15, 27

Saccammina, 12
Saccaminidae, Fam., 8, 11
Saccamininae, Subfam., 11
Saccodendron, 10, 12
Saccorhiza, 10, 74
Sagenina, 12
Sagrina, 44
Saracenaria, 36
Schizamminidae, Fam., 8, 11
Schizammina, 9, 10, 11
Schlumbergerella, 51
Schlumbergerina, 27
Scutuloris, 27
Seabrookia, 39
Seabrookiinae, Subfam., 38, 39
Sejunctella, 49
Shepherdella, 8
Sigmavirgulina, 60
Sigmoidella, 38
Sigmoilina, 26

Sigmoilopsis, 27
Sigmomorphina, 38, 71, 76
Siphogenerina, 45
Siphoglobulina, 39
Siphonaperta, 27
Siphonina, 48, 71
Siphoninella, 48
Siphoninidae, Fam., 32, 33, 45, 48
Siphoninoides, 48
Siphonodosaria, 42
Siphotextularia, 19
Siphotrochammina, 20
Siphovigerina, 38, 44, 45,
Sorites, 29, 67
Soritidae, Fam., 22, 28
Soritinae, Subfam., 28, 29
Sorosphaera, 11, 12
Sphaerammina, 17
Sphaerammininae, Subfam., 16
Sphaeridia, 51
Sphaerogypsina, 58
Sphaeroidina, 19, 40
Sphaeroidinella, 40, 54
Sphaeroidinellinae, Subfam., 54
Sphaeroidinidae, Fam., 32, 40
Spirillina, 49, 68
Spirillinacea, Superfam., 30, 49
Spirillinidae, Fam., 30, 31, 32, 49
Spirillininae, Subfam., 49
Spirillinoides, 13
Spirolina, 29
Spirolocammina, 15
Spiroloculina, 24
Spiroloculininae, Subfam., 23, 24
Spiroplectammina, 18
Spiroplectammininae, Subfam., 18
Sporadogenerina, 38
Sporadotrema, 58, 59
Squamulina, 23
Squamulinidae, Fam., 22, 23
Stainforthia, 41, 42
Stetsonia, 47, 68
Stilostomella, 42
Storthosphaera, 11
Subbotina, 54
Sulcophax, 15
Svratkina, 63
Syringamma, 10, 67

Tawitawia, 19
Tawitawiinae, Subfam., 18, 19
Techitella, 11, 12
Textularia, 18, 74
Textulariella, 18, 21
Textulariidae, Fam., 14, 18
Textulariina, Subord., 8
Textulariinae, Subfam., 18
Textularioides, 18
Thalmannammina, 16, 19, 20
Tholosina, 12
Thurammina, 12
Tinophodella, 55, 69

Tiphotrocha, 20, 76
Tolypammina, 14
Tolypammininae, Subfam., 13, 14
Tomaculoides, 37, 39
Torrosina, 48
Tretomphalus, 45, 54
Trichohyalus, 63
Trifarina, 44
Trilocularena, 15
Triloculina, 26
Trimosina, 44
Triplasia, 17
Tritaxia, 21
Tritaxilina, 22
Tritaxis, 20, 21
Trochammina, 19, 20
Trochamminidae, Fam., 14, 19
Trochammininae, Subfam., 19
Trochamminita, 16, 20
Trochamminoides, 16
Trochamminula, 20
Tubinella, 23, 28
Tubinellinae, Subfam., 25, 28
Turborotalia, 53
Turrilinae, Fam., 34, 40
Turrispirillina, 49
Turritella, 13

Ungulatella, 66
Usbekistania, 14
Uvigerina, 45
Uvigerinella, 43, 44
Uvigerinidae, Fam., 34, 40, 44

Vaginulina, 37
Vaginulinopsis, 36
Valvobifarina, 44, 69
Valvulina, 21
Valvulineria, 47
Valvulininae, Subfam., 20, 21
Vanhoeffenella, 9, 12
Verneuilina, 20
Verneuilininae, Subfam., 20
Verrucina, 12, 13
Vertebralina, 25, 29
Victoriellinae, Subfam., 58, 59
Virgulinopsis, 41
Virgulopsis, 42
Vulvulina, 18, 19

Webbina, 24
Webbinella, 38
Webbinellinae, Subfam., 38
Weisnerella, 24

Zeaflorius, 62, 63
Zoyaella, 24

MEMOIRS OF THE NEW ZEALAND OCEANOGRAPHIC INSTITUTE

Year	Title	Memoir No.	Year	Title
1955	Bibliography of New Zealand Oceanography, 1949–1953. By N.Z. OCEANOGRAPHIC COMMITTEE. <i>N.Z. Dep. scient. ind. Res. geophys. Mem. 4.</i>	16	1963	Bibliography of New Zealand Marine Zoology 1769–1899. By DOROTHY FREED. <i>Bull. N.Z. Dep. scient. ind. Res. 148.</i>
1957	General Account of the Chatham Islands 1954 Expedition. By G. A. KNOX. <i>Bull. N.Z. Dep. scient. ind. Res. 122.</i>	17	1965	Studies of a Southern Fiord. By T. M. SKERMAN (Ed.) <i>Bull. N.Z. Dep. scient. ind. Res. 157.</i>
1959	Contributions to Marine Microbiology. Compiled by T. M. SKERMAN. <i>N.Z. Dep. scient. ind. Res. Inf. Ser. 22.</i>	18	1961	The Fauna of the Ross Sea. Part 1. Ophiuroidea. By H. BARRACLOUGH FELL. <i>Bull. N.Z. Dep. scient. ind. Res. 142.</i>
1960	Biological Results of the Chatham Islands 1954 Expedition. Part 1. Decapoda Brachyura, by R. K. DELL; Cumacea, by N. S. JONES; Decapoda Natantia, by J. C. YALDWYN. <i>Bull. N.Z. Dep. scient. ind. Res. 139 (1).</i>	19	1962	The Fauna of the Ross Sea. Part 2. Scleractinian Corals. By DONALD F. SQUIRES. <i>Bull. N.Z. Dep. scient. ind. Res. 147.</i>
1960	Biological Results of the Chatham Islands 1954 Expedition. Part 2. Archibenthal and Littoral Echinoderms. By H. BARRACLOUGH FELL. <i>Bull. N.Z. Dep. scient. ind. Res. 139 (2).</i>	20	1963	<i>Flabellum rubrum</i> (Quoy and Gaimard). By DONALD F. SQUIRES. <i>Bull. N.Z. Dep. scient. ind. Res. 154.</i>
1960	Biological Results of the Chatham Islands 1954 Expedition. Part 3. Polychaeta Errantia. By G. A. KNOX. <i>Bull. N.Z. Dep. scient. ind. Res. 139 (3).</i>	21	1963	The Fauna of the Ross Sea. Part 3. Asteroidea. By HELEN E. SHEARBURN CLARK. <i>Bull. N.Z. Dep. scient. ind. Res. 151.</i>
1960	Biological Results of the Chatham Islands 1954 Expedition. Part 4. Marine Mollusca, by R. K. DELL; Sipunculoidea, by S. J. EDWARDS. <i>Bull. N.Z. Dep. scient. ind. Res. 139 (4).</i>	22	1964	The Marine Fauna of New Zealand: Crustacea Brachyura. By E. W. BENNETT. <i>Bull. N.Z. Dep. scient. ind. Res. 153.</i>
1961	Hydrology of New Zealand Coastal Waters, 1955. By D. M. GARNER. <i>Bull. N.Z. Dep. scient. ind. Res. 138.</i>	23	1963	The Marine Fauna of New Zealand: Crustaceans of the Order Cumacea. By N. S. JONES. <i>Bull. N.Z. Dep. scient. ind. Res. 152.</i>
1962	Analysis of Hydrological Observations in the New Zealand Region, 1874–1955. By D. M. GARNER. <i>Bull. N.Z. Dep. scient. ind. Res. 144.</i>	24	1964	A Bibliography of the Oceanography of the Tasman and Coral Seas, 1860–1960. By BETTY N. KREBS. <i>Bull. N.Z. Dep. scient. ind. Res. 156.</i>
1961	Hydrology of Circumpolar Waters South of New Zealand. By R. W. BURLING. <i>Bull. N.Z. Dep. scient. ind. Res. 143.</i>	25	1965	A Foraminiferal Fauna from the Western Continental Shelf, North Island, New Zealand. By R. H. HEDLEY, C. M. HURDLE, and I. D. J. BURDETT. <i>Bull. N.Z. Dep. scient. ind. Res. 163.</i>
1964	Bathymetry of the New Zealand Region. By J. W. BRODIE. <i>Bull. N.Z. Dep. scient. ind. Res. 161.</i>	26	1964	Sediments of Chatham Rise. By ROBERT M. NORRIS. <i>Bull. N.Z. Dep. scient. ind. Res. 159.</i>
1965	Hydrology of New Zealand Offshore Waters. By D. M. GARNER and N. M. RIDGWAY. <i>Bull. N.Z. Dep. scient. ind. Res. 162.</i>	27	1965	The Fauna of the Ross Sea. Part 4. Mysidacea, by OLIVE S. TATTERSALL. Part 5. Sipunculoidea, by S. J. EDMONDS. <i>Bull. N.Z. Dep. scient. ind. Res. 167.</i>
1961	Biological Results of the Chatham Islands 1954 Expedition. Part 5. Porifera: Demospongiae, by PATRICIA R. BERGQUIST; Porifera: Keratosa, by PATRICIA R. BERGQUIST; Crustacea Isopoda: Bopyridae, by RICHARD B. PIKE; Crustacea Isopoda: Serolidae, by D. E. HURLEY; Hydroida, by PATRICIA M. RALPH. <i>Bull. N.Z. Dep. scient. ind. Res. 139 (5).</i>	28	1966	Sedimentation in Hawke Bay. By H. M. PANTIN. <i>Bull. N.Z. Dep. scient. ind. Res. 171.</i>
1963	Submarine Morphology East of the North Island, New Zealand. By H. M. PANTIN. <i>Bull. N.Z. Dep. scient. ind. Res. 149.</i>	29	1964	Biological Results of the Chatham Islands 1954 Expedition. Part 6. Scleractinia, by D. F. SQUIRES. <i>Bull. N.Z. Dep. scient. ind. Res. 139 (6).</i>
In prep.	Marine Geology of Cook Strait. By J. W. BRODIE. <i>N.Z. Dep. scient. ind. Res.</i>	30	1966	Geology and Geomagnetism of the Bounty Region east of the South Island, New Zealand. By D. C. KRAUSE. <i>Bull. N.Z. Dep. scient. ind. Res. 170.</i>
		31	In prep.	Marine Biology, Hydrology, Physiography of Manihiki Atoll, Cook Islands. Compiled by C. MCCANN and J. S. BULLIVANT. <i>N.Z. Dep. scient. ind. Res.</i>
		32	1967	The Fauna of the Ross Sea. Part 5. General Accounts, Station Lists, and Benthic Ecology. By JOHN S. BULLIVANT and JOHN H. DEARBORN. <i>Bull. N.Z. Dep. scient. ind. Res. 176.</i>

<i>Memoir No.</i>	<i>Year</i>	<i>Title</i>	<i>Memoir No.</i>	<i>Year</i>	<i>Title</i>
33	1967	The Submarine Geology of Foveaux Strait. By D. J. CULLEN. <i>Bull. N.Z. Dep. scient. ind. Res.</i> 184.	39	1967	Hydrology of the Southern Hikurangi Trench Region. By D. M. GARNER. <i>Bull. N.Z. Dep. scient. ind. Res.</i> 177.
34	In prep.	Benthic Ecology of Foveaux Strait. By E. W. DAWSON. <i>Bull. N.Z. Dep. scient. ind. Res.</i>	40	1967	Sediments of the Western Shelf, North Island, New Zealand. By J. C. MCDUGALL and J. W. BRODIE. <i>Bull. N.Z. Dep. scient. ind. Res.</i> 179.
35	1966	The Marine Fauna of New Zealand: Spider Crabs, Family Majidae (Crustacea Brachyura). By D. J. GRIFFIN. <i>Bull. N.Z. Dep. scient. ind. Res.</i> 172.	41	1967	Bathymetric and Geological Structure of the North-western Tasman Sea—Coral Sea—South Solomon area of the Southwestern Pacific Ocean. By DALE C. KRAUSE. <i>Bull. N.Z. Dep. scient. ind. Res.</i> 183.
36	1967	Water Masses and Fronts in the Southern Ocean South of New Zealand. By TH. J. HOUTMAN. <i>Bull. N.Z. Dep. scient. ind. Res.</i> 174.	42	1968	The Echinozoan Fauna of the New Zealand Subantarctic Islands, Macquarie Island and the Chatham Rise. By D. L. PAWSON. <i>Bull. N.Z. Dep. scient. ind. Res.</i> 187.
37	1968	The Marine Fauna of New Zealand: Porifera, Demospongiae. Part I. Tetractinomorpha and Lithistida. By PATRICIA R. BERGQUIST. <i>Bull. N.Z. Dep. scient. ind. Res.</i> 188.	43	1967	The Marine Fauna of New Zealand. Scleractinian Corals. By I. W. SQUIRES and D. F. KEYES. <i>Bull. N.Z. Dep. scient. ind. Res.</i> 185.
38	1967	The Marine Fauna of New Zealand: Intertidal Foraminifera of the <i>Corallina officinalis</i> zone. By R. H. HEDLEY, C. M. HURDLE, and I. D. J. BURDETT. <i>Bull. N.Z. Dep. scient. ind. Res.</i> 180.	44	1967	A Checklist of Recent New Zealand Foraminifera. By J. V. EADE. <i>Bull. N.Z. Dep. scient. ind. Res.</i> 182.

BY AUTHORITY:
A. R. SHEARER, GOVERNMENT PRINTER, WELLINGTON, NEW ZEALAND—1969

1,530/7/68—35367 A



